**Nocardia pseudobrasiliensis** as an Emerging Cause of Opportunistic Infection after Allogeneic Hematopoietic Stem Cell Transplantation

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We report the case of a 55-year-old man who exhibited a nodular pneumonia 4 months after an allogeneic hematopoietic stem cell transplantation. Culture of the bronchoalveolar lavage fluid revealed *Nocardia pseudobrasiliensis*. This recently described carbapenem-resistant species should be included in the differential diagnosis of fungal infection in this setting.

**CASE REPORT**

We report the case of a 55-year-old man who was admitted to our unit on 7 May 2008 for recent-onset cough and fever. His main medical history was multiple myeloma, diagnosed in February 2001. Treatment of the myeloma included several chemotherapy avenues, including two autologous stem cell transplantations, before he underwent allogeneic hematopoietic stem cell transplantation (HSCT) while in partial remission. Allogeneic HSCT from a matched, unrelated donor was performed at the end of January 2008 following conditioning with intravenous fludarabine (120 mg/m²), oral busulfan (8 mg/kg of body weight), and anti-thymocyte globulin (ATG; 7.5 mg/kg). Prevention of graft-versus-host disease (GVHD) consisted of cyclosporine and mycophenolate mofetil (MMF). Intron A, an interferon-α inducer, was added to steroids, tacrolimus, and MMF for steroid-resistant acute GVHD. He received posaconazole (200 mg three times a day) for primary prophylaxis of invasive fungal infection, oral penicillin V (2 millions units/day), and monthly aerosolized pentamidine (300 mg). At the time of admission to our department, the myeloma remained in stable partial response.

Four months after allogeneic HSCT, the patient developed cough, dyspnea, and fever (38°C). On examination, he appeared asthenic; his blood pressure was 135/60 mm Hg, pulse beats were 90 per minute, respiratory rate was 20 per minute, and temperature was 38°C. Physical examination was normal.

Laboratory test results were hemoglobin, 9.1 g/dl; polymorphonuclear cells, 1,700/mm³; platelet count, 51,000/mm³; serum creatinine, 142 μmol/liter; C-reactive protein level, 69 mg/liter (normal is below 10 mg/liter); and partial O₂ pressure, 74 mm Hg.

Chest radiography was normal, and a computed tomography (CT) scan of the chest revealed a spiculated nodule of the culmen (Fig. 1A).

Blood cultures were negative. A serum *Aspergillus* galactomannan assay was negative. Because of neutropenia occurring a second time, the patient received piperacillin-tazobactam and amikacin. The neutropenia was related to valganciclovir. Bronchoscopy was macroscopically normal, and bronchoalveolar lavage (BAL) was performed; the fluid recovered contained 200 leukocytes per ml (with 64% macrophages and 34% polymorphonuclear neutrophils), with neither *Pneumocystis jirovecii* nor tumoral cells. Mycologic and viral examinations were negative, but direct microbiological examination of the BAL sample revealed numerous branched Gram-positive filamentous rods that fluoresced upon staining with Uvitex (optical brightener belonging to the stilbene derivatives that fluoresces upon excitation with UV light) (Fig. 2). Bacterial cultures were positive for *Nocardia* spp. Identification at the species level was based on the sequence of the *hsp65* gene, which was amplified and sequenced according to the method of Rodriguez-Nava et al. (18). The 430-bp amplified DNA sequence was 100% homologous with that of the *Nocardia pseudobrasiliensis* reference strain DSM44290 (GenBank sequence accession number AY756530; NCBI BLAST and BiB [http://pbil.univ-lyon1.fr/] phylogenetic tools).

Mycologic and viral cultures of the BAL fluid were negative. Brain magnetic resonance imaging (MRI) results were normal. In vitro susceptibility testing (Table 1) was performed by Etest (AB Biodisk, Solna, Sweden) on Mueller-Hinton plates.
The genus *Nocardia* currently contains more than 80 species that have been characterized by phenotypic and molecular methods (8, 10). Disseminated nocardiosis is a well-known opportunistic infection occurring in conjunction with solid organ transplantation (5), AIDS (13), chronic granulomatous disease (CGD) (7), and prolonged steroid therapy but rarely after allogeneic HSCT (4, 24).

