Detection of Hydatid Antigen in Urine by Countercurrent Immunoelectrophoresis

S. C. PARIJA,1* P. T. RAVINDER,1† AND K. S. V. K. SUBBA RAO2

Departments of Microbiology1 and Cardio-Thoracic Surgery,2 Jawaharlal Institute of Postgraduate Medical Education and Research, Pondicherry, India

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Hydatid antigen was demonstrated for the first time in the urine of patients with hydatid disease by countercurrent immuno­electrophoresis (CIEP). The antigen was detected in the concentrated urine of 7 of 16 (43.75% positive) patients with surgically confirmed hydatid disease, 4 of 10 (40% positive) patients with ultrasound-proven hydatid disease (daughter cysts or prominent septation and hydatid sands demonstrated by ultrasound), and 8 of 14 (57.14% positive) patients with clinically diagnosed (presumptive) hydatid disease. No antigen was detected in the concentrated urine from 24 patients with parasitic diseases other than hydatid disease. However, antigen was detected in 2 (8% false positive) of 25 concentrated urine samples collected from healthy control subjects (blood donors and students). These result suggest that the detection of hydatid antigen in the urine by CIEP is a simple, rapid, and noninvasive method of diagnosis of hydatid disease.

The development of diagnostic techniques for hydatid disease that can be used under field conditions is a priority (11). The diagnostic methods in hydatid disease essentially include the immunodiagnostic methods which demonstrate either circulating hydatid antibodies or antigens in the serum. However, collection of the blood for serum is an invasive procedure requiring technical expertise and disposable syringes to prevent the risk of transmission of serious infectious agents such as human immunodeficiency virus and hepatitis B virus. Noninvasive methods would therefore be of immense value in the diagnosis of cases of hydatid disease and in epidemiological studies of hydatid disease. Therefore, emphasis has recently been placed on the demonstration of microbial antigens in various body fluids other than serum such as saliva, sputum, and urine (1, 6). Demonstration of antigen in the urine is suggested to be of much value, because the collection of urine is a safe and noninvasive procedure and the urine can be collected easily and frequently without causing any inconvenience to the patient.

Excreted microbial antigens have been demonstrated in the urine of patients with a variety of parasitic infections such as leishmaniasis (10), Chagas’ disease (7), filariasis (16), and malaria (9). In patients with hydatid disease, the circulating hydatid antigens have been demonstrated in the serum (9). It is therefore believed that the same hydatid antigen excreted in the urine could be detected. Until now, however, no such reports have been available on the detection of hydatid antigen in urine.

Here we report for the first time the development and evaluation of a countercurrent immunoelectrophoresis (CIEP) test for the detection of excreted hydatid antigen in urine. CIEP is widely used as a simple and rapid immunossay for the detection of antibodies as well as antigens in the serum of patients with a variety of microbial infections including hydatid disease (14).

In the present study, CIEP is based on the principle that wells containing hyperimmune hydatid antisera are placed on the anodic side of the electrophoretic chamber and the urine to be tested for hydatid antigen is placed on the cathodic side. If the urine contains hydatid antigen, during the process of electrophoresis in an electric field, the hydatid antigen and antibodies are driven toward each other and form a line of precipitation. A positive reaction is identified by the lines of precipitation between hyperimmune serum and hydatid antigen in the urine.

MATERIALS AND METHODS

Urine. Urine specimens were collected from patients attending the Jawaharlal Institute of Postgraduate Medical Education and Research Hospital, Pondicherry, India. These included 16 specimens from patients with surgically proven hydatid disease and 10 specimens from patients with ultrasound-proven hydatid disease. The cysts which showed daughter cysts or prominent septation and pathognomonic hydatid sands in cysts by ultrasound were diagnosed as ultrasound-proven cases of hydatid disease. This study also included 14 specimens from patients with clinically diagnosed (presumptive) hydatid disease. In the latter group, the patients presented with clinical signs and symptoms of hydatid disease, but were not operated on; hence the diagnosis could not be proved by surgery. In addition to these patients, urine specimens were collected from 49 normal control subjects (24 from patients with parasitic infections and miscellaneous conditions and 25 from healthy controls, which consisted of blood donors and student volunteers who had not suffered from hydatid disease in the recent past).

