Bifidobacterial Lipoglycan as a New Cause for False-Positive Platelia Aspergillus Enzyme-Linked Immunosorbent Assay Reactivity

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We previously hypothesized that a lipoglycan of Bifidobacterium bifidum subsp. pennsylvanicum cross-reacts with the Platelia Aspergillus (PA) enzyme-linked immunosorbent assay (ELISA) based on the presence of galactofuranosyl epitopes in the cell wall (M. A. S. H. Mennink-Kersten, R. R. Klont, A. Warris, H. J. M. Op den Camp, and P. E. Verweij, Lancet 363:325–327, 2004). We tested this hypothesis by testing bacterial suspensions of different bifidobacterial species and other gram-positive and -negative bacteria with the PA ELISA, which is used to detect circulating galactomannan for the serodiagnosis of invasive aspergillosis. Furthermore, neonatal fecal samples were enumerated for bifidobacteria by fluorescence in situ hybridization (FISH) and tested for PA ELISA reactivity. All bifidobacteria, except B. infantis and B. adolescentis, showed reactivity 6- to 600-fold higher compared to the controls (i.e., Micrococcus luteus and Propionibacterium freudenreichii, which contain a cell wall lipomannan). Eggertella lenta showed a 25-fold-higher reactivity. ELISA reactivity was clearly shown to be associated with bacterial lipoglycans containing a β-1,5-galactofuranosyl chain. All neonatal feces showed PA ELISA reactivity and associated numbers of bifidobacteria. Since high concentrations of bifidobacteria are present in the human gut, these bacteria or excreted lipoglycan may cause false serum PA ELISA reactivity in selected patient groups, especially neonates.

Invasive aspergillosis (IA) has become a leading cause of death among immunocompromised patients. A commercial sandwich enzyme-linked immunosorbent assay (ELISA) (Platelia Aspergillus [PA] ELISA; Bio-Rad, Marnes-la-Coquette, France) that is widely used as a diagnostic tool in centers throughout the world detects a fungal antigen which is present in body fluids of patients with IA (5, 24, 44). The PA ELISA specifically detects circulating galactomannan (GM), a component of the total microflora of adults and up to 91% and 75% of the total fecal microflora of adults and up to 91% and 75% of the total fecal microflora in breast-fed and formula-fed infants, respectively (15, 37).

LTAs represent a group of structurally related lipid macroamphiphiles which are hydrophobically anchored to the cytoplasmic membrane, while the hydrophilic chain (mostly polyglycerolphosphate) penetrates the peptidoglycan network and may be detectable as a surface antigen (Fig. 1) (54). In the presence of bifidobacteria that cause bacteremia (47). Cross-reactivity has been documented with blood products and antigens from Aspergillus spp., but these molds very rarely cause invasive infection in humans (47). Besides resulting from IA, circulating Aspergillus GM could result from massive colonization of the gastrointestinal tract by Aspergillus (39) or from foods that are contaminated with Aspergillus (4) or contain its GM (1, 10). This fecal GM may reach the circulation in patients with dysfunction of the intestinal mucosal barrier. Furthermore, intravenous administration of the antibiotics piperacillin-tazobactam and amoxicillin-clavulanic acid, which show in vitro PA reactivity, has been shown to cause serum PA ELISA reactivity in patients without evidence of IA (26, 52). This PA reactivity is probably due to GM that originates from Penicillium that is used for antibiotic production (51).

After a review of the literature for microorganisms that contain galactofuranoses, it was found that a membrane-associated molecule of Bifidobacterium bifidum subsp. pennsylvanicum contained a terminal linear polysaccharide of more than seven β-1,5-linked Galf residues which mimics the epitope recognized by EB-A2 (7). It was suggested that lipoteichoic acid (LTA)-like amphiphiles of bifidobacterial species might cause PA ELISA reactivity in neonates after translocation because of the immaturity of the intestinal mucosa (28). In addition, Bifidobacterium species are members of the normal gastrointestinal microflora of humans, comprising up to 6% of the total fecal microflora of adults and up to 91% and 75% of the total fecal microflora in breast-fed and formula-fed infants, respectively (15, 37).

