Urosepsis can progress toward severe sepsis, septic shock, and, ultimately, death. Rapid antimicrobial susceptibility testing is crucial to decrease mortality and morbidity. This report shows that isothermal microcalorimetry can provide an antibiogram within 7 h with a sensitivity of 95% and specificity of 91% using Vitek-2 system as a reference.

Urine tract infections (UTIs) are the second-most-common type of infection. Patients at risk of urosepsis are those who are elderly, diabetic, immunocompromised, and with obstruction in the urinary tract. Urosepsis might progress to severe sepsis and septic shock, both associated with a high mortality rate ranging between 22% and 76% (1, 2). Treatment of urosepsis includes empirical broad-spectrum antimicrobial therapy and timely de-escalation when antimicrobial susceptibility testing (AST) results become available (3–5). Identification of the pathogen and determination of its susceptibility patterns take at least 48 h on average. Moreover, several methods currently used for AST have additional drawbacks (6, 7). Isothermal microcalorimetry (IMC) that measures metabolic heat production by microbes was recently identified as a promising near-future alternative to conventional methods for AST (6, 7). As urosepsis is virtually always accompanied by UTIs with a high density of uropathogens, urine specimens could be used directly for AST by IMC. Other arguments also advocate such an approach. First, urine is a potent growth medium (8). Second, in 95% of uroseptic cases, urine culture and positive blood culture lead to similar pathogen isolation results (9). And third, polymicrobial infections are rare in bacteremic clinic or performing a clinical study, one needs to determine its sensitivity, specificity, and accuracy. For this study, we used 15 uropathogens (9 Escherichia coli, 3 Enterococcus faecalis, and 3 Enterococcus faecium) previously identified by matrix-assisted laser desorption ionization–time of flight mass spectrometry (MALDI-TOF MS). Susceptibility patterns were obtained by the Vitek-2 system (an automated system for AST). Among the 9 E. coli strains, 3 were sensitive to all antimicrobials tested, 3 were resistant to at least ciprofloxacin, and 3 strains were extended-spectrum-beta-lactamase (ESBL)-producing strains. Among enterococci, all E. faecium strains and one E. faecalis strain were resistant to amoxicillin. Cultures of these strains were diluted in modified artificial urine (12) to obtain an optical density (OD) of 0.1. A 10-μl volume of this dilution was added to a 4-ml microcalorimetric vial prefilled with 3 ml of artificial urine with or without antimicrobial (using the EUCAST guidelines for breakpoint concentration). For AST with E. coli, the following antimicrobials and their respective concentrations were used: ciprofloxacin at 0.5 mg/liter, cotrimoxazole at 2 mg/liter, ceftriaxone at 1 mg/liter, amoxicillin at 8 mg/liter, piperacillin at 8 mg/liter, and ertapenem at 0.5 mg/liter. For enterococci, only amoxicillin at 4 mg/liter and cotrimoxazole at 0.03 mg/liter were tested. After inoculation, the vials were sealed and introduced into the microcalorimeter (TAM48; Waters/TA). The metabolic heat production rate was recorded until the value nearly returned to the baseline. Data were extracted and the maximum growth rate (μ), the lag-phase duration (λ), and the total heat produced (Q) were calculated as described in reference 13. An inhibition index was calculated as follows:

\[
I = \left( \frac{1 - \frac{\mu_2}{\mu_c}}{2} \right) + \left( \frac{Q_2}{Q_c} \right) \times 100
\]