*Nocardia pseudobrasiliensis* is a newly described species that was formerly considered to belong to the *N. brasiliensis* species. Phenotypically, this new taxon is similar to *N. brasiliensis* but hydrolyzes adenine, has nitrate reductase activity less frequently, and displays two specific and early mycolic acid ester peaks in high-performance liquid chromatography (19). Genotypically, this new taxon was confirmed to indeed constitute a new bacterial species on the basis of 16S rRNA gene sequence analysis and DNA-DNA hybridization (19). Furthermore *N. pseudobrasiliensis* exhibits a specific restriction pattern in PCR-restriction fragment length polymorphism analysis of the 65-kDa heat shock protein gene (19, 25).

Wallace et al. (25), in a study of 43 isolates of this new species, described different anatomical sites of infection but reported a strong association with invasive disease, including involvement of the lungs in 30/43 (70%) cases, of the brain and/or meninges in 10/43 (23%), of skin and/or soft tissue in 13/43 (30%), of joints in 3/43 (7%), and disseminated disease in 16/43 (37%). Most of these patients presented with underlying immunosuppression, with 92% (after excluding HIV infection and alcoholism) under corticosteroid therapy.

Ruimy et al. analyzed this taxon by sequencing the gene encoding the small ribosomal subunit (16S RNA) and with DNA-DNA hybridization. They confirmed that this new taxon is indeed a new species and proposed the name *N. pseudobra-

FIG. 1. (A) CT scan of the patient’s chest on admission showing a spiculated macronodule (white arrow) of the culmen. (B) CT scan of the patient’s chest after 2 weeks of antibiotic treatment: the size of the pulmonary nodule was markedly decreased.

FIG. 2. Direct microbiological examination showed branched filaments upon treatment with fluorescent fungal stain (Uvitex).
TABLE 1. Comparative antibiotic susceptibility patterns of *Nocardia brasiliensis* and *Nocardia pseudobrasiliensis*, including the case reported here and others from the literature

<table>
<thead>
<tr>
<th>Drug</th>
<th>MIC breakpoint(s)</th>
<th>This case report</th>
<th>Reference</th>
<th>Result for <em>N. pseudobrasiliensis</em> isolates from reference 4*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(interpretation)</td>
<td>(interpretation)</td>
<td></td>
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</tr>
<tr>
<td>Amikacin</td>
<td>≤0.125, ≥2</td>
<td>ND</td>
<td>24 (S)</td>
<td>S (100) S S</td>
</tr>
<tr>
<td>Amoxicillin-clavulanic acid</td>
<td>≤0.125, ≤16</td>
<td>2 (S)</td>
<td>12 (R)</td>
<td>R (81) ND S</td>
</tr>
<tr>
<td>Ceftriaxone</td>
<td>≤4, ≥0.125</td>
<td>ND</td>
<td>20 (S)</td>
<td>S (69) R S</td>
</tr>
<tr>
<td>Ciprofloxacin</td>
<td>≤4, ≥0.125</td>
<td>0.125 (S)</td>
<td>33 (S)</td>
<td>S (95) S R</td>
</tr>
<tr>
<td>Clarithromycin</td>
<td>≤4, ≥0.125</td>
<td>0.125 (S)</td>
<td>ND</td>
<td>S (91) S R</td>
</tr>
<tr>
<td>Imipenem</td>
<td>≤4, ≥16</td>
<td>&gt;32 (R)</td>
<td>8 (R)</td>
<td>R (&gt;90) R S</td>
</tr>
<tr>
<td>Linezolid</td>
<td>≤4, ≥8</td>
<td>ND</td>
<td>35 (S)</td>
<td>ND S S</td>
</tr>
<tr>
<td>Minocycline</td>
<td>≤1, ≥8</td>
<td>ND</td>
<td>27 (S)</td>
<td>ND R S</td>
</tr>
<tr>
<td>Sulfamethoxazole</td>
<td>≤32, ≥4</td>
<td>ND</td>
<td>6 (S)</td>
<td>S (94) R S</td>
</tr>
<tr>
<td>Tobramycin</td>
<td>≤4, ≥16</td>
<td>ND</td>
<td>32 (S)</td>
<td>ND ND ND</td>
</tr>
</tbody>
</table>

