Cluster of Macrolide-Resistant *Mycoplasma pneumoniae* Infections in Illinois in 2012


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Macrolide-resistant *Mycoplasma pneumoniae* is an increasing problem worldwide but is not well documented in the United States. We report a cluster of macrolide-resistant *M. pneumoniae* cases among a mother and two daughters.

# CASE REPORTS

**Patient 1.** Patient 1 was an 8-year-old female with antecedent history of asthma who developed fatigue and fever of up to 104.3°F on 18 November 2012. On day 2, she consulted with her primary care physician (PCP); her chest examination was normal, and a nasopharyngeal (NP) specimen collected for influenza was negative. Her PCP recommended symptomatic treatment of the fever and a follow-up visit if there was no improvement in 3 to 4 days; no antibiotics were prescribed at this visit. By day 3 she started having additional symptoms such as cough, headache, nausea, and chest pain. She was re-evaluated by her PCP on day 4; chest radiography showed an infiltrate in the right upper lobe with mild volume loss, and a 5-day course of azithromycin was subsequently prescribed. By day 6, fever persisted despite 3 days of azithromycin treatment; a repeat chest radiograph showed slight improvement in the infiltrate. Azithromycin treatment was continued, and albuterol was administered via nebulizer three times daily. Despite completion of the course of azithromycin, fever persisted, prompting the administration of cefdinir on day 8 and of doxycycline on day 11 because of vomiting. The positive PCR result for *M. pneumoniae* was confirmed from the Quest Diagnostics laboratory to the Centers for Disease Control and Prevention on August 29, 2017 by guest.

**Patient 2.** Patient 2 was a healthy 38-year-old female (mother of patient 1) with no history of smoking or asthma. She developed fever of up to 103°F on 15 December 2012, approximately 1 month after onset of fever in patient 1. On day 2, she presented to her PCP with cough, dizziness, aches, and upper respiratory symptoms. An NP specimen collected at this visit for rapid influenza testing was negative. Given the marginal sensitivity of the rapid influenza test and widespread influenza A H3N2 virus activity in the country at the time, her PCP elected to treat an influenza-like illness with oseltamivir. The fever persisted, and azithromycin was added to the treatment regimen on day 3. She was re-evaluated by her PCP on day 5, and a chest radiograph showed left lower lobe pneumonia. Based on the radiographic findings, azithromycin was stopped and levofloxacin was administered on day 5. Because of wheezing, inhaled albuterol was administered. Although the fever resolved by day 7, cough and wheezing persisted, which prompted the administration of a course of oral corticosteroids by her PCP on day 10. Cough and wheezing gradually improved over the following week, resulting in a total illness course of 17 days.

**Patient 3.** Patient 3 was an otherwise healthy 10-year-old female (sister of patient 1 and daughter of patient 2) with no history of asthma. She presented with fever of up to 104.5°F, dizziness, and headache on 27 December 2012, approximately 2 weeks after the onset of fever in patient 2 and 5 weeks after the onset in patient 1. She had a temperature of 98.8°F in the office on day 2, and ronchi were heard in the left anterior chest upon auscultation. Because of persistent headache, dizziness, and cough, she was re-evaluated in the PCP’s office on day 5. Auscultation revealed inspiratory rales in the left anterior chest, and chest radiography showed lingular pneumonia. Her PCP prescribed a 7-day oral course of cefdinir due to the recent history of pneumonia in the family that did not respond to azithromycin. Blood specimens were obtained for complete blood count (CBC), culture, and erythrocyte sedimentation rate (ESR) analyses, and two NP specimens were collected for viral and *Mycoplasma pneumoniae* PCR. The laboratory results came back on day 9 and were as follows: CBC was normal; ESR was elevated (23 mm/h [normal range, 0 to 20]); blood culture was negative; a respiratory viral PCR panel (Quest Diagnostics) was negative; and *M. pneumoniae* DNA qualitative real-time PCR (Quest Diagnostics) was positive. Because of the positive PCR result for *M. pneumoniae*, she was started on doxycycline treatment. She improved with no further fever, but doxycycline was discontinued on day 11 because of vomiting. The total illness course was 11 days.

