Isolation of a Bovine Rotavirus with a ‘‘Super-Short’’ RNA Electrophoretic Pattern from a Calf with Diarrhea

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A rotavirus with a ‘‘super-short’’ RNA electrophoretotype was isolated from a calf with diarrhea and was designated VMRI strain. Segments 10 and 11 of this rotavirus migrated more slowly than did those of bovine rotavirus strains NCDV, B641, and B223. The electrophoretic pattern of the VMRI strain was similar to that reported for rotaviruses with super-short RNA electropherotypes from humans and rabbits. Northern (RNA) blot hybridization indicated that gene 11 of the VMRI strain was altered and migrated between gene segments 9 and 10. The subgroup of the VMRI strain was shown to be subgroup I. The VMRI strain of bovine rotavirus was neutralized by antisera containing polyclonal antibodies to rotavirus serotype 6 (bovine rotavirus serotype I) strains NCDV and B641 and by ascitic fluid containing monoclonal antibodies directed to VP7 of serotype 6 rotavirus. The VMRI strain was not neutralized by either polyclonal or monoclonal antibodies to strain B223 (bovine rotavirus serotype II). Collective data on the neutralization of the VMRI strain with monoclonal antibodies and polyclonal antibodies suggest that this virus is a member of the NCDV group (serotype 6) of rotaviruses (bovine rotavirus serotype I).

Rotaviruses are major causes of enteric disease in animals and humans (4, 7, 8, 10, 11, 27, 37). Although there are several antigenically and electrophoretically distinct groups (A through E) of rotaviruses (2, 3, 31–33), group A rotaviruses appear to be the most prevalent. Group A rotaviruses share a common group antigen on protein VP6 (38). Two subgroups of group A rotaviruses have been reported (15). Neutralizing antigens are contained in two proteins, VP3 and VP7 (16), which also provide serotypic specificity (16). Hoshino et al. (19) reported seven group A rotavirus serotypes based on a minimum of 20-fold differences between the neutralizing titers of antisera with homologous and heterologous viruses. Only serotype 6 (bovine rotavirus serotype I) with an NCDV strain serving as a prototype has been detected in cattle (27). Strains UK, B-14, and B641 belong to this serotype (19, 37, 39, 40). Additional serotypes of bovine rotavirus have been detected (5, 6, 20, 28, 30, 34, 40). One of these serotypes (bovine rotavirus serotype II) has been detected in U.S. cattle, with strain B223 serving as a prototype virus (40). There may also be additional serotypes of rotavirus in U.S. cattle (39).

Group A rotaviruses have a characteristic RNA electrophoretic pattern in polyacrylamide gels in which 11 segments of double-stranded (ds) RNA are arranged in four regions in groups of 4, 2, 3, and 2 segments (22). The group A rotavirus common and subgroup antigens are coded for in gene segment 6 (10, 14). The neutralization antigens in proteins VP3 and VP7 are encoded by gene segments 4 and either 8 or 9, respectively (21, 29). All of the bovine rotavirus designs thus far reported have long RNA electropherotypes. However, rotaviruses with ‘‘super-short’’ RNA electropherotypes have recently been reported for humans and rabbits (1, 17, 25, 36). In these viruses, gene segments 10 and 11 have slower electrophoretic mobilities (1). Human rotavirus with a super-short RNA pattern represents a new serotype (1, 25), whereas the rabbit rotavirus was shown to be a serotype 3 rotavirus (36). In this report, we describe isolation and preliminary characterization of a bovine rotavirus from a calf with diarrhea, designated VMRI strain, with a super-short RNA electrophoretic pattern.

MATERIALS AND METHODS

Case history. A dead, two-week-old Simmental calf from Missouri was submitted to the Iowa State University Veterinary Medical Diagnostic Laboratory. The calf developed diarrhea at 2 days of age and was treated with gentamicin, penicillin, and oral electrolytes. The calf responded but relapsed soon after therapy was discontinued. It was then treated intermittently for 13 days before it died at 15 days of age. Seven other calves died out of a total of ninety. Most of the calves went through episodes of diarrhea, but samples from these animals were not available for investigation.

Diagnostic tests for viral antigens. Intestinal contents were tested for rotavirus common antigens by an enzyme-linked immunosorbent assay (Rotazyme II; Abbott Laboratories, North Chicago, Ill.) according to the protocol of the manufacturer.

