Recombinant Flagellin A Proteins from *Borrelia burgdorferi* Sensu Stricto, *B. afzelii*, and *B. garinii* in Serodiagnosis of Lyme Borreliosis

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Genes for flagellin (FlaA) proteins from European borrelial strains of *Borrelia burgdorferi sensu stricto*, *B. afzelii*, and *B. garinii* were cloned and sequenced. An identity of 92 to 93% was observed in the flaA sequences of the different species. Polyhistidine-tagged recombinant FlaA (rFlaA) proteins were produced in *Escherichia coli* and used as antigens in Western blotting (WB) and enzyme-linked immunosorbent assay (ELISA). In immunoglobulin G (IgG) WB, 71% (10 of 14) of the sera from neuroborreliosis and 86% (12 of 14) of those from Lyme arthritis patients reacted with one to three rFlaAs. In IgG ELISA, 74% (14 of 19) and 79% (15 of 19) of patients with neuroborreliosis and arthritis, respectively, were positive. The immunoreactivity in local European patient sera was stronger against rFlaA from *B. garinii* and *B. afzelii* than against rFlaA from *B. burgdorferi sensu stricto*. Neither IgG nor IgM ELISA was sensitive in the serodiagnosis of erythema migrans. Serum samples from patients with syphilis and systemic lupus erythematosus showed mild cross-reactivity in IgG tests. Sera from *Yersinia enterocolitica* or beta-hemolytic *Streptococcus* infections showed only occasional responses. With IgM ELISA, 58% (11 of 19) and 37% (7 of 19) of patients with neuroborreliosis and arthritis, respectively, were positive. Cross-reactive antibodies to FlaA, especially in serum samples from patients with rheumatoid factor positivity and Epstein-Barr virus infection, reduced the specificity of IgM serodiagnosis. Therefore, rFlaA seems to have a limited role for IgM serodiagnosis, yet rFlaA might be useful in the IgG serodiagnosis of disseminated Lyme borreliosis.

Laboratory diagnosis of Lyme borreliosis (LB) is mainly based on serology, although the present serologic tests have unsatisfactory sensitivity and specificity (34). Routine laboratory testing uses enzyme-linked immunosorbent assays (ELISA) with borrelial whole-cell lysate (WCL) or flagella (consisting mainly of polymerized FlaB protein) as the most commonly used antigens. A two-step approach with ELISA followed by a confirmatory Western blot (WB) has been recommended for positive or borderline results (19, 36). Especially in Europe, the applicability of this procedure has, however, remained doubtful (3, 14) because three or more borrelial species cause LB (38). Several difficulties complicate LB serology. Firstly, immunoglobulin G (IgG) antibody responses are often delayed during the early stages of LB. Even at late-stage LB, 5 to 10% of patients do not have elevated antibody levels (29), perhaps due to diversion of the host immune response towards Th1 immunity by borreliar factors (17). Secondly, viral infections cause false-positive IgM results in several LB tests (4). Thirdly, in a subgroup of patients antibody levels may stay high after successful treatment of LB even for prolonged periods (6, 15).

Several recombinant borreliar antigens (OspA, OspB, OspC, OspE, OspF, p22, BmpA, BBK32, BBK50, VlsE, p100, 14-kDa internal flagellin fragment) (5, 7, 16, 21, 22, 24, 25, 31, 35) and chimeric borreliar proteins OspA, OspB, OspC, flagellin (p41), and p93 (13) have been studied to improve serologic diagnosis. Of these proteins, BmpA (32) and OspC (8, 26, 27, 30, 31) have been suggested as antigens which induce early IgM responses. However, IgG antibodies to recombinant BmpA have been detected mainly in long-standing disease (29, 32). A limiting factor in the use of OspC as a diagnostic antigen is the extensive structural variation of the molecule between borreliar species (18, 23). Use of recombinant antigens has increased the specificity of serologic assays, but the sensitivity of tests using single antigens has thus far remained disappointing. In Europe, where the sequence heterogeneity of antigenic proteins in various borreliar species and strains complicates LB serology (33), supplementary information is needed on the differences between the antigenic properties of the borreliar species.

