Human Immunodeficiency Virus Antibody Testing by Enzyme-Linked Fluorescent and Western Blot Assays Using Serum, Gingival-Crevicular Transudate, and Urine Samples

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The aim of the present study was to evaluate the possible utilization of saliva and urine as alternative samples to serum for the diagnosis of human immunodeficiency virus (HIV) infection. A total of 302 individuals participated in the study: 187 HIV-infected individuals (106 had Centers for Disease Control and Prevention [CDC] stage II infection, 19 had CDC stage III infection, and 62 had CDC stage IV infection) and 115 noninfected persons (46 of the noninfected persons were blood donors and 69 belonged to a group at high risk of HIV infection). Paired saliva and urine samples were taken from each of the participants in the study. The presence of HIV-specific antibodies was detected by an enzyme-linked fluorescent assay (ELFA), and the result was confirmed by Western blot analysis (WB). The ELFA with saliva gave maximum sensitivity and specificity values, while ELFA with blood samples was confirmed to be positive by WB. Among the four reactivity patterns found by WB of these alternative samples, the most frequent included bands against three groups of HIV structural proteins (was ENV, POL, and GAG). The reactivity bands most frequently observed were those for the proteins gp160 and gp120. The least common reactivity band was the band for protein p17. The detection of HIV antibodies in saliva samples by means of ELFA with the possibility of later confirmation by WB makes saliva an alternative to serum for possible use in the diagnosis of infection. In contrast, HIV antibody detection in urine samples by the same methodology (ELFA) could be taken into consideration for use in epidemiological studies.

Over the past few years, deep interest has been shown in the possible use of samples other than serum for the diagnosis of infection or screening for various infectious processes (1, 4–8, 33, 37, 45). Among these, infection caused by the human immunodeficiency virus (HIV) has been the most evaluated (38). Traditionally, tests for the detection of HIV infection have been based on the detection of HIV antibodies in serum or plasma derived from whole blood collected by venipuncture and hence require trained health care workers for collection and laboratory facilities for testing. Among these alternative biological fluids, saliva has already been considered an option in numerous studies (11, 17, 24, 25, 27, 34, 40, 41, 44); in fact, the U.S. Food and Drug Administration has authorized the use of an oral device for the collection of saliva for HIV antibody screening (42).

Whole saliva is a mixture consisting of the secretions of the salivary glands together with oral cavity microorganisms, cells, and a gingival-crevicular transudate (GCT). GCT is a fluid into which immunoglobulins and other blood components passing through the mucosa to the oral cavity are passively excreted. Mortimer and Parry (32) have already shown that the GCT of HIV-infected individuals contains high concentrations of HIV-specific immunoglobulin G (IgG) antibodies. This antibody concentration is certainly lower than that found in serum but is quite sufficient to render GCT an adequate sample for anti-HIV antibody detection under certain conditions.

Use of this type of sample affords clear advantages, such as easy, rapid sample collection that does not require specialized personnel, a minimization of contamination risk for health professionals, a lack of trauma for the patient, and applicability to special population groups for whom blood sample collection is difficult (e.g., individuals addicted to parenterally administered drugs).

Meanwhile, other biological fluids with similar advantages, such as urine, are still being evaluated by various groups (10, 15, 16, 36).

The goal of the study described here was to evaluate GCT and urine samples as possible alternatives to serum for the detection of HIV antibodies with an automated system optimized for these sample types and confirmation by Western blot analysis (WB). The use of paired samples from the same individual facilitates better evaluation of the sensitivity and specificity of the technique with these alternative samples.

MATERIALS AND METHODS

A total of 302 individuals participated in the study: 187 infected with HIV type 1 (HIV-1) (106 had Centers for Disease Control and Prevention [CDC] stage II infection, 19 had CDC stage III infection, and 62 had CDC stage IV infection)

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Each strip contained a serum control (human anti-immunoglobulin antibody). The WB combined purified HIV-1 antigens (corresponding to structural genes gag, env, and pol) with an HIV-2 synthetic peptide (gp36) in one strip. This enzyme-linked fluorescent assay (ELFA) associates a final phosphate's conversion into a fluorescent 4-methyl-umbelliferyl product through liberation of the phosphate. The final readings are given as relative fluorescence detected in the reading (the alkaline phosphatase catalyzes the substrate's conversion into a fluorescent 4-methyl-umbelliferyl product through liberation of the phosphate). The final readings are given as relative fluorescence values (RFVs).