A total of 5 ml of urine was collected from each patient in sterile glass vials by aseptic technique, and specimens were labelled and stored at ~20°C until use. Concentration of urine. The urine was concentrated by the method of ethanol precipitation described by Dokseland and Berdal (6). One milliliter of urine was mixed with 0.1 ml of a cold solution of 3 M sodium acetate, 0.1% bovine serum albumin, and 0.1% (wt/vol) sodium azide. To this solution, 1.5 ml of 95% (vol/vol) ethanol stored at 20°C was added. The mixture was blended in a vortex mixture and was centrifuged at 10,000 × g for 10 min at 4°C. The supernatant was discarded, and the concentrated pellet of urine was resuspended in 0.1 ml of phosphate-buffered saline (pH 7.2).

Both normal (unconcentrated) and concentrated urine specimens from each patient were tested in parallel for hydatid antigen by CIEP.

Hyperimmune antisera. Sterile human hydatid cyst fluid was emulsified with an equal volume of Freund’s complete adjuvant. Adult rabbits (weight, 3 to 4 kg) were immunized by intramuscular inoculation of 0.5 ml of emulsion into each of the four limbs. Ten days after the final inoculation, the rabbits were bled and the blood samples were monitored for antibodies to human hydatid cyst fluid by an indirect hemagglutination (IHA) test (12). The IHA test antibody titer was 1 in 1,024.

Hyperimmune antisera containing hydatid antibodies was purified by the method described by Gottstein (8). Briefly, 1 ml of cold serum was mixed with 1
ml of cold saline at pH 7. The serum-saline mixture (2 ml) was added to 2 ml of 50% cold saturated ammonium sulfate (pH 7) dropwise while stirring for 30 min on ice and was then centrifuged (3,000 rpm at 0°C) in a cold centrifuge. The supernatant was discarded, the precipitate was suspended in 2 ml of saline, and the procedure was repeated until the supernatant was colorless. The final precipitate was suspended in 1 ml of saline and was dialyzed against phosphate-buffered saline (pH 7.2) to remove all the residual ammonium sulfate. The titer of the purified antigen serum was 1 in 2,048 by the IHA test.

CIEP. CIEP was performed by the method described by Shariff and Parija (14). Briefly, a gel was made on a glass slide by using 1% Difco Bacto Agar. It was allowed to set at room temperature and thereafter was stored at 4°C overnight before use.

CIEP was performed on the gel slides. Parallel rows of wells (4 mm in diameter and 3 mm apart) were punched out on the slides with the aid of a template. Six pairs of wells were punched in each slide. Wells containing hyperimmune antiserum (titer, 1 in 2,048) were placed on the anodic side of the electrophoretic chamber, and those containing the urine to be tested were placed on the cathodic side. Twenty microliters of urine was put in the well for detection of hydatid antigen, and then a current of 8 V/cm was applied for 30 min. The slides were read unstained and also after staining with amido black.

A positive reaction was defined by a precipitation line between the side with hyperimmune serum and the side with hyperimmune serum. Positive and negative controls were included with each test.

Washing and staining of slides were carried out by immersing the slides for 24 h in Veronal buffer. The slides were then washed for 15 min in a 1% solution of amido black made in a solvent containing 1 ml of glacial acetic acid, 49 ml of distilled water, and 50 ml of methylated spirit for 30 min. Excess stain was washed with the solvent to give a preparation with dark bluish precipitation bands on a clear background. The slide was finally soaked in 1% acetic acid containing 1% aldehyde-stabilized human O cells sensitized with the optimum sensitizing dose of hydatid antigen (12). Hydatid antigen in the sera was detected by CIEP by the method that we described earlier (14). The test was performed to detect hydatid antigen in the serum in the same way as described above for the urine antigen, except that serum instead of urine was used in the cathode wells.

RESULTS

In a positive CIEP test, the presence of hydatid antigen in urine is detected as a line of precipitation between two wells, with one containing the test urine and the other containing raised hyperimmune antiserum rich in hydatid antibodies. In a negative test, the lines of precipitation were absent for urine negative for the hydatid antigen.

Hydatid antigens in the urine were detected in the unconcentrated urine of four (25%) patients with surgically confirmed hydatid disease, two (20%) patients with ultrasound-proven hydatid disease, and three (21.42%) patients with clinically diagnosed cases of hydatid disease. The antigens were also detected in the unconcentrated urine of two (8%) patients with parasitic diseases but were absent from the urine of healthy controls.

