Relationships Among the Results of Coagulase, Staphylococcal Toxin, and Thermonuclease Tests on Staphylococci from Cow Milk

DONALD E. JASPER,* FIDEL INFANTE, AND JON D. DELLINGER

Department of Clinical Pathology, School of Veterinary Medicine, University of California, Davis, California 95616

Received 25 October 1984/accepted 18 December 1984

Production of staphylococcal alpha- or alpha-beta-toxins correlated well with production of coagulase or thermonuclease (or both) in 203 Staphylococcus aureus isolates from milk and should be reliable indicators of S. aureus in the absence of Staphylococcus intermedius. Failures to produce toxin, tube coagulase, or thermonuclease occurred in only 1 to 2% of S. aureus. Evidence of beta- or alpha-beta-toxins was not found among 321 other staphylococci isolated from milk. A few coagulase- or thermonuclease-positive isolates not producing beta- or alpha-beta-toxins were found among the Staphylococcus hyicus isolates.

Positive tube coagulase tests have long been accepted as justification for classification of staphylococci of both human and animal origin as Staphylococcus aureus. This practice is no longer totally valid in view of recent more definitive methods for classifying staphylococci (11, 20) and the finding of other coagulase-positive species among animal isolates (3, 4, 15, 20, 21). Hajek (6) described a new species, Staphylococcus intermedius, which is coagulase positive and produces staphylococcal beta- and delta-toxins (100%) and alpha-toxins (22%). (Staphylococcal hemolysins are herein referred to as alpha, beta, etc., toxins to avoid confusion with the streptococcal hemolysin.) Little is known of the distribution of S. intermedius in milk samples or of its possible association with mastitis. Five isolates were reported among 93 staphylococcal isolates from intramammary infections from dairy cows and 6 from 60 intramammary infections from beef cows (20). In another study, 525 isolates from intramammary infections of beef cows were classified as 141 S. intermedius, 151 Staphylococcus simulans, 85 S. aureus, and small numbers of 10 other staphylococcal species (21). In that study 133 of 241 (55.2%) coagulase-positive isolates were classified as S. intermedius, 83 (34.4%) were classified as S. aureus, and 15 of 151 (10.0%) were classified as S. simulans, a coagulase-negative organism reported to produce staphylococcal beta-toxin (21). Reliance upon coagulase or hemolytic characteristics (or both) to identify S. aureus was therefore considered to be unsatisfactory in that study because of the large number of S. intermedius isolates. Many of these latter isolates produced a weak maltose reaction, and reclassification as atypical S. aureus is being considered (personal communication). Langlois et al. (14) did not find any S. intermedius among 581 isolates including 10 staphylococcal species from milk of dairy cows. Coagulase-positive staphylococci from cow milk usually produce beta-toxin, alone or in combination with alpha-, delta-, or epsilon-toxins, which become evident on properly prepared bovine erythrocyte agar (BEA) plates (2, 3, 5, 7-10, 15-18). Several have emphasized the reliability of beta-toxin production as an indicator of coagulase-positive staphylococci isolated from milk (5, 7-9, 18). Hajek and Marsalek (7) found that 100% of coagulase-positive strains produced beta-toxin, and DeVriese (3) more recently considered the presence of beta-toxin sufficient evidence to identify S. aureus. It has been a common practice in applied bovine mastitis bacteriology to consider the production of beta-toxin as indirect evidence for coagulase production and therefore for identification of an organism as S. aureus. If S. intermedius or coagulase-positive Staphylococcus hyicus appears with significant frequency in milk samples, or if other species are identified that produce coagulase or beta-toxin, common field laboratory diagnostic practices will need revision. This research was done to investigate the diagnostic significance of staphylococcal hemolytic toxin, tube coagulase, and thermostable nuclease production by staphylococci isolated from milk samples and identified by biochemical means now available (11, 21).

MATERIALS AND METHODS

The staphylococci for this study were obtained from milk samples taken for general surveys (composite samples) or for mastitis diagnosis (quarter samples) from 26 California dairy herds. Milk (0.01 ml) was streaked on BEA plates prepared with washed cow blood cells (2). Colonies of catalase-positive cocci were picked and transferred to a second BEA plate as a test for purity and then transferred to a Trypticase soy agar (BBL Microbiology Systems, Cockeysville, Md.) slant and refrigerated at 4°C pending analysis. Revitalization was assured by making transfers on BEA before further testing was done. Hemolytic toxin activity was determined visually, as recommended previously (2), by an experienced observer after incubation of BEA plates at 37°C for 24 h. The standard tube coagulase test was done as recommended previously (2) with Difco rabbit plasma and was read at 4 and 24 h. Thermonuclease tests were performed on both P agar (19) and Mueller-Hinton (M-H) agar plates by an agar overlay modification (1) of the method of Lachica et al. (13). Final identity of staphylococcal isolates was made biochemically by the methods of Kloos and Schleifer (11) as modified to include species not found in humans (21). Anaerobic mannitol utilization testing was not done. Staphylococcal isolates having at least two of the following characteristics (beta- or alpha-beta-toxin, coagulase positive, thermonuclease positive), and which were maltose positive, were considered to be S. aureus. Other staphylococci were examined by the Kloos and Schleifer simplified scheme (11) as modified previously (21).

* Corresponding author.
RESULTS

By the criteria selected, 203 of the isolates were S. aureus (Table 1). The results of the tests for alpha- or alpha-beta-toxin, tube coagulase, and thermonuclease were all positive for 96.6% of the S. aureus isolates in this study. The tube coagulase and hemolytic toxin tests were each positive for 99% of the isolates tested. The thermonuclease tests were positive for 98% of the isolates, being identical on M-H and P agar plates.