Flagellin A (FlaA) is a 37-kDa outer sheath protein of the periplasmic *Borrelia burgdorferi* flagella. It has been suggested that FlaA could potentially be a useful antigen for detecting antibodies in early LB. Gilmore et al. (12) obtained promising results for erythema migrans (EM) patients with IgM WB using recombinant FlaA (rFlaA) as an antigen. In contrast, Ge et al. (9) failed to show any useful serologic role in LB for another rFlaA construct. The purpose of the present study was to expand our knowledge of FlaA proteins in *B. burgdorferi sensu lato* spirochetes. We present the cloning and expression of FlaA proteins from three European borreliar strains of *B. burgdorferi sensu stricto*, *B. afzelii*, and *B. garinii* and the results...
of WB assays and ELISAs using FlaA recombinants as antigens.

MATERIALS AND METHODS

Borrelial strains. Domestic borrelial strains of *B. burgdorferi* sensu stricto (ia) isolated from cerebrospinal fluid and *B. afzelii* (A91) and *B. garinii* (46) isolated from skin biopsies of Finnish patients with LB were used. The genotyping of these strains was performed by PCR of *flaB* and subsequent sequencing of the PCR products, as described previously (20).

Borrelia culture and DNA isolation. Borrelial strains were cultured in Barbour-Stoenner-Kelly-H medium (Sigma) at 33°C in a 5% CO₂ atmosphere until the growth was approximately 2 × 10⁸ cells/ml. The genomic DNA was isolated with a DNeasy Tissue kit (Qiagen, Hilden, Germany).

PCR and cloning of the genes. For each borrelial strain the *flaA* sequence was studied by PCR amplification of the genomic DNA (Table 1). Approximately 1 ng of template DNA was used and the parameters in the PCR amplification reaction were 30 cycles at 94°C for 1 min, 50°C for 1 min, and 72°C for 1 min 30 s with AmpliTaq Gold DNA polymerase (Perkin Elmer). DNA products were visualized by gel electrophoresis on a 1% Agarose NA gel (Amersham Pharmacia, Uppsala, Sweden) containing ethidium bromide. The PCR products were cloned into the pCR 2.1-TOPO vector (Invitrogen, Groningen, The Netherlands). *Escherichia coli* INFA (Invitrogen) host cells were used for cloning.

DNA sequencing. Plasmid DNA containing *flaA* inserts was isolated from *E. coli* by using a QIAprep-Spin plasmid kit (Qiagen). DNA sequencing was performed with the DyePrimer (T7, M13Rev) cycle sequencing kit (Applied Biosystems Inc.) in accordance with the manufacturer’s instructions by the Core Facility of the Haartman Institute, University of Helsinki. Sequencing reactions were run and analyzed by an automated sequencing apparatus model 373A (Applied Biosystems, Inc.). DNA and protein sequences were analyzed with Lasergene software (DNASTAR, Inc.). To eliminate possible sequencing errors, the whole gene was sequenced twice, and in some cases three times, following discordant results.

Construction of the expression plasmid and expression of rFlaA. The starting codon for the *flaA* construct was chosen according to previous successful experiences of other investigators to express FlaA (12). The *flaA* constructs included the protein coding sequence without the sequence for the leader and three subsequent codons encoding the first three amino acids after a predicted signal.

FIG. 1. FlaA amino acid sequence comparison of the translated full-length gene in *B. burgdorferi* sensu stricto strains B31, 212, and ia (Bbia), *B. afzelii* A91 (BaA91), and *B. garinii* 46 (Bg46). Boxes indicate amino acid heterogeneity. The starting point of our recombinant constructs is indicated by an arrow.