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high-G+C subdivision of gram-positive bacteria, LTAs are functionally replaced by lipoglycans, which contain a linear or branched polysaccharide as a hydrophilic moiety; this polysaccharide may carry monoglycerophosphate side chains (Bifidobacterium species) or succinyl esters (lipomannan from Micrococcus, Mycobacterium, or Propionibacterium) (9). The macroamphiphiles are secreted even during normal growth both with and without their lipid anchor. Secretion is stimulated by exposure to β-lactam antibiotics (9, 12, 33, 35). In aqueous solutions, the acylated molecules form micellar aggregates (53).

The aim of our research was to test the in vitro reactivity of different bacteria including bifidobacterial species and other members of the gut microbiota of humans with the PA ELISA. Any reactivity was correlated with the presence and structure of bacterial lipid macroamphiphiles based on review of the literature.

MATERIALS AND METHODS

Bacterial strains and culture conditions. Reactivity with the PA ELISA was investigated in vitro by testing cell suspensions of a range of bacteria, including 11 bifidobacterial species and 15 other bacteria (gram-positive and -negative species), most known to be common inhabitants from the human gut (Table 1). For each bacterium the structure of the lipid macroamphiphile was documented based on review of the literature (7–9, 13, 16, 18, 30, 48). Bacteria were cultured anaerobically (80% N2, 10% CO2, 10% H2) on agar plates containing fastidious anaerobic agar (Lab M) or aerobically on Columbia agar (BBL), at 37°C. B. bifidum subsp. pennsylvanicum was also cultured on reinforced clostridial medium (RCM; Oxoid). Because this liquid medium showed ELISA reactivity, it was hydrolyzed with acid (pH 2 with HCl for 3 h at 100°C) and neutralized with NaOH at pH 6.8 before inoculation. Acid hydrolysis removes Galf chains that are present in the RCM, which originate from contaminated medium components like meat extracts.

Fecal sample collection and FISH analysis. Fecal samples from nine healthy neonates were obtained. Babies were fully breast-fed or formula fed and ranged in age from 3 days to 4.5 months. No baby had been on antibiotic treatment prior to fecal sampling. Fecal samples were stored at −80°C until shipment on dry ice to Groningen for fluorescence in situ hybridization (FISH) analysis. Samples were processed and FISH analysis was performed as described by Harmsen et al. (14). The bifidobacterial probe Bi6164 (21), a 16s rDNA-based oligonucleotide probe, was used to enumerate total bifidobacteria in the fecal samples.

PA ELISA. Bacterial cells were scraped of the agar plates and suspended in 0.9% NaCl (wt/vol). Fresh fecal samples were suspended in saline to give a final concentration of 100 mg/ml (wet weight). The complete mixtures were serial diluted and used for detection of reactivity by the PA ELISA. The PA ELISA was performed according to the manufacturer’s instructions. However, the pretreatment step intended to dissociate immune complexes was omitted, except for the R3, R4, and R5 calibration samples (serum spiked with GM) and the fecal samples. Briefly, 50 μl of a reaction mixture containing horse-radish peroxidase-conjugated anti-GM monoclonal antibody EB-A2 was added to each well of a microtiter plate coated with the same monoclonal antibody EB-A2, followed by addition of 50 μl of the bacterial cell suspension or pretreated sample. After 90 min of incubation at 37°C, the plates were washed five times with washing buffer before 200 μl of buffer containing tetramethylbenzidine solution was added. Then the plates were incubated for another 30 min in the dark at room temperature, followed by the addition of 100 μl of 1.5 N sulfuric acid to stop the reaction. The optical density (OD) was read at 450 and 620 nm. A test sample was considered positive when the OD at 450 nm was higher than the cutoff sample (i.e., 1.0 ng GM). Pretreatment of the R3, R4, and R5 calibration samples and the fecal samples was done by mixing 300 μl of each sample with 100 μl of treatment solution (4% EDTA), and the mixture was subsequently boiled for 5 min. After centrifugation (10,000 × g, 10 min), the supernatant was used for further testing.