NP specimens (*n* = 2) collected from patient 3 were transferred from the Quest Diagnostics laboratory to the Centers for Disease Control and Prevention on 15 December 2012, for confirmatory testing for *M. pneumoniae* by qPCR and culture and for determination of macrolide susceptibility. Isolation was performed using SP4 medium (Remel) as previously described (1). Total nucleic acid was extracted from NP specimens and liquid culture medium using a MagNA Pure Compact instrument with total nucleic acid isolation kit I (Roche Applied Science, Indianapolis, IN) according to the manufacturer’s instructions. *M. pneumoniae*-specific quantitative PCR (qPCR) was performed as previously described (2). Both NP specimens were positive for *M. pneumoniae*; the threshold cycle (*C*<sub>T</sub>) values were approximately 15 for both speci-
M. pneumoniae extracts (data not shown), indicating a relatively large amount of M. pneumoniae nucleic acid present in each specimen. An isolate was obtained from each specimen. Both NP specimens and the corresponding isolates were tested for macrolide resistance using a qPCR assay with Light Upon Extension (LUX) chemistry and high-resolution melt (HRM) analysis to identify single-base mutations in the 23S rRNA gene that confer resistance to macrolide antibiotics (3). Melt patterns consistent with macrolide resistance were observed for both specimens and isolates (Fig. 1). DNA sequencing of the amplicon was performed as previously described (3); sequencing analysis revealed the presence of the A2063G mutation in the 23S rRNA gene, a single-base mutation previously shown to confer macrolide resistance in M. pneumoniae (3) (Table 1).

Primary specimens and isolates were characterized by multilocus variable-number tandem-repeat (VNTR) analysis (MLVA) using previously described methods (4, 5). Both primary specimens and isolates displayed virtually identical MLVA types (5/4/5/7/2), although variation at the Mpn1 locus was identified in one of the primary specimens and the corresponding isolate, a phenomenon that has been reported previously (4). This specimen and isolate displayed two distinct fragment sizes corresponding to two and five repeats at this VNTR locus (Table 2). Molecular typing of the gene encoding the P1 major adhesion protein using qPCR with high-resolution melt analysis identified both isolates as type 1 (Table 2) (6).

M. pneumoniae is a major cause of community-acquired pneumonia among children and adults. Since 2000, reports of the incidence of macrolide-resistant M. pneumoniae isolates have increased worldwide, perhaps due in part to the expanded use of improved diagnostic methods (7). Macrolide-resistant M. pneumoniae has been implicated in individual cases, family clusters, and outbreaks in various settings in the United States and other countries (8–12). The prevalence of resistance among isolates from case series and surveillance studies in the literature ranges from 8% to 27% (13–15). We describe a household cluster of presumed macrolide-resistant M. pneumoniae infections diagnosed 45 days after the onset of symptoms in the index case, including laboratory confirmation in the third case. This household

![FIG 1](http://jcm.asm.org/)

**FIG 1** High-resolution melt (HRM) profiles of M. pneumoniae present in nasopharyngeal specimen collected from patient 3 and in macrolide-resistant and susceptible reference strains. The presence of macrolide-resistant M. pneumoniae is evident from the distinct melting profile. Specimens from patient 3 displayed HRM profiles consistent with that of the macrolide-resistant M. pneumoniae reference strain.