The results of these tests were also very much as expected for the other staphylococci studied (Table 2). However, four isolates identified as S. hyicus had a narrow zone of clear hemolysis, as did one isolate identified as Staphylococcus sciuri. Tube coagulase and thermonuclease test results were negative for 242 S. hyicus isolates, but varied among 13 isolates that were positive to one or more of these tests. Beta- or alpha-beta-toxin production was not observed among any staphylococci other than S. aureus, although one isolate designated S. simulans was nuclease positive.

In one instance only, the thermonuclease test was positive on M-H agar and not on P agar.

DISCUSSION

There was a strong association of tube coagulase (99.5%), beta- or alpha-beta-toxin (99%), and thermonuclease (98%) with S. aureus isolates from milk. All three tests were positive 97% of the time. Toxin was not observed in only two isolates, and thermonuclease was not observed in only three isolates found to be S. aureus. At the moment it appears that a test for maltose fermentation would be sufficient for routine differentiation (12, 21).

Since identification of staphylococcal species from cow milk is still not well established on an extensive data base of its own, it is likely that misidentifications do occur. The thermonuclease-positive S. simulans isolate in this series may in fact be S. hyicus, although it is phosphatase negative, and the four weakly hemolytic S. hyicus isolates may in fact be S. simulans, although they are phosphatase positive. We did not find coagulase-negative S. simulans producing staphylococcal beta-toxin as was previously reported (21).

Observations for production of hemolytic staphylococcal alpha- or alpha-beta-toxin made on good quality BEA plates used for primary inoculation of bovine milk samples provide an easy and reliable presumptive diagnosis for S. aureus from milk in the absence of S. intermedius. It is important, however, that properly prepared plates be used (2), and that good lighting is available for reading by a competent observer.

ACKNOWLEDGMENT

This work was supported in part by funds provided by the U.S. Department of Agriculture under the Animal Health Act of 1977 PL95-113.

LITERATURE CITED


### Table 1. Staphylococcal toxin, tube coagulase, and thermonuclease test results on S. aureus isolates from milk

<table>
<thead>
<tr>
<th>No. of isolates</th>
<th>Alpha- or alpha-beta-toxin</th>
<th>Hemolytic toxin</th>
<th>Tube coagulase</th>
<th>Thermonuclease</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>P agar M-H agar</td>
</tr>
<tr>
<td>197</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>1</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

### Table 2. Staphylococcal toxin, tube coagulase, and thermonuclease tests results on 321 other staphylococci isolated from milk

<table>
<thead>
<tr>
<th>Species</th>
<th>No. of isolates</th>
<th>Hemolytic toxin</th>
<th>Tube coagulase</th>
<th>Thermonuclease</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staphylococcus capitis</td>
<td>1</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Staphylococcus cohnii</td>
<td>1</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Staphylococcus epidermidis</td>
<td>7</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Staphylococcus hyicus</td>
<td>238</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Staphylococcus sciuri</td>
<td>4 N</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Staphylococcus simulans</td>
<td>5</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Staphylococcus xylosis</td>
<td>14</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Unknown</td>
<td>17</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Micrococcus spp.</td>
<td>3</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

* N, Narrow zone of weak hemolysis.
584 JASPER, INFANTE, AND DELLINGER


ERRATA

Early Detection of Streptococci in Swabs by Latex Agglutination Before Culture

D. N. PETTS

Microbiology Department, Basildon Hospital, Nethermayne, Basildon, Essex, SS16 5NL England

Volume 19, no. 3, p. 433, column 1, Table 1: Footnote b is missing. It should read “These specimens all contained K. pneumoniae type 54, and the positive results were due to a cross-reaction.”

Relationships Among the Results of Coagulase, Staphylococcal Toxin, and Thermonuclease Tests on Staphylococci from Cow Milk

DONALD E. JASPER, FIDEL INFANTE, AND JON D. DELLINGER

Department of Clinical Pathology, School of Veterinary Medicine, University of California, Davis, California 95616

Volume 21, no. 4, p. 582, abstract, line 1: “alpha- or alpha-beta toxins” should read “beta- or alpha-beta toxins.”

Page 583, column 1, lines 2–3 and 48 and Table 1, heading 2: “alpha- or alpha-beta toxin” should read “beta- or alpha-beta toxin.”

Identification of 22 Legionella Species and 33 Serogroups with the Slide Agglutination Test

W. LANIER THACKER, BONNIE B. PLIKAYTIS, AND HAZEL W. WILKINSON

Division of Bacterial Diseases, Center for Infectious Diseases, Centers for Disease Control, Atlanta, Georgia 30333

Volume 21, no. 5, p. 781, column 2, lines 43–45: “... and more recently, L. hackeliae, L. rubrilucens, L. maceachernii, and L. parisiensis” should read “... and more recently, L. hackeliae and L. maceachernii.”

Accuracy and Reproducibility of a Four-Hour Method for Anaerobe Identification

PETER C. APPELBAUM, CINDY S. KAUFMANN, AND JOSEPH W. DEPENBUSCH

Department of Pathology (Clinical Microbiology), Hershey Medical Center, Hershey, Pennsylvania 17033, and Department of Laboratory Medicine (Clinical Microbiology), The Johns Hopkins Hospital, Baltimore, Maryland 21205

Volume 21, no. 6, p. 894, column 2, lines 18–20: “Isolates were maintained anaerobically at room temperature in chopped-meat-glucose medium” should read “Isolates were maintained in a frozen state at −70°C before testing.”