All bacterial isolates were tested twice and in duplicate. In vitro reactivity was quantified as ELISA index (EI) correlated to the total protein content of the tested cell suspension. Total protein concentrations were determined with a protein assay kit (Bio-Rad, Richmond, CA), with bovine β-globulin as a standard, after boiling the cell suspensions for 15 min in 1 M NaOH and neutralizing with 1 M HCl. The reactivity of Micrococcus luteus and Propionibacterium freudenreichii, which are known to have a lipomannan in the cell wall (9), served as negative controls. Furthermore, 1 ng/ml of purified GM (22) was added to M. luteus samples with different total protein contents. Fecal samples were tested twice, and in vitro reactivity was expressed as EI per gram of feces.

Cell count. Direct microscopic counts of tested bacterial cell samples were obtained by using duplicate smears of 0.01 ml of a 10-fold dilution spread over 1 cm2 of a glass slide. The smears were heat fixed and gently Gram stained. Ten fields were counted, and the counts were then correlated with the actual sample size.

Preparation and deacylation of cell extracts and cell pellet samples. After growth of B. bifidum subsp. pennsylvanicum on liquid RCM, samples were taken for PA ELISA. Part of the culture (1-ml samples) was centrifuged at 8,000 × g (10 min, room temperature) followed by filtration of the supernatant (0.2 μm). The clear supernatant was used as a cell-free culture fluid. The cell pellet was suspended in 200 μl 0.9% NaCl. The supernatant and suspended-pellet samples were serially diluted and analyzed for PA ELISA reactivity and compared with the total culture sample. Diluted samples were also deacylated by treatment with 0.1 M NaOH in ethanol for 30 min at 37°C as described before (34).

RESULTS

ELISA reactivities of different bacterial samples. B. bifidum subsp. pennsylvanicum bacterial suspensions showed reactivity when tested in the PA ELISA. Reactivity was tested with a range of sample dilutions and showed saturation kinetics (Fig.
Reactivity was determined in the linear part of the graph and was expressed as EI per milligram of protein. Protein concentrations of the undiluted samples ranged from 0.1 to 0.5 mg/ml. Some cell samples reacted so strongly that even a 40-fold dilution gave a positive PA ELISA result (EI/H11022 1.0) as shown for Bifidobacterium lactis (Fig. 2). The reactivity of all tested bacteria is shown in Fig. 3. Most bifidobacteria (from human, animal, or food origin) showed cross-reactivity with the PA ELISA. Reactivities ranged from 6-fold (Bifidobacterium breve) to 600-fold (B. lactis) higher than the controls. Some of these species are known to contain a lipoglycan with more than 7 β-1,5-linked Gal residues (B. bifidum, B. breve, and Bifidobacterium longum; Table 1). Bifidobacterium infantis and Bifidobacterium adolescentis, which do not seem to have β-1,5-

![FIG. 2. PA EI related to the protein concentration of the total bifidobacterial cell sample. ●, B. bifidum subsp. pennsylvaniaic; ■, B. lactis. The detection limit of the assay is indicated as a dotted line.](http://jcm.asm.org/)