The system was adapted to detect IgG-type immunoglobulins in GCT and urine samples. The RFVs obtained with the GCT and urine samples belonging to the group of noninfected individuals were used to find the cutoff (10 times the average RFV obtained with urine samples [cutoff, 2.5 \times 4 = 10 RFVs]). Samples with RFVs above 10 were considered to be positive, and those with RFVs equal to or less than 10 RFVs were considered to be negative.

All samples were processed according to the manufacturer's instructions, with only one HIV antibody test performed with each sample type (GCT and urine).

The positive samples and negative samples belonging to seropositive individuals were confirmed by WB (Bioblot HIV-1 plus; Genelabs Diagnostic, Geneva, Switzerland). The WB combined purified HIV-1 antigens (corresponding to structural genes env, gag, and pol) with an HIV-2 synthetic peptide (gp36) in one strip. Each strip contained a serum control (human anti-immunoglobulin antibody).

The dilutions used were 1/10 (200 \mu l of GCT plus 1,800 \mu l of blotting buffer) for the GCT samples and 3/4 (1,500 \mu l of urine plus 500 \mu l of blotting buffer) for the urine samples. The GCT and urine samples were incubated for 16 to 20 h.

The 1990 World Health Organization (WHO) criterion was used as positive (reactivity to two bands including bands for gp41, gp120, or gp160) (46). Samples that did not meet this criterion but that displayed reactivities to other bands were classified as indeterminate. Negative samples did not display any reactive bands.

The result of the WB confirmation of HIV antibodies with serum was used as the "gold standard" for sensitivity and specificity calculations. Bonferroni's test was used to evaluate the significance between the RFVs thus obtained.

In order to find the most appropriate time for the collection of alternative samples, a preliminary trial was run before the material used in the study was obtained to evaluate possible variations in the elimination of HIV antibodies throughout the day in the same individual. To do so, six individuals with CDC stage II infection were asked at random to cooperate. They were tracked over 24 h. Sample collection (urine and GCT simultaneously) began when they woke up in the morning and then continued throughout the 24-h period with no set schedule; every time that they needed to urinate, both GCT and urine samples were collected as described above. All samples were analyzed by ELFA, and the result was confirmed by WB procedures described above. In all cases at least four paired saliva and urine samples were collected from the same participant.

RESULTS

All 187 GCT samples from infected individuals were reactive by ELFA (Table 1). Likewise, all GCT samples from noninfected individuals were negative. These results provided sensitivity and specificity values of 100% and positive and negative predictive values of 1 (P < 0.0001).

Detection of HIV antibodies in urine samples was positive by ELFA for 181 individuals (Table 2). Of these, 178 were individuals with some stage of HIV infection (102 with CDC stage II infection, 19 with CDC stage III infection, and 57 with CDC stage IV infection), and 3 were noninfected individuals. Nine samples from seropositive individuals were negative by ELFA (four and five samples belonging to individuals with CDC stage II and IV infections, respectively). Of the 115 urine samples from seronegative persons, only 3 urine samples displayed reactivity by the ELFA technique (1 sample belonged to...
TABLE 1. Confirmation of ELFA reactivities by WB of GCT samples

<table>
<thead>
<tr>
<th>Group</th>
<th>Total ELFA positive</th>
<th>WB positive</th>
<th>No. of GCT samples with the indicated results for HIV antibodies by the following assay:</th>
</tr>
</thead>
<tbody>
<tr>
<td>CDC stage II</td>
<td>106</td>
<td>106</td>
<td>No. of samples with the following RFVs:</td>
</tr>
<tr>
<td>CDC stage III</td>
<td>19</td>
<td>19</td>
<td>Total 302 187 187 187</td>
</tr>
<tr>
<td>CDC stage IV</td>
<td>62</td>
<td>62</td>
<td>Blood donors 46 1 1</td>
</tr>
<tr>
<td>Individuals with high risk of HIV infection</td>
<td>69</td>
<td>0</td>
<td>CDC stage I</td>
</tr>
<tr>
<td>Blood donors</td>
<td>46</td>
<td>0</td>
<td>CDC stage III</td>
</tr>
<tr>
<td>Total</td>
<td>302</td>
<td>187</td>
<td>CDC stage IV</td>
</tr>
</tbody>
</table>

* None of the samples had a negative or indeterminate result.