RNA analysis. The electrophoretic pattern of genomic viral RNA was determined by polyacrylamide gel electrophoresis. Viral RNA was extracted from fecal samples by a procedure similar to that described by Herring et al. (18), except that all of the steps were performed by using a microcentrifuge. Briefly, 0.2-g fecal samples were diluted with 0.6 ml of sodium acetate buffer, sonicated, and clarified by centrifugation at 13,000 × g for 1 min. The supernatants were harvested, extracted with 0.5 ml of phenol-chloroform solution, and clarified by centrifugation at 13,000 × g in a
microcentrifuge for 1 min. Electrophoresis of RNA segments was performed in Laemmlli 0.75-mm-thick polyacrylamide slab gels (23) with 10% resolving and 3.5% stacking gels. Electrophoresis was done at a constant current of 9 mA for 16 h at 4°C, and gels were stained with silver (18). Cells infected with rotavirus were extracted with Freon (E. I. du Pont de Nemours & Co., Wilmington, Del.) and pelleted through 30% sucrose at 100,000 × g for 3 h, and viral RNA was extracted with phenol-chloroform. Electropherotypes of viral RNA were analyzed by polyacrylamide gel electrophoresis.

Viral isolation. Rotavirus was isolated from intestinal contents in MA-104 cells in roller tube cultures as described previously (12). Briefly, 10% suspensions of intestinal contents were prepared in modified Eagle medium containing gentamicin (50 μg/ml), sonicated, clarified by centrifugation at 400 × g for 10 min, and filtered through 0.45-μm filters. The inoculum was pretreated with 10 μg of trypsin (type IX; Sigma Chemical Co., St. Louis, Mo.) per ml for 1 h at 37°C. The inoculum was adsorbed onto cultures of MA-104 cells in roller tubes. After adsorption for 1 h, the monolayers were washed twice with modified Eagle medium and the culture medium was replaced with modified Eagle medium supplemented with trypsin (0.5 μg/ml) and gentamicin (50 μg/ml) without serum. Mock-infected cells were treated similarly, except that modified Eagle medium containing trypsin was substituted for viral inoculum. The cultures were examined for cytopathic effect daily. Once cytopathic effect was extensive, cultures were frozen and thawed twice and passed once more in roller tube cultures. A three-passage virus stock was prepared and used for characterization. Bovine rotavirus strains NCDV, B641, and B223 were propagated in MA-104 or BSC-1 cells.

Antisera and monoclonal antibodies. Bovine rotaviruses were propagated in MA-104 cells (VMRI strain) or BSC-1 cells (B641, NCDV, and B223). Virions were purified on cesium chloride gradients (26), and double-capsid virions were used to prepare hyperimmune serum in guinea pigs against strains VMRI and NCDV. Production of antisera to B223 (GC47), B641 (GC51), and NCDV (GC54) in gnotobiotic calves and antisera to B14, canine serotype 3 virus, and OSU-1 (a porcine virus) in guinea pigs has been described previously (12, 13, 38, 39). Monoclonal antibody IC3 was directed to VP7 of NCDV and monoclonal antibody HRB223 had specificity for VP3 of B223 virus.

Virus neutralization. Neutralization was performed by a fluorescent focus neutralization test as described previously (40). Briefly, samples of rotavirus containing about 200 fluorescent-focus-forming units were mixed with equal volumes of culture medium or with dilutions of antisera and incubated at 37°C for 1 h. The virus-serum mixture (0.1 ml) was inoculated into each of four wells of cells on 96-well microdilution plates. At 24 h after infection, the cells were fixed and stained for rotavirus antigen by an indirect immunofluorescence test. The neutralization titer was the reciprocal of the highest dilution of serum that neutralized 50% or more of the virus. Results are reported as the mean neutralizing titer of the four determinations.