<table>
<thead>
<tr>
<th>Bacterial species</th>
<th>Strain or source*</th>
<th>Origin</th>
<th>Characteristics</th>
<th>Reference(s)</th>
</tr>
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<tr>
<td>Bifidobacterial species</td>
<td></td>
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<tr>
<td>B. adolescentis</td>
<td>ATCC 15703</td>
<td>Human intestine</td>
<td>Soluble polysaccharide without β-1,5-Galf</td>
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<td>B. animalis</td>
<td>Wincolve 53</td>
<td>Fermented milk</td>
<td>Lipoglycan</td>
<td></td>
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<td>B. bifidum</td>
<td>CUETM 89/13</td>
<td>Rat feces</td>
<td></td>
<td></td>
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<tr>
<td>B. brev</td>
<td>CUETM 89/27</td>
<td>Breast-fed neonate intestine</td>
<td>Lipoglycan with 11–18 β-1,5-Galf residues</td>
<td>7, 18</td>
</tr>
<tr>
<td>subsp. pennsylvaniaic</td>
<td>Wincolve 1</td>
<td>Human intestine</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>DSM20239</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>B. breve</td>
<td>Wincolve 9</td>
<td>Neonate intestine</td>
<td>Lipoglycan with 11–18 β-1,5-Galf residues</td>
<td>18</td>
</tr>
<tr>
<td></td>
<td>Gavini crohn 16</td>
<td>Neonate intestine</td>
<td></td>
<td></td>
</tr>
<tr>
<td>B. catenulatum</td>
<td>CUETM 15700</td>
<td>Human intestine</td>
<td>Cell wall polysaccharide with 1 internal β-1,5-Galf residue</td>
<td>30</td>
</tr>
<tr>
<td>B. dentium</td>
<td>CUETM 89/20</td>
<td>Human intestine</td>
<td></td>
<td></td>
</tr>
<tr>
<td>B. infantis</td>
<td>ATCC 15697</td>
<td>Infant intestine</td>
<td>Cell wall polysaccharide with 1 internal β-1,3-Galf residue</td>
<td>48</td>
</tr>
<tr>
<td>B. lactis</td>
<td>Wincolve 18</td>
<td>Fermented milk</td>
<td>Lipoglycan</td>
<td></td>
</tr>
<tr>
<td>B. longum</td>
<td>Wincolve 3</td>
<td>Not specified</td>
<td>Lipoglycan with 11–18 β-1,5-Galf residues</td>
<td>18</td>
</tr>
<tr>
<td></td>
<td>Wincolve 16</td>
<td>Not specified</td>
<td></td>
<td></td>
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<tr>
<td>B. pseudocatenulatum</td>
<td>Gavini BS 175</td>
<td>Neonate intestine</td>
<td>Lipoglycan</td>
<td></td>
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<tr>
<td>B. pseudolongum</td>
<td>Gavini BS 82</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>subsp. pseudolongum</td>
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<td>Animal feces</td>
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<td></td>
</tr>
<tr>
<td>subsp. globosum</td>
<td>Gavini RU 224</td>
<td>Animal feces</td>
<td>Lipoglycan</td>
<td></td>
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</table>

* CUETM, Collection de l’Unité d’Ecotoxicologie Microbienne, Institut National de la Santé et de la Recherche Médicale, 59651 Villeneuve d’Ascq, France; DSM, Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Mascheroder Weg 1B, D-38124 Braunschweig, Germany; Gavini, strains received from F. Gavini, INRA, Villeneuve d’Ascq, France; Wincolve, Collection from Wincolve BioIndustries BV, P.O. Box 37239, 1030 AE Amsterdam, The Netherlands; AZN, Collection of the Department of Medical Microbiology, Radboud University Nijmegen Medical Center, P.O. Box 9101, 6500 HB Nijmegen, The Netherlands.

TABLE 1. Origin and characteristics of tested bacterial species and strains

- **Other gram-positive bacteria**
  - Actinomyces viscosus
    - AZN, Human infection
    - Lipoglycan
  - Bacillus subtilis
    - AZN, Human infection
    - LTA
  - Clostridium perfringens
    - ATCC 13124, Not specified
    - LTA
  - Enterococcus faecalis
    - ATCC 29212, Urine
    - LTA
  - Eggerthella lenta
    - ATCC 25559, Rectal tumor
    - Lipoglycan
  - Lactobacillus acidophilus
    - AZN, Human infection
    - LTA
  - Lactobacillus casei
    - AZN, Human infection
    - Unglycosylated LTA
  - Listeria monocytogenes
    - AZN, Human infection
    - LTA with αGal monomeric residues
  - Micrococcus luteus
    - AZN, Human infection
    - Lipomannan
  - Nocardia farcinica
    - AZN, Human infection
    - Lipoglycan
  - Propionibacterium acnes
    - ATCC 6919, Facial acne
    - Lipoglycan
  - Propionibacterium freudenreichii
    - ATCC 6207, Dairy
    - Lipomannan
  - Staphylococcus epidermidis
    - ATCC 12228, Not specified
    - LTA
  - Streptococcus pyogenes
    - ATCC 43202, Not specified
    - Unglycosylated LTA