![Figure 1](http://jcm.asm.org/issue/10/issue/10_0001.png)

**TABLE 1.** Primers used in PCR reactions for *flaA* sequencing and expression of the respective protein.

<table>
<thead>
<tr>
<th>Target DNA</th>
<th>Primer (5'-3')y</th>
<th>Location (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Sequencing</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>ATA TAA GGA GTT GGT TTA CAT</td>
<td>−19–2</td>
</tr>
<tr>
<td>2</td>
<td>ATG AAA AGG AAA GCT AAA AGT</td>
<td>1–21</td>
</tr>
<tr>
<td>3</td>
<td>GCT ATT CTC AAT CAT CTG C</td>
<td>386–404</td>
</tr>
<tr>
<td>4</td>
<td>TTT ATG AGA CTA GGC GAA CT</td>
<td>860–879</td>
</tr>
<tr>
<td>5</td>
<td>TTA AAT AAA CCT TGC CAT CAA</td>
<td>1,438–1,418</td>
</tr>
<tr>
<td>6</td>
<td>TTT TCT AAA TCT AAT ATT TCC AT</td>
<td>1,117–1,094</td>
</tr>
<tr>
<td>7</td>
<td>CAT TTT ACT TGA AGC AAG AG</td>
<td>717–698</td>
</tr>
</tbody>
</table>

**Expression**

|      | CCG CAT CCG ATG GAT TAG CAG AGG GTT | 67–85           |
|      | CCG GTA CCC TAA TTT TTC GGA GAT GAT TC | 1,026–1,006   |
|      | CCG CAT CCG ATG GAT TAG CAG AGG GC  | 67–84          |
|      | CCG GTA CCC TAA TTT TTC GGA GAT GAT TC | 1,026–1,006   |
|      | CCG CAT CCG ATG GAT TAG CAG AGG GC  | 67–84          |
|      | CCG GTA CCC TAC TTT TTT AAG GAT GAT TC | 1,029–1,009 |

* Underlining indicates BamHI and KpnI cleavage sites.
peptide I cleavage site (28) (Fig. 1). Expression primers for flaI of each strain were designed individually (Table 1), and the sense area to be expressed was multiplied by PCR and inserted into the pCR 2.1-TOPO vector as above. The plasmid containing the insert was purified and digested with BamH1 and KpnI. The cleaved flaI was ligated into similarly cut pQE-30 expression plasmid (Qia-
gen) with T4 ligase. The ligation mixture was used to transform E. coli M15 host cells as described in the manufacturer’s instructions (Qiagen). The transformation mixture was plated onto Luria-Bertani agar containing 100 μg of ampicillin and 25 μg of kanamycin per ml. A primary culture for expression of rFlaA was started by inoculating a single colony from a fresh transformant plate into 50 ml of Luria-Bertani broth containing antibiotics as described above. The culture was incubated at 37°C with shaking overnight. This starter culture was diluted 1:100 to 1,500 ml of Luria-Bertani broth with antibiotics as described above and incubated at 37°C for 3 h (the growth reached the mid-log phase; the optical density at 600 nm was ca. 0.6). Isopropyl-β-D-thiogalactoside (Calbiochem) was added to a final concentration of 0.6 mM, and an additional incubation of 3 h was performed. The cells were harvested, washed, and sonicated. The expressed rFlaA was recovered as insoluble protein in the centrifugation pellet. This was dissolved in 4 M guanidinium hydrochloride and attached by its oxidHistidine tag to a Chelating Sepharose Fast Flow column (Amersham Pharmacia) containing Ni2+ rFlaA was subsequently eluted from the column by 160 mM potassium phosphate (pH 7.2). After centrifugation the resulting insoluble protein pellet was resolubilized in 4 M guanidinium hydrochloride eluting buffer. Protein expression and purity were confirmed by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE).