After concentration of the urine by ethanol precipitation, the antigen was detected in the concentrated urine of seven (43.75%) patients with surgically confirmed hydatid disease, four (40%) patients with ultrasound-proven hydatid disease, and eight (57.14%) patients with clinically diagnosed cases of hydatid disease. The antigens were detected in the urine of two (8%) healthy controls. Antigens were not detected in the urine of any of the control patients with parasitic diseases.

Tables 1 and 2 present a comparison of the sensitivity and specificity of the IHA test for demonstration of antibodies in serum and CIEP for the detection of antigen in the serum and the detection of antigen in both unconcentrated and concentrated urine for the diagnosis of hydatid disease. Table 3 presents a comparison of the sensitivity of antigen detection in serum and urine of the different patients with hydatid disease and the controls by CIEP.

DISCUSSION

CIEP is a simple, inexpensive, and rapid test. In this laboratory, CIEP was standardized and evaluated earlier to demonstrate circulating hydatid antigen in serum for the diagnosis of hydatid disease (14).

The present study shows for the first time the detection of hydatid antigen in urine by CIEP. In our study, CIEP detected antigen in the unconcentrated urine of four (25%) patients with surgically confirmed hydatid disease, two (20%) patients with ultrasound-proven hydatid disease and three (21.42%) patients with clinically diagnosed hydatid disease. The sensitivity of CIEP for the detection of hydatid antigen in unconcentrated urine was very low compared to that of detection of antigen in the serum (Tables 1 and 2). The lower sensitivity of CIEP for the detection of antigen in urine in comparison to that in serum is possibly due to the lower quantity of hydatid antigen excreted in a large volume of urine. In several studies carried out on the detection of antigen in the urine for diagnosis of leishmaniasis, Chagas’ disease, malaria, filariasis, etc., (7, 9, 10, 16) the urine samples have been concentrated with the aim of concentrating and detecting antigen in an increased number of urine specimens. Therefore, in the present study, an

### TABLE 1. Evaluation of immunoassays for the diagnosis of hydatid disease

<table>
<thead>
<tr>
<th>Subject group</th>
<th>No. of subjects</th>
<th>IHA test</th>
<th>CIEP</th>
</tr>
</thead>
<tbody>
<tr>
<td>Healthy controls</td>
<td>25 0 (0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Controls with other parasitic diseases</td>
<td>24 3 (12.5)</td>
<td>0 (0)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Presumptive hydatid disease</td>
<td>14 (71.42)</td>
<td>6 (42.85)</td>
<td>3 (21.42)</td>
</tr>
<tr>
<td>Surgically confirmed hydatid disease</td>
<td>16 11 (68.75)</td>
<td>8 (50)</td>
<td>4 (25)</td>
</tr>
<tr>
<td>Ultrasound-proven hydatid disease</td>
<td>10 7 (70)</td>
<td>4 (40)</td>
<td>2 (20)</td>
</tr>
</tbody>
</table>

### TABLE 2. Sensitivity, specificity, and positive predictive value of immunoassays for the diagnosis of hydatid disease

<table>
<thead>
<tr>
<th>Test</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>Positive predictive value</th>
</tr>
</thead>
<tbody>
<tr>
<td>IHA test</td>
<td>Serum antibody</td>
<td>70</td>
<td>93.87</td>
</tr>
<tr>
<td>CIEP test</td>
<td>Serum antigen</td>
<td>45</td>
<td>100</td>
</tr>
<tr>
<td>Unconcentrated urine</td>
<td>22.5</td>
<td>95.91</td>
<td>81.81</td>
</tr>
<tr>
<td>Concentrated urine</td>
<td>47.5</td>
<td>95.91</td>
<td>90.47</td>
</tr>
</tbody>
</table>
The failure of CIEP to detect antigen in urine, which can be more safely obtained than blood, use in clinical laboratories because detection of hydatid antigen offers a noninvasive, easy, and safe alternative means of diagnosis of hydatid disease not only under field conditions but also in routine clinical laboratories. It has the potential for wider use in clinical laboratories because detection of hydatid antigen in urine, which can be more safely obtained than blood, would be immensely useful for establishing the etiological diagnosis of hydatid disease.

**REFERENCES**