- **Gram-negative bacteria**
  - Bacteroides fragilis
    - AZN, Human infection
    - LPS
  - Escherichia coli
    - ATCC 25022, Human infection
    - LPS
  - Klebsiella pneumoniae
    - ATCC 27736, Human infection
    - LPS with 1 internal β-1,3-Galf residue
  - Pseudomonas aeruginosa
    - ATCC 10145, Not specified
    - LPS

2). Reactivity was determined in the linear part of the graph and was expressed as EI per milligram of protein. Protein concentrations of the undiluted samples ranged from 0.1 to 0.5 mg/ml. Some cell samples reacted so strongly that even a 40-fold dilution gave a positive PA ELISA result (EI > 1.0) as shown for Bifidobacterium lactis (Fig. 2). The reactivity of all tested bacteria is shown in Fig. 3. Most bifidobacteria (from human, animal, or food origin) showed cross-reactivity with the PA ELISA. Reactivities ranged from 6-fold (Bifidobacterium breve) to 600-fold (B. lactis) higher than the controls. Some of these species are known to contain a lipoglycan with more than 7 β-1,5-linked Gal residues (B. bifidum, B. breve, and Bifidobacterium longum; Table 1). Bifidobacterium infantis and Bifidobacterium adolescentis, which do not seem to have β-1,5-
linked Gal residues in their cell wall, showed low reactivity. Furthermore, other gram-positive bacteria containing LTA or lipoglycan showed low reactivity, including the negative controls containing a lipomannan. Gram-negative bacteria containing lipopolysaccharide (LPS) also reacted negatively. The only nonbifidobacterial species that showed PA ELISA reactivity was *Eubacterium lentum*. This species, which is one of the predominant microorganisms from the human intestine, has been reclassified as *Eggerthella lenta* (19). In contrast to other eubacteria, which belong to the low-G+C group, this bacterium has a high G+C content and by definition a lipoglycan instead of an LTA (31).

**PA ELISA reactivity per gram of feces.** Table 2 shows a calculation of the EI per gram of neonatal feces for the most frequently found bifidobacterial species, i.e., *B. longum*, *B. bifidum*, and *B. breve*. Matsuki et al. (25) studied the distribution of bifidobacterial species in the intestinal tract with 16S rRNA gene-targeted species-specific primers and found about one to three species per neonate (25). The EI was corrected for the dilution factor. This reactivity was then correlated to the

![Cross-reactivity of different bacterial species and strains with the PA ELISA. Reactivity is quantified as EI correlated to the total protein content of the cell suspension tested. Black bars indicate the control samples. Total protein contents of cell samples ranged from 0.1 to 0.5 mg/ml. Control samples include 1 ng/ml GM (clinical cutoff value) added to *M. luteus* samples with different total protein contents.](image)