a MIC resistance breakpoints are those from the NCCLS (16); the first value indicates the susceptibility breakpoint, and the second value indicates the resistance breakpoint.

b Inhibition zones, measured in millimeters, were determined by the Mueller-Hinton agar disk diffusion method.

c The percentage of susceptible or resistant strains as determined by the Etest method.

d MICs were determined by the Etest method.

e The table includes results from the reports of Wallace et al. (25), Mongkolrattanathai et al. (15), and Brown-Elliot et al. (4). MIC measurements are in μg/ml.

S, susceptible; R, resistant; ND, not determined.

*N. brasiliensis* (19). Three additional cases have since been reported, including invasive infections; these include ventriculitis in an immunocompetent child (15), pneumonia in a 71-year-old man with a recent history of esophageal and gastric cancer (12), and disseminated infection in a patient with AIDS (3). In contrast, *N. brasiliensis* is mostly isolated from posttrauma cutaneous infection (21) and is responsible for the majority (approximately 80%) of primary cutaneous nocardiosis (4, 14, 20).

The antimicrobial susceptibilities of *Nocardia* spp. are heterogeneous, and six antimicrobial susceptibility patterns have been proposed to classify the most frequently isolated *Nocardia* spp. strains. Importantly, *N. pseudobrasiliensis* does not fit this classification and its antimicrobial susceptibilities differ from those of *N. brasiliensis*. In *Nocardia* spp., several mechanisms may lead to imipenem resistance, including decreased permeability, efflux pump overexpression, and high-level beta-lactamase production (11). In our patient, the observation that EDTA restored susceptibility to imipenem suggests the presence of a metallo-beta-lactamase (17).

To our knowledge, this is the first reported case of *Nocardia pseudobrasiliensis* in an allogeneic HSCT recipient. van Burik et al. (24) retrospectively reported 25 cases of nocardiosis among 6,759 HSCT recipients that included 4,570 allogeneic HSCT recipients. All cases occurred in allogeneic HSCT recipients, with an incidence of 0.54% in this subpopulation. Infections were attributed to *Nocardia asteroides* in 90% of the cases. However, the infections reported in the study were probably caused by other species. According to current taxonomy, there are very few, if any, clinical isolates with genetic similarity to the type strain of *N. asteroides*. Pulmonary involvement (nodule and/or infiltrate) was noticed in 56% of the patients, 44% exhibited skin lesions, and 8% had brain abscesses. Those patients exhibited a high prevalence of GVHD (13/25 [52%]). The median day on which nocardiosis was diagnosed after HSCT was day 210.

Another retrospective study (6) reported 10 cases (corresponding to 1.7% of the allogeneic HSCT recipients) and found 40% coinfection with *Aspergillus fumigatus*. The high mortality rate (70%) was attributed to aspergillosis and GVHD (4/5 patients diagnosed with nocardiosis exhibited GVHD). In that study, cases occurred at a median of 6.2 months after HSCT (ranging from 2 to 18 months) and the *Nocardia* species found were *N. asteroides* (6/10), *N. nova* (3/10), and *N. otitidiscaviarum* (1/10). In these two studies, the beneficial effect of SXT (used as *Pneumocystis jirovecii* prophylaxis) remained uncertain, as nocardiosis occurred in 40% (10/25) of patients receiving this prophylaxis (24).

*Nocardia* spp. infections should be included in the differential diagnosis of invasive fungal infections in allografted patients, and carbapenem-resistant species should be considered when initiating antimicrobial therapy in this setting.

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REFERENCES