Detection of a $\text{bla}_{\text{SHV}}$ Extended-Spectrum β-Lactamase in *Salmonella enterica* Serovar Newport MDR-AmpC

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*Salmonella enterica* serovar Newport MDR-AmpC expressing TEM-1b and extended-spectrum β-lactamase SHV-12 was isolated from affected animals during an outbreak of salmonellosis that led to a 3-month closure of one of the largest equine hospitals in the United States.

*Salmonella enterica* serovar Newport MDR-AmpC is undergoing epidemiologic spread in animals and humans in the United States (4, 5, 7, 18, 22–24). This strain is characterized by a plasmid-mediated *ampC* gene (*bla*<sub>CMY-2</sub>) that encodes resistance to extended-spectrum cephalosporins (3, 5, 22). Serovar Newport MDR-AmpC strains are commonly resistant to ampicillin, chloramphenicol, streptomycin, sulfonamides, tetracycline, amoxicillin-clavulanate, cefoxitin, and cefotiofur and show reduced susceptibility to ceftriaxone (23). Serovar Newport MDR-AmpC was isolated from animals between July 2003 and May 2004 during what has now been defined as a protracted outbreak of salmonellosis that subsequently led to a 3-month closure of the George D. Widener Hospital for large animals at the University of Pennsylvania’s New Bolton Center (NBC), one of the largest equine hospitals in the United States.

*Salmonella* strains that express extended-spectrum β-lactamases (ESBLs) are rare. Coexistence of *bla*<sub>CMY</sub> and ESBL mechanisms in the same *Salmonella* strain has been documented infrequently in the United States and has been associated with foreign travel (8). To our knowledge, this is the first report of ESBL genes in an *S. enterica* serovar Newport MDR-AmpC strain.

All *Salmonella* isolates from the NBC clinical microbiology laboratory that were obtained from July 2003 to May 2004 were referred to the *Salmonella* Reference Center, also at NBC, for serotype confirmation and molecular characterization. Antimicrobial susceptibility profiles of *S. enterica* serovar Newport isolates were determined using a Sensititre CMV2ECOF Companion/Equine MIC veterinary plate (Trek Diagnostics, Cleveland, Ohio), and the interpretation of breakpoints was as determined by the manufacturer according to CLSI (formerly NCCLS) guidelines, when available (14–16). Serovar Newport strain SRC0307-213, isolated in July 2003 and identified as the index case, was shown to be resistant to ampicillin, chloramphenicol, tetracycline, cephalexin, cefotiofur, amoxicillin-clavulanate, gentamicin, and trimethoprim-sulfamethoxazole. Isolates from all of the 60 *S. enterica* serovar Newport cases included in the present report were susceptible to amikacin, imipenem, and enrofloxacin.

Plasmids were transferred by conjugation from *S. enterica* serovar Newport strain SRC0307-213 to a nalidixic acid-resistant *Escherichia coli* recipient strain (ATCC 27662) using nalidixic acid (50 µg/ml) and ampicillin (50 µg/ml) as the selective agents. Isoelectric focusing (IEF) for β-lactamases was performed on all strains at the Centers for Disease Control and Prevention (CDC) using a small-scale freeze-thaw method (12). Detection of *bla*<sub>CMY</sub> and *bla*<sub>TEM</sub> genes was performed by PCR at NBC as described previously (18). Detection of *bla*<sub>SHV</sub> was performed at CDC using a modification of the method described by Rasheed and colleagues (19). Briefly, the *bla*<sub>SHV</sub> gene was amplified on a PTC-200 DNA engine (MJResearch, Waltham, MA) in a final volume of 25 µl. A final primer concentration of 0.2 µM was used, and cycling conditions were as follows: 96°C for 5 min, 35 cycles of 96°C for 1 min, 50°C for 1 min, and 72°C for 1 min, and a final extension at 72°C for 10 min.

β-Lactamase genes were sequenced using standard methods. Briefly, total genomic DNA was extracted from SRC0307-213 and *E. coli* transconjugants using a Wizard genomic DNA purification kit (Promega Corp., Madison, WI). β-Lactamase genes were amplified with Promega 2× PCR master mix using primers shown in Table 1. PCR amplicons were cleaned up using a QiaQuick PCR purification kit (QIAGEN, Valencia, CA). Clean amplicons were used to obtain 2× DNA sequence coverage in both directions using a CEQ2000 capillary sequencer (Beckman Coulter, Fullerton, CA), and sequence alignments were performed using SeqManII version 5.06 (DNASTar Inc., Madison, WI).

*Salmonella enterica* serovar Newport SRC0307-213 was phenotypically negative for ESBL production by double-disk diffusion testing with ceftazidime and ceftazidime-clavulanate and also cefotaxime and cefotaxime-clavulanate (BD BBL Sensi-Disk; Becton Dickinson, Franklin Lakes, NJ) but was positive for *bla*<sub>CMY</sub>, *bla*<sub>TEM</sub>, and *bla*<sub>SHV</sub> genes by PCR. It has been noted previously that the presence of an ESBL can be masked by the expression of an AmpC-like enzyme, such as *bla*<sub>CMY</sub> (1, 2, 11). IEF showed enzymes with pIs of ≈8.4, 8.0, and 5.4, consistent with the CMY, SHV, and TEM enzymes, respectively. Two *E. coli* transconju-
transconjugants showed that only two Enterobacteriaceae grants from the Pennsylvania Department of Agriculture. bla 

gants (SRC0307-213-1 and SRC0307-213-2) were tested by dou-

bacteria Resistance Monitoring System indicate that many express a 
port MDR-AmpC strains submitted to the National Antimicro-

because certain combinations of genes could effectively limit all 
AmpC has considerable implications for veterinary and public 

This is the first report of an ESBL-producing 

bla 

BLA-producing salmonellae are rare in the United States, and 
this is the first report of an ESBL-producing S. enterica serovar 
Newport MDR-AmpC strain from animals. bla 

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TABLE 1. Oligonucleotide primers used to amplify genes for 

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