**TABLE 2. Calculated and measured PA ELISA reactivity per gram of neonatal feces**

<table>
<thead>
<tr>
<th>Tested sample</th>
<th>Occurrence (%)&lt;sup&gt;a&lt;/sup&gt;</th>
<th>EI × dilution factor</th>
<th>No. of cells (10&lt;sup&gt;7&lt;/sup&gt;)&lt;sup&gt;b&lt;/sup&gt;</th>
<th>No. present in feces g&lt;sup&gt;−1&lt;/sup&gt;</th>
<th>EI g feces&lt;sup&gt;−1&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>B. longum</em></td>
<td>37</td>
<td>80.7</td>
<td>1.0</td>
<td>10&lt;sup&gt;8.6–10.1c&lt;/sup&gt;</td>
<td>51,000–102,000</td>
</tr>
<tr>
<td><em>B. bifidum</em></td>
<td>22</td>
<td>10.2</td>
<td>0.7</td>
<td>10&lt;sup&gt;10.5c&lt;/sup&gt;</td>
<td>29,000</td>
</tr>
<tr>
<td><em>B. breve</em></td>
<td>70</td>
<td>4.8</td>
<td>3.8</td>
<td>10&lt;sup&gt;9.5–10.7c&lt;/sup&gt;</td>
<td>4,000–6,000</td>
</tr>
<tr>
<td>Fecal samples from neonates</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>5.6</td>
<td></td>
<td></td>
<td>1.40 × 10&lt;sup&gt;7d&lt;/sup&gt;</td>
<td>1,120</td>
</tr>
<tr>
<td>2</td>
<td>72.0</td>
<td></td>
<td></td>
<td>1.90 × 10&lt;sup&gt;7d&lt;/sup&gt;</td>
<td>14,400</td>
</tr>
<tr>
<td>3</td>
<td>1.0</td>
<td></td>
<td></td>
<td>2.50 × 10&lt;sup&gt;9d&lt;/sup&gt;</td>
<td>200</td>
</tr>
<tr>
<td>4</td>
<td>11.5</td>
<td></td>
<td></td>
<td>1.20 × 10&lt;sup&gt;7d&lt;/sup&gt;</td>
<td>2,300</td>
</tr>
<tr>
<td>5</td>
<td>181.6</td>
<td></td>
<td></td>
<td>3.00 × 10&lt;sup&gt;9d&lt;/sup&gt;</td>
<td>36,320</td>
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<tr>
<td>6</td>
<td>54.8</td>
<td></td>
<td></td>
<td>1.90 × 10&lt;sup&gt;7d&lt;/sup&gt;</td>
<td>10,960</td>
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<tr>
<td>7</td>
<td>125.8</td>
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<td></td>
<td>3.60 × 10&lt;sup&gt;9d&lt;/sup&gt;</td>
<td>28,160</td>
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<td>8</td>
<td>45.1</td>
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<td></td>
<td>2.10 × 10&lt;sup&gt;9d&lt;/sup&gt;</td>
<td>9,020</td>
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<tr>
<td>9</td>
<td>57.6</td>
<td></td>
<td></td>
<td>5.10 × 10&lt;sup&gt;7d&lt;/sup&gt;</td>
<td>11,520</td>
</tr>
</tbody>
</table>

<sup>a</sup> Expressed as % of the total number of fecal samples that were tested, taken from reference 25.

<sup>b</sup> Present in 50 μl undiluted cell sample.

<sup>c</sup> Taken from reference 29.

<sup>d</sup> Determined by FISH analysis.
number of cells present in 50 µl undiluted cell sample. So knowing the reactivity per cell, the reactivity per gram of feces can be calculated by multiplying it with the amount of cells present per gram of feces. Theseecal numbers were taken from the literature but may vary depending on the method used for quantification (15, 29). Table 2 also shows the EI per gram of feces as measured in the nine collected neonatal fecal samples. All fecal samples exhibited PA ELISA reactivity. Furthermore, all fecal samples contained bifidobacterial species as determined by FISH.

**ELISA reactivity of liquid cultures.** The result of the ELISA reactivity of the B. bifidum subsp. pennsylvanicum RCM cultures is shown in Table 3. When theundiluted total cell culture was tested in the PA ELISA, no reactivity was observed. However, 40-fold dilution of the samples gave a positive reaction (EI = 1.553). Part of this reactivity was associated with the cells (25.8%) and another part (37.8%) with secreted products. Blank media showed no reactivity. Deacetylation of the same samples completely removed ELISA reactivity in contrast to deacetylation of GM samples containing 1.5, 3, and 5 ng GM (purified GM was a kind gift from Marc Tabouret, Bio-Rad, Steenvoorde, France) per ml water, which showed a 50% decrease in reactivity upon this treatment (results not shown).

