Molecular Evidence of a New Strain of *Ehrlichia canis* from South America

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**Blood samples from dogs with clinical signs compatible with ehrlichiosis were examined for infection of *Ehrlichia canis* using PCR, multiplex real-time PCR, and DNA sequencing analysis. Eleven of 25 samples were positive for a new strain of *E. canis*. This is the first molecular identification of *E. canis* infection in dogs from Peru.**

*Ehrlichia canis* is a rickettsial gram-negative bacterium responsible for canine monocytic ehrlichiosis (CME) in dogs (6, 7, 19) and is mainly transmitted by ticks of the *Rhipicephalus* group (9, 15). *E. canis* has a wide distribution in the world but most frequently occurs in tropical and subtropical regions (2, 8, 11, 12, 21). Recently there have been reports of *E. canis* in South America (1, 6, 16, 17), including the first report of a human case of *E. canis* infection in Venezuela (24). The severity of disease depends on the host's immune status, age, breed, and any current coinfection. Clinical manifestations may vary geographically, but in general the main clinical signs are anemia, thrombocytopenia, and leukopenia (14, 27). Microscopic and serological evaluations have been used in conjunction with clinical signs for diagnosis of CME. Recently, molecular tools, like PCR, have become more sensitive and specific methods to aid in the diagnosis of CME. The specific PCR amplifies a segment of the 16S rRNA gene, and sequences of this gene have been used to identify different strains of *E. canis* (26). Here we report the first genetic characterization of *E. canis* in dogs from Peru.

Twenty-five dogs with clinical signs compatible with *Ehrlichia* sp. infection that were brought by their owners to the Veterinary Teaching Hospital (Universidad de San Marcos, Lima, Peru) from July 2002 to June 2003 were selected for the study. The ages of the animals ranged from 6 months to 11 years, and 80% were males. More than 90% had a history of tick exposure as well as either thrombocytopenia or nonregenerative anemia, fever, anorexia, weight loss, ecchymoses, petechiation, and/or epistaxis.

Serum samples were evaluated for *E. canis* (SNAP 3 DX; IDDEX Laboratories, Westbrook, ME) according to the manufacturer’s protocol. All 25 samples were positive by *E. canis* enzyme-linked immunosorbent assay.

DNA was extracted from EDTA-treated blood samples using a QiaAmp kit (QIAGEN, Inc., Valencia, CA) according to the manufacturer’s protocol. Specific *E. canis* amplification was accomplished using a previously described technique (26) with some modifications. In this PCR, we used the forward primer CANIS (CAATTATTATAGCTCTGGCATTAGGA), reverse primer GA1UR (GAGTTTGCCGGGACTTCTTCT), and a HotStarTaq master mix kit (QIAGEN, Inc., Valencia, CA). Also, the multiplex real-time PCR (MRT-PCR) was used to detect *E. canis*, *E. chaffeensis*, *E. ewingii*, *Anaplasma platys*, and *A. phagocytophilum* (20), with DNA isolated as described above. All PCRs were performed using the Smart Cycler system (Cepheid, Sunnyvale, CA), as described previously. Eleven of the 25 blood samples tested positive for *E. canis* by specific PCR. The same 11 samples were also positive based on the MRT-PCR. None of the samples was positive for *E. chaffeensis*, *E. ewingii*, *A. platys*, or *A. phagocytophilum*.

The 16S rRNA gene of *E. canis* was amplified as two fragments for sequencing. Therefore, two sets of oligonucleotide primers targeting a highly conserved 16S rRNA gene in *E. canis* were used for PCR to obtain the near-full-length 16S rRNA gene. The nucleotide sequences of the forward and reverse primer pairs used for amplification of 16S rRNA were as follows: 15F and 842R, ATCATGGCTCAGAACGAACG; 15F and 842R, ATCATGGCTCAGAACGAACG; 537F and 1442R, CAGCAGCCGCGGTAATACG and GTGACGGGCAGTGTGTACAAG, respectively; and 537F and 1442R, CAGCAGCCGCGGTAATACG and GTGACGGGCAGTGTGTACAAG, respectively (26). The PCR products were purified from the gel (Millipore Corp., Bedford, MA) and cloned with the pGEM-T cloning kit (Promega, Madison, WI). Plasmids were isolated from the clones with the QIAprep Spin miniprep kit (QIAGEN, Inc., Valencia, CA). Plasmids with inserts were tested by specific *E. canis* PCR. Inserted sequences were sequenced using vector primers T7 and SPS6, and a CEQ 8000 genetic analysis system (Beckman Coulter, Fullerton, CA) was used for sequencing. Both the sense and antisense strands of each ampiclon were sequenced, and sequences were aligned and edited manually. The 16S rRNA gene sequences were subjected to BLAST analysis of GenBank.