<table>
<thead>
<tr>
<th>TABLE 1</th>
<th>Clinical findings for the three patients</th>
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<tbody>
<tr>
<td>Characteristic</td>
<td>Patient 1</td>
</tr>
<tr>
<td>Age (yrs)</td>
<td>8</td>
</tr>
<tr>
<td>Sex</td>
<td>Female</td>
</tr>
<tr>
<td>Relationship</td>
<td>Younger daughter</td>
</tr>
<tr>
<td>Antecedent medical history</td>
<td>Asthma</td>
</tr>
<tr>
<td>Date of fever onset</td>
<td>18 November 2012</td>
</tr>
<tr>
<td>First antibiotic administered</td>
<td>Azithromycin</td>
</tr>
<tr>
<td>Second antibiotic administered</td>
<td>Cefdinir</td>
</tr>
<tr>
<td>Steroids administered</td>
<td>Yes</td>
</tr>
<tr>
<td>T_max (°F)</td>
<td>104.3</td>
</tr>
<tr>
<td>No. of days of fever prior to antibiotic administration</td>
<td>3</td>
</tr>
<tr>
<td>No. of days of fever after initial antibiotic administration</td>
<td>5</td>
</tr>
<tr>
<td>Total illness course (no. of days)</td>
<td>24</td>
</tr>
<tr>
<td>Chest X-ray result</td>
<td>Dense infiltrate in right upper lobe</td>
</tr>
</tbody>
</table>
The clinical courses of the three patients were similar; all patients had radiological evidence of pneumonia, multiple medical consultations, and multiple courses of antimicrobials. Two of the patients described in this cluster received macrolide treatment with no clinical improvement. The last patient (for whom the diagnosis was laboratory confirmed) did not receive macrolide treatment but was given cefdinir, to which she did not respond. Clinical improvement was noted after the administration of doxycycline. The prolonged duration of symptoms and nonresponse to macrolide therapy suggest that the level of macrolide resistance conferred by the genetic mutation detected here was clinically significant and resulted in treatment failure in these otherwise healthy patients.

Macrolide resistance in *M. pneumoniae* is caused by specific point mutations within the single-copy 23S rRNA gene which inhibit the interaction of macrolides with the large ribosomal subunit, thereby allowing protein synthesis to proceed (22). Sequencing analysis of specimens obtained from patient 3 demonstrated the presence of the A2063G transition in the 23S rRNA gene, a mutation commonly associated with macrolide resistance in *M. pneumoniae* (23, 24). The high burden of *M. pneumoniae* in patient 3, as indicated by the low Ct values in the detection assay, allowed the clear identification of macrolide resistance directly from the primary specimen extract without the need for an isolate.

Assays for the characterization and typing of *M. pneumoniae*, including P1 typing and MLVA, were utilized in this investigation. These assays can help to elucidate the relatedness of strains in outbreaks or clusters such as the one reported here. Unfortunately, specimens were available from only one of the three related cases in this report, precluding any comparison of *M. pneumoniae* data among family members. However, the close family contact, similarity of symptoms, and time courses of the three cases, with onset of the second and third cases occurring within the known incubation period of *M. pneumoniae* after onset of the first case, suggest that all three illnesses were caused by the same strain.

Macrolides such as erythromycin, clarithromycin, and azithromycin are used as first-choice therapeutic agents for treating *M. pneumoniae* infections. This is especially important for children, as other classes of antibiotics with activity against *M. pneumoniae* are not indicated or may cause adverse events in young patients (25). The advantages of clarithromycin and azithromycin over erythromycin include improved oral bioavailability, longer half-life (allowing once- or twice-daily administration), higher tissue concentrations, enhanced antimicrobial activity, and reduced gastrointestinal adverse effects (25). Recent studies have demonstrated that tetracyclines and fluoroquinolones are effective treatments for patients with macrolide-resistant *M. pneumoniae* infections (26). In particular, administration of minocycline was shown to decrease the number of DNA copies found in the nasopharynxes of children with macrolide-resistant *M. pneumoniae* infections (27).

This familial cluster of macrolide-resistant *M. pneumoniae* infections highlights the need for increased awareness among clinicians of circulating macrolide-resistant *M. pneumoniae* strains. Clinicians considering *M. pneumoniae* as part of a differential diagnosis should consider macrolide resistance if there is no clinical improvement after the initiation of first-choice antibiotics. Earlier detection of *M. pneumoniae* and its resistance profile would likely reduce the inappropriate or unnecessary use of antibiotics, the duration and severity of illness, and the spread of infection in the community. Early detection and proper treatment of macrolide-resistant *M. pneumoniae* pneumonia could also be important for prevention of outbreaks in congregate settings. Reliable diagnostic tests for rapid identification of both macrolide-sensitive and macrolide-resistant *M. pneumoniae* strains as well as public health programs to monitor disease are needed to understand the true burden of infections due to this pathogen and to monitor trends in antibiotic resistance.

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We declare that we have no conflicts of interest.

There was no prior presentation of the data presented here.

The findings and conclusions in this report are ours and do not necessarily represent the official position of the Centers for Disease Control and Prevention.

REFERENCES