**RESULTS**

Cloning and nucleotide sequence analysis of *flaA*. *flaA* nucleotide sequences of the borrelial strains were obtained by PCR and sequencing. The primers were designed according to the published flanking region sequences to ensure correct sequences at the gene ends. The analyzed sequences were compared with the published *flaA* sequences of *B. burgdorferi* sensu stricto strains B31 and 212. The *flaA* sequence of the *B. burgdorferi* sensu stricto strain ia was identical with that of the American strain B31 whereas it differed by nine nucleotides (identity of 99% at the nucleotide level) from the *flaA* sequence in strain 212. The identity of *flaA* sequences between the Finnish strains *B. burgdorferi* sensu stricto ia and *B. afzelii* A91, between *B. burgdorferi* sensu stricto ia and *B. garinii* 46, and between *B. afzelii* A91 and *B. garinii* 46 was 93, 92, and 95%, respectively. The identity of *flaA* in *B. afzelii* A91 and in *B. garinii* 46 compared to *flaA* in strain 212 was 92 and 91%, respectively.

**Protein sequence analysis.** The deduced amino acid sequences of *flaA* were compared with those from *B. burgdorferi* sensu stricto, *B. afzelii* A91, and *B. garinii* 46 were 95% identical (Fig. 1). The identity of both *flaA* and *flaA* with *flaA* of *B. burgdorferi* sensu stricto 212 was 94%. The *flaA* gene of *B. garinii* 46 has an additional triplet of nucleotides, AAA, at positions 1000 to 1002, leading to a lysine insertion into an otherwise hydrophilic sequence area. None of the sequence differences change the local hydrophilicity of the sequence area, indicating conservation of the overall structure (Fig. 1). The deduced amino acid sequence of *FlaA* protein (without the leader peptide) was rich in phenylalanine and tyrosine (15 and 16 amino acids, respectively) but contained no cysteine. The count of these amino acids was identical in all three *FlaA* proteins examined in this study. Thirty-three percent of the deduced amino acids were hydrophobic. The calculated (Lasergene Software; DNASTAR, Inc.) isoelectric points of *flaA* were 6.23, 6.49, and 6.35 for *B. burgdorferi* sensu stricto, *B. afzelii* A91, and *B. garinii* 46 for the mature protein without leader sequence were 5.70, 5.33, and 7.27, respectively. Three additional lysines and fewer aspartic acids in *flaA* than in *flaA* and *flaA* may account for the higher isoelectric point of *flaA*. 

**IgG WB.** For the WB analysis, serum samples from 14 patients with NB, 14 with LA, 10 with syphilis, and 13 healthy blood donors were tested. Ten samples from patients with NB (71%) and 12 from patients with LA (86%) reacted with one or
TABLE 2. IgG WB results with rFlaA antigen from B. garinii, B. afzelii, and B. burgdorferi sensu stricto

<table>
<thead>
<tr>
<th>Antigen</th>
<th>NB patients (n = 14)</th>
<th>LA patients (n = 14)</th>
<th>Syphilis patients (n = 10)</th>
<th>Healthy blood donors (n = 13)</th>
</tr>
</thead>
<tbody>
<tr>
<td>rFlaA_Bg46</td>
<td>9</td>
<td>12</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>rFlaA_AfA91</td>
<td>6</td>
<td>9</td>
<td>6</td>
<td>0</td>
</tr>
<tr>
<td>rFlaA_Bbia</td>
<td>6</td>
<td>5</td>
<td>5</td>
<td>2</td>
</tr>
<tr>
<td>Total*</td>
<td>10</td>
<td>12</td>
<td>2</td>
<td>1</td>
</tr>
</tbody>
</table>

* Total refers to the total number of patients in each group having antibodies to one or more of the rFlaA proteins.

more of the rFlaAs. Two of 10 patients with syphilis and 1 out of 13 healthy blood donors were positive. The combined results of the WB analysis are presented in Table 2. rFlaA_Bg46 and rFlaA_BaA91 were superior to rFlaA_Bbia in detecting antibodies to FlaA.