Northern blot hybridization. Approximately 10 μg of dsRNA extracted from VMRI and NCDV bovine rotavirus particles purified by cesium chloride density gradients (26). The dsRNA segments of the viral genome were separated by polyacrylamide gel electrophoresis as described by Laemmlli (23). The dsRNA was denatured partially and transferred electrophoretically to nylon membranes (Bio-Rad Laboratories, Richmond, Calif.) according to the instructions of the manufacturer. Northern (RNA) blot hybridizations were done by using 32P-labeled cDNA probes synthesized from templates of all 11 genes of genomic RNAs or from the single gene 10 or gene 11 of NCDV. The probes were synthesized from RNA with reverse transcriptase by using random primers (Pharmacia, Inc., Piscataway, N.J.) as described by Eiden et al. (8). Hybridization conditions were adapted from Dimitrov et al. (7). After overnight hybridization at 65°C, the membranes were washed as specified by Dimitrov et al. (7), except that the final wash was at 65°C with 1 × SSC (0.15 M NaCl plus 0.015 M sodium citrate). The filters were then exposed to XAR film (Eastman Kodak Co., Rochester, N.Y.).

Detection of subgroup antigen. Rotavirus group A subgroup antigens were determined by a modification of an enzyme-linked immunosorbent assay described by Berg et al. (15). Briefly, wells of microdilution plates (Immunon-I; Dynatech, Chantilly, Va.) were coated with 1:1,000 dilutions each of rabbit anti-OSU-1 and anti-Gottfried porcine rotavirus antiserum in carbonate-bicarbonate buffer. After overnight incubation at 4°C, the plates were washed, and the free sites were blocked with 1% bovine serum albumin in phosphate buffer and 0.1% Tween 20. Rotavirus diluted 1:2 in phosphate buffer-Tween 20-bovine serum albumin was added to wells and incubated for 16 h at 4°C. The wells were washed, and monoclonal antibodies (provided by Y. Hoshino, National Institutes of Health, Bethesda, Md.) reactive with group A rotavirus, subgroup I (255/60/125/14) or subgroup II (631/9/104/56), were added. After 2 h of incubation, the wells were washed and horseradish peroxidase-conjugated goat anti-mouse immunoglobulin G was added. The reactions were developed by adding 0.2 ml of a substrate-chromogen solution consisting of 2 mM hydrogen peroxide and 0.2 mM ABTS [2,2′-azino-di(3-ethyl-benz-thiazoline-6-sulfonic acid)] in 0.5 M citrate buffer, pH 4.0, to each well. The A405 was recorded after 30 min in an enzyme-linked immunosorbent assay reader (Biotech Instruments Inc., Winooski, Vt.). Results were expressed as optical density as well as subgroup ratio. The subgroup ratio was determined by dividing the optical density of the subgroup I monoclonal antibody by that of the subgroup II monoclonal antibody. A subgroup ratio of more than 3.0 was taken as rotavirus belonging to subgroup I, whereas OD ratios of less than 0.5 were indicative of rotavirus subgroup II. Porcine OSU-1 (subgroup I) and Gottfried (subgroup II) strains were used as controls.

RESULTS AND DISCUSSION

Intestinal contents were positive for rotavirus by enzyme-linked immunosorbent assay and RNA electropherotyping. The VMRI strain of rotavirus had a 4,2,3,2 electrophoretic pattern typical of group A rotaviruses, except that in comparison with the NCDV, B641, and B223 strains of bovine rotavirus (Fig. 1), it had a super-short RNA electropherotype. Gene segments 10 and 11 of the VMRI strain migrated more slowly than did those of the NCDV, B641, and B223 strains, and the electropherotype of the cultivated virus was identical to that of the fecal virus. Minor differences in the migration of other segments were also observed among the NCDV, B641, B223, and VMRI strains (Fig. 1). Slow electrophoretic migration of gene segments 10 and 11 was similar to that described for super-short rotaviruses from humans and rabbits (1, 36).

There are two possible explanations for the slower electrophoretic migration of segments 10 and 11 of the VMRI
strain of rotavirus. Either both genes 10 and 11 are altered or only gene 11 is altered. Close examination of the electrophoretic patterns suggested that only gene 11 of VMRI is altered, since it is similar in size to gene 10 of the NCDV, B641, and B223 strains. Additionally, in other rotaviruses with a short pattern, such as the DS-1 strain of human rotavirus, gene 11 is inverted in relation to gene 10 (for a review, see reference 10). To determine if gene 11 or both genes 10 and 11 were altered in the VMRI strain, Northern blots of viral RNA were hybridized with gene 10 or 11 probes. NCDV gene 10 probe hybridized to segment 11 of the VMRI strain (Fig. 2) and segment 10 of the NCDV strain (data not shown). NCDV gene 11 probe hybridized to segment 10 of the VMRI strain (Fig. 2) and segment 11 of the NCDV strain (data not shown), which indicated that gene 11 was altered in the VMRI strain and was longer and migrated more slowly than gene 11 in NCDV. Gene 11 of VMRI migrated between genes 9 and 10. The biological significance of this alteration is unclear.