The distribution of RFVs with GCT and urine samples can be seen in Tables 3 and 4, respectively. It can be seen from the data in Table 3 that 77% of the samples from seronegative individuals yielded an RFV of 0 for 200, with no samples having values equal to or less than 10 RFVs. In Table 4 it can be seen that only 24.1% of the urine samples from HIV-seronegative individuals had RFVs of >200, while 5.3% of the samples had negative RFVs (four samples from individuals with CDC stage II infection and five samples from individuals with CDC stage IV infection). On the other hand, the seronegative individuals displayed very similar RFVs: an RFV of 0 for 47.8% of GCT samples and 44.3% of urine samples and RFVs of between 1 and 10 for 52.2 and 54.9% for GCT and urine samples, respectively. Among the samples from noninfected persons, three urine samples were positive by ELFA, with RFVs in the range of 11 to 50.

The average RFVs for GCT and urine samples from seronegative individuals were 946 ± 3,153 and 409 ± 980, respectively. Statistical comparison of the RFVs for samples from the HIV-infected groups did not show any significant differences between RFVs belonging to individuals with CDC stage II, III, and IV infections (Bonferroni's test). This was true for both the GCT and the urine samples. However, significant differences were found between each of these HIV infection stages and the values obtained for noninfected groups.

The ELFA results were confirmed by WB. All GCT samples reactive by ELFA fulfilled the WHO criterion for positivity (Table 1). In contrast, among the WBs performed with 190 urine samples, only 181 met the positivity criterion (96.8%) (Table 2). The samples were distributed by stage as follows: 104 samples belonged to individuals with CDC stage II infection (2 samples were negative by ELFA), 19 samples belonged to individuals with CDC stage III infection, and 58 samples belonged to individuals with CDC stage IV infection (1 sample was negative by ELFA). The seven samples with negative WB results included two urine samples each from individuals with CDC stage II and IV infections (the samples were negative by ELFA), one sample from the blood donor group (the sample was positive by ELFA), and two samples from the group of individuals in a group at high risk of HIV infection (the samples were positive by ELFA). Finally, two samples with indeterminate WB results were found (the samples were from individuals with CDC stage IV infection) and had bands that were reactive to proteins gp160 and gp41, respectively (both samples were negative by ELFA).

The reactivity band most frequently observed by WB was the gp160 protein band (in 100% of GCT samples versus 97.3% of urine samples), and the least common reactivity band was the p17 protein band (in 31.5% of GCT samples versus 21.4% of urine samples) (Tables 5 and 6). The arithmetic mean of the observed number of bands was 6.7 for GCT and 6.5 for urine. Nevertheless, 23.2% of the WBs run with GCT samples showed all nine bands, while 32.2% of the WBs run with urine samples yielded five or fewer reactive bands. In the WBs with GCT proportion dropped to 25.5%.

Only 1.6 and 0.5% of the WBs with GCT and urine, respéc-

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**DISCUSSION**

When gathering GCT samples for testing, the device that is used and the mouth site chosen for sample collection seem to be crucial to the results of the trial (29). However, the time of sample collection does not appear to be decisive if the results obtained in the follow-up performed with the six individuals infected with HIV (CDC stage II infection) over 24 h are taken into account. Several devices specifically designed for the collection of GCT samples exist (31, 32). For this project we selected a device which we feel affords a greater content of GCT in the final sample volume that is collected because the device remains in the lateral gingival fold long enough and has a large absorbent surface. Another advantage of this device is that it does not seem to alter the original concentration of IgG-type immunoglobulins present in the GCT sample. This is not true of the devices used in other studies (20, 43), which incorporate solutions that dilute the sample and that thus may cause a decrease in antibody levels and therefore the sensitivity of the detection technique. Nevertheless, use of such devices may be best when no freezing facility that can preserve the sample once it has been taken is available.

Screening for HIV antibodies in biological samples from populations for whom the final objective is not diagnosis involves the use of techniques in which the most important criterion for selection must be sensitivity (3, 9, 14, 18, 21, 35). The sensitivity shown in the ELFA study with GCT samples for the detection of HIV antibodies could make it acceptable for epidemiological studies.

Another important aspect is the possibility of using WB to confirm the reactivity of the alternative samples that are positive by the preliminary screening.