**DISCUSSION**

Most bifidobacteria showed significant reactivity with the PA ELISA in contrast to other gram-positive bacteria containing LTA or lipomannan or gram-negative bacteria containing LPS (Table 1; Fig. 3). ELISA reactivity was clearly shown to be associated with bacterial lipopolysaccharides containing a β-1,5-galactofuranosyl chain. Only a few bifidobacterial lipopolysaccharides have been structurally analyzed, but also other bifidobacterial species seem to contain reactive residues in their cell wall (7, 18, 30). This specific epitope can be detected as a surface antigen, shown by the reactive suspension of cell pellets, but also as an exoantigen, shown by the reactivity of the cell-free supernatant of a liquid culture (Table 3). Secretion of lipopolysaccharides during normal growth was already shown by Op den Camp et al. with B. bifidum subsp. pennsylvanicum (33). The complete removal of PA ELISA reactivity after deacetylation of the supernatant, compared to the 50% decrease with GM, suggests that only the acylated micellar form reacts in the PA ELISA.

When our results were extrapolated to the neonatal host, a calculation was made to estimate the PA ELISA reactivity of feces as potential source of cross-reactivity in the neonate. The bifidobacterial numbers found in feces (15, 29) are very high, and the EI per gram of feces is also high, ranging from 4,000 to 100,000. In order to confirm this calculation, FISH analysis was performed on neonatal fecal samples with a genus-specific probe for bifidobacteria. This 16S rRNA hybridization technique is a fast method to quantitify bifidobacteria in the human gut (21). The presence of bifidobacteria was associated with PA ELISA reactivity of the fecal samples, but reactivity showed some variation, which probably depends on the bifidobacterial species present, as shown by the calculated examples. The results clearly show that the bifidobacterial community in the gut is a significant source of PA ELISA reactivity and that, considering the dilution in the blood volume, the serum concentration could become high enough after transmucosal passage to be detected with the PA ELISA. Furthermore, false-positive PA reactivity in fully breast-fed neonates could not come from galactomannan since breast milk reacts negatively in the PA ELISA (results not shown).

E. lenta is, like Bifidobacterium species, a gram-positive, obligatory anaerobic, non-spore-forming rod and has been found in adult feces at concentrations of 10^6 cells/g, in 4 out of 12 subjects that were tested (detection limit, 10^7 cells/g) (41). Furthermore, the number increased in the intestinal microflora of healthy subjects after administration of antibacterial agents (32). Comparable species have also been found in feces of children at 10^6 cells/g, but the bacteria were not identified to species level (15, 29). So, in addition to Bifidobacterium, this species might also form an important source of PA ELISA reactivity in the intestine. Since E. lenta is present in the fecal flora of adults, it might also be a cause of false PA ELISA serum reactivity in adults. Recently developed species-specific oligonucleotide probes might be helpful with future studies of E. lenta in the human intestine (19, 41).

Positivity of fecal samples has already been shown (1) but was always correlated with consumption of food containing fungal GM from contaminated sources (1, 23). In addition to the host’s own microflora as a source of bifidobacteria, ELISA reactivity of certain food products is more likely to result from bifidobacterial lipoglycan than from fungal GM. Because of their health-promoting effects, bifidobacteria are widely used as probiotics and food additives and are present in fermented foods like milk, olives, sauerkraut, yogurt, butter, and cheese (3). Especially B. lactis and Bifidobacterium animalis are often used, and amounts of 10^7 to 10^8 of bifidobacteria per gram of food can be found (20, 40, 50). As can be seen in Fig. 3, B. lactis (recently reclassified as a subspecies of B. animalis [50]) and B. animalis show high reactivities and could act as a source of ELISA serum reactivity after consumption of food products. Furthermore, bifidobacteria are used as fecal indicator organisms and have been found on meat and meat products and in raw milk (2, 11). Especially meat products are often GM positive (23) but are more likely contaminated with bifidobacteria instead of fungal GM.