The VMRI strain contained the rotavirus group A subgroup I antigen but not the subgroup II antigen (Table 1). Control viruses OSU-1 (subgroup I virus) and Gottfried (subgroup II virus) were shown to contain subgroup I and II antigens, respectively (Table 1).

The VMRI strain of rotavirus was serotyped by a fluorescent focus neutralization test with antisera to the B641, NCDV, and B-14 strains, as well as to the VMRI strain (Table 2). Antiserum to serotype 3 canine rotavirus neutralized the VMRI strain to low titers (Table 2). Antiserum to bovine rotavirus B223 and serotype 5 OSU-1 rotavirus did not neutralize VMRI. Anti-VMRI serum neutralized bovine rotaviruses NCDV and B641 but not B223; however, some differences were noted in the neutralization titers with the homologous and heterologous viruses. These data suggested that the VMRI strain is a member of the NCDV/B641 group of serotype 6 rotaviruses. To further study the relationship between the VMRI and NCDV strains, monoclonal antibi-
TABLE 2. Neutralization titers of antisera and monoclonal antibodies to various rotaviruses

<table>
<thead>
<tr>
<th>Virus</th>
<th>Serotype</th>
<th>GP17 (canine)</th>
<th>GP41 (OSU-1)</th>
<th>GP38</th>
<th>GC51</th>
<th>GC54</th>
<th>GP2</th>
<th>GP47 (B223)</th>
<th>GP3</th>
<th>GP4</th>
<th>IC3 (NCDV)</th>
<th>HRB223 (B223)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Canine</td>
<td>3</td>
<td>256,000 &lt;100</td>
<td>&lt;100</td>
<td>ND‡</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>OSU-1</td>
<td>5</td>
<td>&lt;500 12,800</td>
<td>&lt;100</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>B641</td>
<td>(bovine I)</td>
<td>1,000 &lt;100</td>
<td>6,400 640</td>
<td>1,280</td>
<td>20</td>
<td>160</td>
<td>1,280</td>
<td>320</td>
<td>2,560</td>
<td>2,560</td>
<td>&lt;100 10,240</td>
<td>&lt;100 10,240</td>
</tr>
<tr>
<td>NCDV</td>
<td>6 (bovine I)</td>
<td>2,000 &lt;100</td>
<td>12,800 640</td>
<td>5,120</td>
<td>640</td>
<td>320</td>
<td>2,560</td>
<td>&lt;100 10,240</td>
<td>&lt;100 10,240</td>
<td>2,560</td>
<td>&lt;100 10,240</td>
<td>&lt;100 10,240</td>
</tr>
<tr>
<td>B223</td>
<td>Bovine II</td>
<td>500 &lt;100</td>
<td>&lt;100</td>
<td>&lt;20</td>
<td>&lt;20</td>
<td>&lt;10</td>
<td>&lt;10</td>
<td>&lt;100 10,240</td>
<td>&lt;100 10,240</td>
<td>2,560</td>
<td>&lt;100 10,240</td>
<td>&lt;100 10,240</td>
</tr>
<tr>
<td>VMRI</td>
<td>Bovine I</td>
<td>500 &lt;100</td>
<td>6,400 640</td>
<td>1,280</td>
<td>20</td>
<td>160</td>
<td>1,280</td>
<td>320</td>
<td>2,560</td>
<td>2,560</td>
<td>&lt;100 10,240</td>
<td>&lt;100 10,240</td>
</tr>
</tbody>
</table>

* Neutralization titers of antisera are the reciprocal of the highest dilution of antisera which neutralized 50% or more of the virus (200 fluorescent-focus-forming units) in a fluorescent focus neutralization assay. Results represent the mean of four determinations. Numbers in boldface type are homologous titers.

‡ ND, Not done.

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LITERATURE CITED