Most positive samples displayed reactivities to more than six bands. This feature seems to be favored by an increased sample volume and overnight incubation, which cause an increase in band intensity, which in turn facilitates band observation and

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**TABLE 5. Reactivity patterns of GCT samples from HIV-infected individuals by WB for HIV**

<table>
<thead>
<tr>
<th>Reactivity pattern</th>
<th>No. (%) of samples with the following bands by WB:</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ENV gp160 gp120 gp41 POL p66 p51 p31 GAG p55 p24 p17</td>
<td></td>
</tr>
<tr>
<td>ENV</td>
<td>9 9 9 9 6 0 0 0 0 0 0 0</td>
<td></td>
</tr>
<tr>
<td>ENV + GAG</td>
<td>9 9 9 7 0 0 0 0 2 9 0 0</td>
<td></td>
</tr>
<tr>
<td>ENV + POL</td>
<td>21 21 21 16 7 6 21 0 0 0</td>
<td></td>
</tr>
<tr>
<td>ENV + POL + GAG</td>
<td>148 148 148 145 113 126 130 105 138 59</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>187 187 (100) 187 (100) 174 (93) 120 (64.2) 132 (70.6) 151 (80.8) 107 (57.2) 147 (78.6) 59 (31.5)</td>
<td></td>
</tr>
</tbody>
</table>

**TABLE 6. Reactivity patterns of urine samples from HIV-infected individuals by WB for HIV**

<table>
<thead>
<tr>
<th>Reactivity pattern</th>
<th>No. (%) of samples with the following bands by WB:</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ENV gp160 gp120 gp41 POL p66 p51 p31 GAG p55 p24 p17</td>
<td></td>
</tr>
<tr>
<td>ENV</td>
<td>9 8 7 7 0 0 0 0 0 0 0</td>
<td></td>
</tr>
<tr>
<td>ENV + GAG</td>
<td>8 8 8 4 0 0 0 1 7 1</td>
<td></td>
</tr>
<tr>
<td>ENV + POL</td>
<td>31 31 31 22 11 6 26 0 0 0</td>
<td></td>
</tr>
<tr>
<td>ENV + POL + GAG</td>
<td>135 135 135 131 105 120 122 108 129 39</td>
<td></td>
</tr>
<tr>
<td>None</td>
<td>4 0 0 0 0 0 0 0 0 0</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>187 182 (97.3) 181 (96.8) 164 (87.7) 116 (62.0) 126 (67.4) 148 (79.1) 109 (58.3) 136 (72.7) 40 (21.4)</td>
<td></td>
</tr>
</tbody>
</table>

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In the Western blots displayed only two bands, those for the envelope proteins gp160 and gp120. In the Western blots with three bands (5.3% of GCT samples versus 6.4% of urine samples), in addition to the bands of reactivity to proteins gp160 and gp120, reactivity was observed to other proteins: gp41 (60% of GCT samples versus 50% of urine samples), p24 (20% of GCT samples versus 25% of urine samples), and p31 (20% of GCT samples versus 25% of urine samples).

Only WBs with urine samples displayed one reactive band (indeterminate WB results belonging to samples from individuals with CDC stage IV infection had two reactive bands to proteins gp160 and gp41, respectively).

Four reactivity patterns were observed for both types of samples: ENV (4.8% of GCT samples versus 4.8% of urine samples), ENV plus POL (11.2% of GCT samples versus 16.6% of urine samples), ENV plus GAG (4.8% of GCT samples versus 4.3% of urine samples), and ENV, POL, and GAG (79.2% of GCT samples versus 72.2% of urine samples).

Finally, Table 7 gives the results from the ELFA analysis of the GCT and urine samples taken over a 24-h period from a group of six volunteers infected with HIV (CDC stage II infection). These results draw attention to the disproportionate RFVs for GCT samples and their paired urine samples. In general, the RFVs for the GCT samples were always higher. The RFVs for samples subsequently obtained from each individual were similar. The same was true for urine samples. There seems to exist a certain parallel between the RFVs found for GCT samples and their paired urine samples. In general, the RFVs for GCT samples versus 72.2% of urine samples).
therefore enhances test sensitivity. Sometimes a short incubation period combined with very stringent criteria for positivity by WB can reduce the sensitivity of WB with alternative biological fluids (19).

As far as the intensities of the bands observed in the WBs with GCT samples is concerned, our results coincide with those of other researchers (31, 33, 39), who have observed that reactivity to HIV envelope antigens is usually strong, while reactivity to GAG antigens is weak.

The reactivity pattern most frequently observed by WB included bands to ENV, POL, and GAG, which makes for easier interpretation of the results of confirmation tests. The presence of a predominant reactivity to the envelope proteins (gp120 and gp160) in all of the WBs performed with GCT samples coincides with the results obtained by other groups (2, 19, 22, 29). This fact seems to indicate that the WHO criteria for serum samples could be used with GCT samples with no decrease in the operational characteristics in terms of the sensitivity and specificity of WB (28). In some studies, use of other criteria causes a drop in the sensitivity of WB with an increase in the number of samples with indeterminate results by WB (19, 22).