The frequent isolation of bifidobacteria from clinical infections in recent years has raised debate whether the bacteria are actually infective (3, 17, 36). However, even if there’s a lack of pathogenicity in immunocompromised patients (3), bifidobacteria can invade the host by bacterial translocation (17). This phenomenon is caused by a diminished intestinal barrier, resulting in the passage of bacteria or bacterial components or products across the mucous membrane and epithelium. Not only intestinal mucosal injury by for instance cytotoxic chemotherapy or an immature intestinal mucosa but also immuno-

**TABLE 3. PA ELISA reactivity of liquid RCM cultures of B. bifidum subsp. pennsylvanicum**

<table>
<thead>
<tr>
<th>Sample</th>
<th>EI^a (n = 2)</th>
<th>Recovery (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total culture</td>
<td>62.1</td>
<td>100</td>
</tr>
<tr>
<td>Cell-free supernatant</td>
<td>23.5</td>
<td>37.8</td>
</tr>
<tr>
<td>Suspended cell pellet</td>
<td>16.0</td>
<td>25.8</td>
</tr>
</tbody>
</table>

*Sample EI was corrected for dilution (20 × to 40 ×), and cell pellet values are also corrected for the five-fold concentration step (from 1,000 µl to 200 µl).
deficiency in the host, overgrowth of intestinal bacteria, and treatment with antibiotics and/or immunosuppressive agents have been shown to promote translocation of intestinal bacteria (i.e., bifidobacteria) (6, 17). The actual translocation of reactive components across the intestinal wall remains to be proven but seems even more likely for an exocellular lipoglycan molecule of 10 kdA. Penicillin treatment of B. bifidum subsp. pennsylvaniaicum resulted in an increase of lipoglycan excretion (33). This was not the result of bacteriolysis, as was also observed for acylated or deacylated LTAs of other bacteria and for other inhibitors of cell wall synthesis (33, 35, 43, 49). Consequently, antibiotic treatment of immunocompromised patients will increase the free lipoglycan pool in the intestine which may easily translocate to the blood.

In the blood, lipoglycan might bind to different components, resulting in a mixture of Galf-containing molecules that react in the PA ELISA, as is the case with GM (27). LTAs and lipoglycans spontaneously bind to mammalian cell membranes. In a study on B. bifidum subsp. pennsylvaniaicum it was shown that its lipoglycan bound reversibly to human colonocytes (8.3 × 10⁵ binding sites/cell) (34). The lipid part of the molecule was shown to be responsible for binding. Furthermore, the lipoglycan of B. bifidum subsp. pennsylvaniaicum exhibited strong binding to the macrophage scavenger receptor, comparable to the LTA of Staphylococcus aureus macrophage scavenger receptor, and for other inhibitors of cell wall synthesis (33, 35, 43, 49).

Bacterial translocation in the intestine is a major cause of false-positive serum samples, a method is needed that specifically recognizes the presence of circulating Gal₃/H₁₁₀₀₃ in the blood, lipoglycan might bind to different components, resulting in a mixture of Galf-containing molecules that react in the PA ELISA, as is the case with GM (27). LTAs and lipoglycans spontaneously bind to mammalian cell membranes. In a study on B. bifidum subsp. pennsylvaniaicum it was shown that its lipoglycan bound reversibly to human colonocytes (8.3 × 10⁵ binding sites/cell) (34). The lipid part of the molecule was shown to be responsible for binding. Furthermore, the lipoglycan of B. bifidum subsp. pennsylvaniaicum exhibited strong binding to the macrophage scavenger receptor, comparable to the LTA of Staphylococcus aureus macrophage scavenger receptor, and for other inhibitors of cell wall synthesis (33, 35, 43, 49).

The Bifidobacterium lipoglycan offers an explanation for the occurrence of a high rate of false PA ELISA reactivity among neonates and infants. Furthermore, together with E. lenta, these bacteria might also be the cause of false-positive reactivity in adult patients.

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REFERENCES