Probiotic-Associated Aspiration Pneumonia Due to Lactobacillus rhamnosus

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Lactobacilli are low-virulence, commensal organisms of the gastrointestinal and genitourinary tracts and are commonly used as “probiotic supplements.” Herein, we describe an episode of respiratory syncytial virus (RSV) bronchiolitis with bacterial superinfection secondary to administration of Lactobacillus rhamnosus in an 11-month-old female with trisomy 21.

CASE REPORT

An 11-month-old female with trisomy 21 presented with vomiting and fever. The patient had a past medical history significant for esophageal atresia, which was repaired 10 months earlier, subsequently requiring multiple balloon dilations. The patient had dysphagia and was gastrostomy tube (G-tube) dependent. Approximately 1 week prior to admission, the patient was admitted for an esophageal dilation and developed hematemesis, which required intubation and admission to the pediatric intensive care unit (PICU). She was extubated the next day and ultimately discharged home after 4 days of hospitalization given the lack of evidence for esophageal perforation on esophagram and clinical improvement. She had been doing well until 1 day prior to admission, when she had an episode of emesis and developed a fever to 101°F. In the emergency department, a chest X ray was performed and interpreted as concerning for a right-sided infiltrate versus atelectasis. The patient was administered intravenous clindamycin and admitted for presumed aspiration pneumonia. Overnight, the patient remained febrile and became hypoxic. A respiratory pathogen panel (RPP) (BioFire; FilmArray, Salt Lake City, UT) was performed and found to be positive for respiratory syncytial virus (RSV), which subsequently led to a discontinuation of clindamycin. The patient remained febrile for 3 days, and O2 saturations continued to be between 80 and 90%. A repeat chest X ray demonstrated progression of pneumonia, demonstrated by complete opacification secondary to her atelectasis of the right lung. At that time, ceftriaxone was initiated, and a pulmonary consult was sought. During this assessment, she was diagnosed with RSV bronchiolitis and presumed bacterial superinfection, which required an admission to the PICU for further management of her hypoxia and increasing respiratory distress. Following admission to the PICU, a bronchoalveolar lavage (BAL) was performed, and the BAL fluid sample was submitted to microbiology for RPP testing as well as bacterial and fungal culture. The RPP was again positive for RSV, while fungal cultures were negative. The BAL fluid specimen Gram stain showed many white blood cells (WBC), rare ciliated respiratory epithelial cells, rare alveolar macrophages, many red blood cells (RBC), and many Gram-positive rods. The bacterial culture grew >100,000 CFU/ml of Gram-positive rods and <1,000 CFU/ml of mixed respiratory flora. The Gram-positive rod grew both aerobically and anaerobically and was alpha-hemolytic and catalase negative. The organism was identified by matrix-assisted laser desorption ionization—time of flight mass spectrometry (MALDI-TOF MS). Both the Biotype version 3.0 (Bruker Daltonics, Billerica, MA) and the Vitek MS (IVD Knowledge Base v2.0) (bioMérieux, Marcy l’Etoile, France) identified the organism as Lactobacillus rhamnosus. Using gradient diffusion testing performed on blood Mueller-Hinton agar and incubated for 24 h (Etest; bioMérieux, Marcy l’Etoile, France), the isolate was found to be susceptible to penicillin (MIC, 0.5 µg/ml) but resistant to vancomycin (MIC, ≥256 µg/ml) (1). Given these findings, ceftiraxone was discontinued and intravenous ampicillin-sulbactam was administered for 10 days. The patient gradually recovered over a 2-week period. Two endotracheal tube aspirates were collected 1 and 15 days after initiation of ampicillin-sulbactam, which were negative for the presence of Lactobacillus by both direct Gram stain and culture.

During the course of this patient’s illness, it was discovered that she had been taking the Lactobacillus rhamnosus-containing probiotic Culturelle (probiotic strain referred to as “LGG”) (Culturelle, Cromwell, CT) for 3 months prior to her illness. A capsule of the probiotic was obtained and cultured, and the isolated organism was confirmed to be L. rhamnosus by MALDI-TOF MS. Strain typing of the L. rhamnosus isolates was performed using repetitive sequence PCR (REP-PCR) with the primer RW3A as previously described (2, 3). PCR products were resolved on the Agilent 2100 (Agilent, Santa Clara, CA), and the relatedness of the strains was evaluated using Diversilab software (bioMérieux, Durham, NC). The probiotic strain and the patient’s strain were identical, with a similarity index of >99%. A control strain of Lactobacillus rhamnosus isolated by the lab during the same time frame was also analyzed and found to be unrelated to the index patient’s isolate (similarity index of
<70% to the index isolate) (Fig. 1). To our knowledge, this is the first reported case of *Lactobacillus* pneumonia related to supplementations with probiotics.