Our findings seem to indicate that HIV antibodies can be detected in GCT as well as in serum. Some studies have compared serum and GCT in a longitudinal manner during the period of immunocompetence and hypothesize that immunocompetence can likely be detected GCT and serum at about the same time (3, 12, 18).

The use of urine as an alternative to serum offers worse results than the use of GCT; the antibody concentration present in urine samples is much lower than that present in serum; some investigators state that it is even 10⁴ times less (13). On the other hand, other factors may influence the antibody concentration found in urine samples and may cause a sample to be nonreactive by the screening and confirmation techniques used in study, even though the sample belonged to an HIV-infected individual. Among such factors which would explain the presence of false-negative results are certain medications (i.e., diuretics), consumption of large quantities of liquids, urine collection without a long enough waiting period after the last urination to allow the antibody concentration to return to normal, and deterioration in the individual's immunological state (30). Other factors to be borne in mind when analyzing results for the three urine samples that had false-negative results by the screening test but that were reactive by WB are sample preservation time and lower ELFA sensitivity with respect to the results of WB. Therefore, for studies that rely on the analysis of urine samples, it seems recommendable that samples be analyzed as quickly as possible, even when they are preserved at 4°C. Another aspect which should be considered is sample freezing, because it seems that the processes of freezing and defrosting can cause a drop in the specificity of the

TABLE 7. Evolution of RFVs in GCT and urine samples over a 24-h period

<table>
<thead>
<tr>
<th>Individual</th>
<th>RFV in GCT (urine) samples for the following sample no.:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1</td>
</tr>
<tr>
<td>A</td>
<td>438 (53)</td>
</tr>
<tr>
<td>B</td>
<td>477 (90)</td>
</tr>
<tr>
<td>C</td>
<td>3,404 (200)</td>
</tr>
<tr>
<td>D</td>
<td>2,314 (102)</td>
</tr>
<tr>
<td>E</td>
<td>2,180 (115)</td>
</tr>
<tr>
<td>F</td>
<td>7,920 (4,288)</td>
</tr>
</tbody>
</table>

*a All individuals had CDC stage II HIV infection.

FIG. 2. Reactivities observed by WB of GCT and urine samples from the same individual obtained over 24 h.
technique (26). In our study none of the urine samples were frozen.

The intensities of the bands observed by the WB of urine samples were generally weaker than the intensities observed by WB of GCT samples. As with GCT samples, a predominant reactivity to envelope proteins (gp41, gp120, and gp160) was observed among the samples with reactive bands by WB. This particular reactivity and the possibility of detecting antibodies against all the structural antigens of the virus have already been described by Cao et al. (10).

As has been shown here, the results obtained with urine samples permit the use of this easily obtained and easily preserved sample in epidemiological studies that deal with large populations similar to those handled in this study. In contrast, use of urine samples for diagnosis does not seem to be advisable when one takes into consideration the sensitivities that were found.

The concentration in serum of an antibody against a specific infectious disease agent can undergo oscillations over time. Such variations tend to be slow and are rarely established in a matter of days except in patients in the acute stage of the infection. When samples other than serum are used for diagnosis of a chronic infection such as HIV infection, one problem that may arise is the presence of different HIV antibody concentrations in the fluids used as alternatives to serum (16, 23, 31). The concentration may be influenced by the antibody levels that previously existed in the serum, by the quantity of serum filtrate that passes to the fluids, or by its final dilution, depending on the secretion volume of the alternative fluid. With this in mind it seems reasonable to venture that the antibody concentration in urine may suffer more circadian alterations than that in GCT. According to Mortimer and Parry (31), the immunoglobulin concentration in GCT is between four and five times lower than in plasma. Slight fluctuations in RFVs for both GCT and urine throughout the day were observed in our study. At any rate, it should be indicated that just as when GCT and urine reactivities were observed, very significant quantitative reactivity differences were found between GCT and urine samples from the same individuals. The fact that the values for GCT samples were always higher than those for urine samples confirms what various investigators have indicated, that more immunoglobulins are present in sera than in urine samples (16, 23, 31). The concentration may be influenced by the antibody level in the fluids used as alternatives to serum (16, 23, 31). The concentration may be influenced by the antibody level in the fluids used as alternatives to serum (16, 23, 31).

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