Sequences were aligned manually using Se-Al (18); the matrix was trimmed slightly on each end and was complete for the included sequences. Two sequences of *Anaplasma* were used as an outgroup. All phylogenetic analyses were conducted using PAUP* v4.0b10 (23), with gaps treated as missing data. Unweighted parsimony analysis was conducted by heuristic

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TABLE 1. Comparison of nucleotide differences among 16S rRNA genes of E. canis from different sources

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* Bullets represent positions conserved relative to the sequence based on the PDE sequence. Slashes (’/’) indicate the sequences were unavailable. Data are presented as modified from those in the study by Unver et al. (24).

b E. canis DNA from a dog in Peru reported in this study.
c E. canis DNA from a dog in Venezuela (24).
d E. canis cDNA from R. sanguineus ticks from Venezuela (24).
e Venezuelan human culture isolate (17).
f E. canis type strain culture isolated from a dog in Oklahoma (4).
g E. canis culture isolated from a dog in Florida (4).
h E. canis culture isolated from a dog in Peru reported in this study.
i E. canis DNA from a dog in The People’s Republic of China (10).
j E. canis DNA from a dog in South Africa (3).
k E. canis DNA from a sheep in Turkey (5).
l DNA from a dog in Spain (2).
m E. ovina DNA from a sheep in Greece.
n E. ovina DNA from a dog in Greece.
o E. ovina DNA from a dog in Spain (2).
p E. ovina DNA from a dog in Thailand.
q DNA from a dog in Okinawa, Japan (22).
r Corrected bases based on the sequence data that were obtained by resequencing (24).

search with 100 random additions (with tree bisection-reconnection branch swapping and the Multrees option on), followed by full heuristic bootstraping to support branches (8a; with 1,000 replicates, each with 10 random additions and tree bisection-reconnection branch swapping). A neighbor-joining analysis was also conducted, with an uncorrected p (uncorrected number of changes between two sequences) distance measure. The aligned sequence data matrix including partial sequences of the 16S rRNA gene (PDE and other sequences from GenBank) was 1,300 bp long. All sequences from Peru were identical (100%) in all samples analyzed and named “PDE,” for Peruvian dog Ehrlichia (GenBank accession no. DQ915970). The PDE sequence was nearly identical (99.9%) to the E. canis VHE, VDE, VTE, Ovina, and Kagoshima sequences (Table 1). Specific differences between PDE and almost all E. canis reported strains in this article were found at nucleotide positions 648, 941, and 1001 (Table 1).

There were 134 variable characters, 100 of which were potentially parsimony informative. Parsimony analysis yielded two most parsimonious trees of 141 steps each, with a consistency index of 0.99. The new strain, PDE, grouped within a well-supported E. canis clade (Fig. 1). However, relationships within E. canis were not well resolved and did not enable any robust inferences regarding patterns of diversification. Indeed, all E. canis sequences and the included sequence of E. ovina were nearly identical (Table 1 and Fig. 1).

Canine ehrlichiosis is a potentially fatal disease, and diagnosis cannot be made based on clinical signs or serological results alone. Clinical signs may vary between different geographical regions, and serology does not differentiate between current infection and previous exposure to Ehrlichia, whereas PCR might indicate an active infection. Therefore, the assessment of clinical manifestations, serologic evaluation, and molecular evidence are all important for accurate diagnosis of ehrlichiosis.

The presence of dogs with E. canis in Peru raises concern that dogs may act as a reservoir of agents of human ehrlichiosis in this region, as purported for Venezuela (24). The possibility that dogs may facilitate transmission of these bacteria to humans increases their zoonotic importance.

The present work is the first to document infection of dogs with E. canis from Peru using molecular methods. It is clear that the new strain reported here, PDE, represents E. canis. While sequencing of the 16S gene enables positive identification of the bacterium, there is too little sequence variation within this evolutionarily conserved coding region to enable
inference of the phylogenetic history of *E. canis* strains. An interesting future investigation would be the exploration of phylogeographic patterns of these strains: sequencing of a non-coding region of DNA for a diversity of *E. canis* samples might yield valuable information on the epidemiology of the disease.

**Nucleotide sequence accession numbers.** The GenBank accession numbers for the 16S rRNA nucleotide sequences of organisms used for comparison in this study are as follows: *E. canis* VDE, AF373613; *E. canis* VTE, AF373614 and AF373615; *E. canis* VHE, AF373612; *E. canis* Venezuela, AF287154; *E. canis* Oklahoma, M73221; *E. canis* Florida, M73226; *E. canis* 611 (Israel), U26740; *E. canis* Gzh982, AF162860; *E. canis* Germishuys, U54805; *E. canis* Gxht67, AF156786; *E. canis* Gdt3, AF156785; *E. canis* G95E10, U96437; *E. canis* okinawa, AF308455; *E. canis* Madison, YA39465; *E. canis* Germishuys, AF536827; *Ehrlichia ovina* Turkey, AF318946; *E. canis* Turkey, YA621071; *E. canis* Greece, EF011110 and EF011111; *E. chaffeensis*, M773222, *E. ewingii*, M73227; *A. platys*, AF156784; *A. phagocytophila*, DQ449948; *Neocricetotis helminthoeca*, U12457; and the new *E. canis* PDE reported here, DQ915970.

**REFERENCES**