**IgM WB.** In repeated experiments using serum samples from patients with EM or NB, we consistently observed moderate to strong IgM immunoreactivity to rFlaAs, yet samples from patients with EBV infection or RF positivity were similarly immunoreactive. However, the evaluation of the immunoblot bands visually and with the MacBAS program showed differences in the intensity of the bands (data not shown). Based on the intensities of healthy blood donor serum reactions, one of five samples from patients with EM, four of five from patients with NB, two of five from patients with EBV, and two of five with RF positivity gave positive IgM reactions for one or more of the rFlaAs. Alterations in the dilution of the serum (1:25 to 1:500), the buffer (varying the protein and using different salt concentrations), or the applied conjugate did not lead to any better discrimination of the IgM bands between the patients and the controls (data not shown).

**IgG ELISA.** For IgG ELISA optical density values above the means plus 2 SD of values of healthy blood donors were defined as positive. Results for IgG ELISA of B. garinii and B. burgdorferi were comparable with the results obtained from IgG WB analysis: positive reactions were found mainly against rFlaA_Bg46 and rFlaA_BaA91 while reactions to rFlaA_Bbia were less frequent (Fig. 2). Of the samples from 15 patients with EM, two (13%) of the acute-phase samples and three (20%) of the convalescent-phase samples reacted with one or more of the rFlaAs. Samples from 14 of the 19 (74%) patients with NB and from 15 of 19 (79%) patients with LA reacted with either rFlaA_Bg46 or rFlaA_BaA91. The 19 patient samples of NB and LA used here include those examined by IgG WB. None of the samples from patients with NB or LA reacted with rFlaA_Bbia only. Twelve of the 14 positive samples from NB patients recognized both rFlaA_BaA91 and rFlaA_Bg46 and the remaining 2 recognized rFlaA_Bg46 only. Of the 15 positive samples from LA patients, 11 recognized both antigens, 2 recognized rFlaA_BaA91 only, and 2 recognized rFlaA_Bg46 only. Depending on the rFlaA antigen used, 1 to 3 of 10 patients with syphilis, 1 to 2 of 8 patients with RF positivity, 0 to 3 of 8 patients with ASO positivity, 1 to 3 of 8 patients with EBV infection, 0 to 2 of 8 patients with *Yersinia* infection, 2 to 3 of 8 patients with SLE, and 2 of 27 healthy blood donors were positive.

**DISCUSSION**

In the present study, flaA sequences of *B. afzelii* and *B. garinii* are reported for the first time. The identity of the flaA sequences of *B. afzelii*, *B. garinii*, and *B. burgdorferi* sensu stricto was over 90%. Although the exact function of the FlaA protein is not known, it has been shown that periplastic flagella of *B. burgdorferi* comprise a core protein FlaB and an outer sheath protein FlaA (11). Given the essential role of flagella in motility of the spirochete it is not surprising that the flaA was highly conserved among *B. burgdorferi* sensu lato strains.

FlaA has been evaluated as a WB antigen in LB serodiagnosis in only a few studies, and the results have been contradictory (9, 12). No previous studies using rFlaA in ELISA have been published. Our aim was to study the antigenic potential of FlaA in the European context. Therefore, we cloned flaA from all three borrelial species and produced the respective recombinant proteins. For IgG serodiagnosis of NB and LA, FlaA appeared to be a sensitive antigen. Seventy-one or seventy-four percent of patients with NB were positive either by WB or ELISA, respectively. The corresponding positivities for LA patient samples were 86 and 79%. The specificity in our series was slightly lowered due to low positive values occurring more frequently in samples from other diseases than in samples from healthy controls. Although the sequences of the three FlaA proteins were highly homologous, rFlaA antigens from *B. garinii* and *B. afzelii* seemed to perform better regarding sensitivity and specificity than FlaA from *B. burgdorferi* sensu stricto. Obviously, various FlaA proteins have both common and divergent antigenic epitopes. In Europe, *B. garinii* and *B. afzelii* are the most prevalent genospecies (38). This concurs with the predominance of immunoreactivity to FlaA from these two genospecies.