where I is the percent inhibition index. The s and c indices indicate samples and controls, respectively. The results were obtained in 7 h, including 25 to 30 min for sample handling and preparation, 6 h for measurements, and, finally, 30 min for curve fitting and data processing using the R statistical package (14, 15). The calculated inhibitions were concordant with the Vitek-2 system results in 95% of cases. Similarly, IMC results were confirmed by a final OD measurement of the microcalorimetric ampoule content after 48 h (Table 1). For ciprofloxacin and cotrimoxazole, there was a clear difference (Mann-Whitney test; \( P < 0.05 \)) in the inhibition indices of susceptible strains (showing very little growth) and resistant strains (showing rapid growth) (Fig. 1). On the other hand, for beta-lactams (amoxicillin, ceftriaxone, and piperacillin), the inhibition index calculated for susceptible strains using IMC was lower (Fig. 1) but still exhibited a significant difference from the indices determined for resistant strains in all cases (Mann-Whitney test; \( P < 0.05 \)). This is explained by the mode and speed of action of this antimicrobial class. Early in vitro time-kill-curve studies showed that a decrease of CFU or OD after addition of
these antimicrobials in the medium can take between 2 and 4 h at concentrations up to 10× higher than the MIC (16–18). Such delayed action results in an apparently lower level of inhibition. However, for both ceftriaxone and amoxicillin, in most cases the resistant strains had an inhibition index of 0%. Only one amoxicillin-resistant strain had an inhibition index of 6% (Fig. 1). None of the tested strains was resistant to piperacillin, but one would expect that resistant strains would also have an inhibition index very close to 0%. Similarly, none of the tested strains was resistant to ertapenem; however, the very narrow distribution of inhibition indices suggests that clear discrimination would be possible as well (Fig. 1). For AST determination with IMC, the data from ciprofloxacin, cotrimoxazole, ceftriaxone, and amoxicillin showed a sensitivity of 95%, a specificity of 91%, and an accuracy of 93% using the Vitek-2 results as a reference. To avoid bias, the data of piperacillin and ertapenem were not used since only susceptible strains were investigated. These results warrant the use of IMC in larger clinical studies focusing on AST in UTIs and urosepsis. For such studies, additional antibiotics such as pivmecillinam, which has been reported to be valuable against ESBL strains, might be considered. Accurate and reproducible results can be obtained by diluting the original sample in artificial urine to allow bacterial growth (needed to clearly observe an inhibition). The use of arti-

<table>
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<tr>
<th>Test category</th>
<th>IMC = Vitek-2</th>
<th>IMC ≠ Vitek-2</th>
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<tbody>
<tr>
<td>IMC = OD</td>
<td>55 (92)</td>
<td>2 (3)</td>
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<tr>
<td>IMC ≠ OD</td>
<td>2 (3)</td>
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*a* = similar results were obtained by the two methods; † = discordant results were obtained by the two methods. OD values were measured from the microcalorimetric ampule content after 48 h.

![Figure 1](http://jcm.asm.org/)

**FIG 1** (A) Raw data (i.e., heat flow curves) from an *E. coli* isolate resistant to ciprofloxacin and amoxicillin. Note the similarity of the control and ciprofloxacin and amoxicillin heat flow patterns; for the other antimicrobials, the heat flow curves show a rapid decrease. (B, C, D, E, F, and G) Box plots showing the inhibition index obtained by IMC (see equation 1) for antimicrobial-resistant and -susceptible *E. coli*, *E. faecalis*, and *E. faecium* strains previously investigated using the Vitek-2 system. R, resistant; S, susceptible.
ficial urine also improves the reproducibility, as urine composition varies in patients and over time. In addition, artificial urine more closely matches the in vivo situation, as urine exhibits growth characteristics different from those of artificial growth media. Moreover, the concentration of bacteria used as inoculum here was rather low (\(10^5\) CFU/ml) compared to concentrations observed in severe infections (\(10^6\) CFU/ml). An increased inoculum concentration might reduce the measuring time by an additional 2 h (19), thus allowing an antibiogram to be delivered in 5 h.

Moreover, the concentration of bacteria used as inoculum media. Furthermore, the concentration of bacteria used as inoculum for 1 antibiogram with 6 antibiotics to be \$28 (personnel and overhead were not included in the estimate because of large variations between institutions).

REFERENCES