**Discussion and review of the literature.** We have described a case of probiotic-associated pneumonia secondary to RSV infection in a patient with trisomy 21 and a history of esophageal atresia but who was otherwise thought to be immunocompetent. Herein, we provide both clinical and microbiological evidence that the probiotic strain (LGG) was a cause of disease in this patient. Clinically, the treating physicians had suspicion of bacterial superinfection based on the patient’s deteriorating respiratory status as well as imaging studies. In addition, the patient was managed as though she had a *Lactobacillus* infection, where empirical vancomycin and ceftriaxone were switched to ampicillin-sulbactam in response to isolation of *Lactobacillus* in culture. Laboratory evidence was also consistent with an infection; the specimen’s Gram stain demonstrated inflammatory cells as well as abundant Gram-positive rods. The culture results supported this finding with abundant and predominant growth of *Lactobacillus rhamnosus* and strongly suggested a pathogenic role.

The mechanism by which the patient developed this infection is not clear. We hypothesize that the patient aspirated the probiotic strain while being predisposed to serious bacterial infection as a result of her ongoing RSV infection. Indeed, the patient would later be diagnosed with a tracheoesophageal fistula, further supporting this hypothesis. RSV-infected individuals are at risk for secondary bacterial infection, and those with trisomy 21 are at particular risk for severe RSV infection (4, 5). Despite the patient’s risk factors, *Lactobacillus* pneumonia is very rare. A review of the literature identified only 11 cases of *Lactobacillus* pneumonia (6–14). The rarity of cases makes it difficult to draw conclusions about risk factors for disease, but being immunocompromised (especially having AIDs) appears to be the primary risk factor.

*Lactobacilli* are low-virulence organisms that are commensals of the genitourinary and gastrointestinal tracts and are commonly used as probiotic supplements (15). In the rare instances where *Lactobacillus* causes disease, it is usually in the form of bacteremia in patients who are either immunocompromised or have intravenous catheters (16). This is the first published case of probiotic-associated pneumonia. It is one of only 19 previous cases in which the infecting *Lactobacillus* isolate was confirmed by strain typing to be identical to the probiotic strain being taken by the patient (17–23). Eighteen of these 19 cases were bloodstream infections, with the 19th being a case of liver abscess. Interestingly, 5 of the 19 confirmed cases were pediatric patients, 3 of which had short gut syndrome (22).

To date, a number of other studies have associated probiotic use with infection. The majority of cases are bloodstream infections, but other disease manifestations, such as urinary tract infection and peritonitis, have also been reported (24–28). Seven studies have had a look at the safety of LGG probiotic use at the institutional level and found that its use is safe and does not lead to an increase in infectious complications. Whelan and Meyers conducted a literature review of randomized and nonrandomized trials with the goal of investigating the safety of probiotic use (29). Of the 72 articles that fulfilled their inclusion criteria, they found 20 case reports of adverse events in 32 patients. Of these, the majority of infections were due to *Saccharomyces boulardii*, with only 5 identified cases resulting from LGG, all of which were bloodstream infections. Honeycutt and colleagues conducted a randomized placebo-controlled trial in which they asked whether probiotic use could reduce the rates of nosocomial infection in a pediatric intensive care unit (30). The numbers were small in this study, and they were not able to show that LGG administration reduced infection in this patient population. Importantly though, they did not identify any LGG infections during the study. These findings are contrary to those of Luong et al., who found an increase in LGG disease in their hospital immediately after the hospital formulary switched from *Lactobacillus acidophilus* or *Lactobacillus helveticus* to *Lactobacillus rhamnosus* (31).

In conclusion, we describe the first case of probiotic-associated *L. rhamnosus* pneumonia. Our case is of particular interest because it occurred secondary to a respiratory viral infection. One of the reported benefits of *L. rhamnosus* probiotic use is to help prevent the development of severe respiratory tract infection, especially those due to respiratory viruses, including RSV (32–34). These data provide an important example of how normal nonpathogenic probiotic strains can cause real disease in vulnerable patient populations. With that being said, it is important to point out that while probiotics are frequently consumed, there is an extremely low incidence of infection due to these strains. The purpose of this report is to serve as an example of how extenuating circumstances can render a patient susceptible to probiotic infection.

**REFERENCES**


August 2014 Volume 52 Number 8  jcm.asm.org  3125

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**FIG 1** Strain typing of *Lactobacillus* strains. The dendrogram illustrates the results of repetitive sequence PCR (repPCR). Lane 1, patient BAL strain; lane 2, Culturelle strain; lane 3, unrelated *Lactobacillus rhamnosus* control strain. The solid horizontal line denotes a similarity index of 95%.