In IgM serology our patient selection differed from the series by Gilmore et al. (12). They studied EM patient sera with antibodies to the *B. burgdorferi* 37-kDa protein (putative FlaA antigen) in WCL immunoblot which also reacted with the rFlaA. However, only 38 and 57% of all patients with EM at the acute and convalescent phase, respectively, had antibodies to rFlaA (12). Serum samples from EM patients with no observed immunoreactivity against the *B. burgdorferi* 37-kDa protein did not recognize rFlaA (12). It is not clear how the group of EM patients without immunoreactivity to rFlaA differed from those reacting positively with rFlaA. In our study with unselected culture-confirmed EM patients, sensitivity of the IgM ELISA was performed with the same panel of patient and control samples as the IgG ELISA. Of the 15 samples from patients with EM, 3 (20%) of the acute-phase samples and 4 (27%) of the convalescent-phase samples reacted with one or more of the rFlaAs. Samples from 11 of the 19 (58%) patients with NB and from 7 of 19 (37%) patients with LA reacted with one or more of the rFlaAs. Depending on the rFlaA antigen used, 0 to 1 of 10 patients with syphilis, 2 to 4 of 8 patients with RF positivity, 1 of 8 patients with ASO positivity, 2 to 6 of 8 patients with EBV infection, 1 to 4 of 8 patients with *Yersinia* infection, 1 of 8 patients with SLE, and 1 of 27 healthy blood donors were positive. Results with three variant rFlaAs as antigen in the IgM ELISA are shown in Fig. 2.
IgM serology in WB and ELISA assays was only 20 to 27%. Another study failed to show FlaA as an immunodominant antigen in LB (9). The authors did not specify whether an IgM or IgG WB or early- or convalescent-phase serum samples of LB patients were used. In our assays for EM patient samples, IgG ELISA performed almost equally with IgM ELISA. For NB patients, IgG FlaA ELISA had better sensitivity than IgM ELISA. It is possible that our patient selection, including NB patients at early and late stages, may account for this finding.

Furthermore, cross-reactive antibodies to FlaA reduced the specificity of IgM serodiagnosis both in WB and ELISA experiments. Possibly, antibodies to FlaA of ubiquitous bacteria account for the cross-reactivity in IgM serology. Therefore, we do not see rFlaA as a useful antigen in IgM serodiagnosis for early LB.

As is typical for signal sequences, the leader of the FlaA protein includes a central hydrophobic segment (10). Trials to express a construct with the N-terminal leader have failed (12), perhaps due to toxicity to the E. coli host cell. However, as shown by Gilmore et al. (12) and in our study, FlaA without the leader peptide was well expressed, yet our recombinant protein tended to form inclusion bodies. The Protein Analysis program (Lasergene) predicted a \( \beta \)-sheet-rich region between amino acids 50 and 180 of the full-length protein (data not shown). We anticipate that this region of FlaA is involved in binding to the FlaB protein in the flagellar structure and may contribute to its property to form inclusions. A more hydrophilic construct formed by exclusion of the N-terminal part of FlaA might lead to a soluble recombinant protein. However, in another laboratory a construct where the first 79 amino acids were excluded failed to lead to successful expression (12).

Unlike in other spirochetes, B. burgdorferi FlaA is expressed at a considerably lower level than FlaB (11). The surface of borrelial flagella may have only a small amount of FlaA protein.
attached to the FlaB backbone, leaving a large part of the FlaB surface uncovered. This is in accordance with the observed difficulty of demonstrating flagellar shearlas in Borrelia (2). The smaller amount of FlaA protein expressed by Borrelia spp. during infection than of FlaB may account for the difficulty in finding specific IgM or IgG to FlaA and to the lower proportion of patients with positive immunoreactivity to FlaA than to FlaB. Alternatively, differences in the time of collection of sera with respect to clinical course might cause discrepancies in the immunoreactivity.

Besides Borrelia immunity, FlaA in other bacteria may be associated with interesting phenomena. In another spirochete, B. valaisiana, the flaA 37-kilodalton immunoblot band (P37) used in serodiagnosis of early Lyme disease is the flaA gene product. J. Clin. Microbiol. 37:548–552.


References
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