

Triazole Cross-Resistance among *Candida* spp.: Case Report, Occurrence among Bloodstream Isolates, and Implications for Antifungal Therapy

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Received 12 August 2005/Returned for modification 14 October 2005/Accepted 16 November 2005

***Candida* spp. are common causes of bloodstream infections among hospitalized patients. Fluconazole (FLC) remains a first-line therapy for candidemia; and voriconazole (VRC), an expanded-spectrum triazole, was recently approved for the treatment of candidemia in nonneutropenic patients. In vitro studies have suggested that VRC has potent activity against *Candida* spp. with reduced susceptibilities to FLC. We present a case report of invasive candidiasis and candidemia due to a *Candida glabrata* isolate that developed resistance to all currently available triazole antifungals after a course of FLC treatment. This case prompted us to determine the frequency of cross-resistance among bloodstream *Candida* isolates collected during a recent 12-month period at a large, academic medical center. FLC MICs were determined for 125 of 153 isolates (81.7%). Thirty of 125 isolates (24%) were resistant or showed reduced susceptibilities to FLC (MICs ≥ 16 $\mu\text{g/ml}$). When 28 of these 30 isolates were tested for their VRC susceptibilities, 9 (32%) had MICs that were ≥ 2 $\mu\text{g/ml}$. Five of these nine isolates were *C. glabrata*, two isolates were *Candida tropicalis*, one isolate was *Candida albicans*, and one isolate was *Candida parapsilosis*. All five *Candida krusei* isolates tested had VRC MICs ≤ 0.5 $\mu\text{g/ml}$. These data have prompted the introduction of reflexive FLC susceptibility testing of first bloodstream *Candida* isolates at our institution. The case report and our data also suggest that VRC should be avoided as initial therapy in unstable patients with invasive candidiasis, particularly in the setting of prior azole exposure. Studies are needed to define the clinical significance of in vitro resistance to the newer antifungal agents.**

Candida spp. account for 9% of nosocomial bloodstream infections (BSIs), making *Candida* the fourth most common cause of nosocomial BSIs in the United States (23). *Candida glabrata* has emerged as the second most common *Candida* species causing BSIs among hospitalized adults (11, 22). While it is generally regarded as less virulent than *Candida albicans*, *C. glabrata* presents a particular therapeutic challenge in that it either may be intrinsically resistant to triazole antifungals (20) or may acquire resistance during relatively short periods of exposure to these drugs (1, 2). Expanded-spectrum triazoles such as voriconazole (VRC) have been shown to have potent in vitro activities against most *Candida* spp., although the VRC MICs appear to be higher for *C. glabrata* and *Candida krusei* than for other species (4, 10). In a recently published clinical trial, the efficacy of VRC was comparable to that of amphotericin B followed by fluconazole (FLC) (sequential treatment) for the treatment of candidemia in nonneutropenic patients (6). The drug was recently approved by the U.S. Food and Drug Administration for the treatment of candidemia in nonneutropenic patients, which makes the evaluation of the in vitro VRC susceptibility profiles of *Candida* species of particular importance.

In the present study we describe a patient with intra-abdominal and bloodstream infections due to a *C. glabrata* isolate that developed resistance to triazole antifungals after a course of FLC treatment. We document the in vitro resistance of this strain to all available triazole antifungal agents, and we also provide a perspective on this resistance by analyzing in vitro

FLC and VRC susceptibility data for *Candida* isolates recovered from blood cultures over a 12-month period at Johns Hopkins Hospital (JHH). The therapeutic implications of these results are discussed.

CASE REPORT

A 45-year-old woman in her usual state of health approximately 1 year prior to admission was found to have multiple hepatic masses. She was diagnosed with a pancreatic neuroendocrine tumor metastatic to the liver. A pylorus-preserving pancreaticoduodenectomy and radiofrequency ablation of hepatic metastases were performed 42 days prior to the current admission. The patient remained in the surgical intensive care unit (ICU) overnight following the surgery. She did not receive FLC prophylaxis in the ICU.

Thirty-four days prior to the current admission the patient developed a fever, with her temperature increasing to 39.2°C. Her abdominal wound was erythematous. A computed tomographic (CT) scan of the abdomen showed no evidence of abscesses. Blood cultures grew *Enterococcus faecalis*, and vancomycin treatment was begun. She defervesced; but a few days later she was again febrile, and a repeat CT scan revealed new, low-attenuation hepatic lesions compatible with abscesses. A blood culture grew *Escherichia coli* and *E. faecalis*, and gatifloxacin treatment was begun. The patient continued to experience fever and rigors. Ultrasound-guided drainage of the patient's hepatic abscesses was performed, and cultures of the abscess material grew *E. faecalis*, *Escherichia hermannii*, and *C. glabrata*. Blood cultures grew *E. faecalis* and *Lactobacillus* spp. Two catheters were placed percutaneously into the patient's hepatic abscesses for continued drainage, and broad-spectrum

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antibacterials were administered. FLC was begun at 800 mg per day intravenously (i.v.) 21 days prior to the current admission. She underwent manipulations of her existing drainage catheters and placement of a third catheter into an undrained hepatic abscess 18 days prior to the current admission. Cultures of fluid aspirated from this collection grew *E. faecalis*, a coagulase-negative *Staphylococcus* sp., and *C. glabrata*. She was discharged to home with the drainage catheters in place to complete a course of antimicrobial therapy, including FLC.

After approximately 2 weeks at home and 3 days after completing her course of antimicrobial therapy, she was readmitted to the hospital with fever and hypotension. Broad-spectrum antibacterial therapy was begun on admission (day 0); and caspofungin (CAS) therapy was initiated on hospital day +1, with a loading dose of 70 mg i.v., followed by 50 mg i.v. per day. Blood cultures were positive for *Prevotella* spp., and liver abscess drainage cultures grew a coagulase-negative *Staphylococcus* sp. and *Stenotrophomonas maltophilia*. Cultures of specimens collected from the hepatic abscess drains on day +4 grew a coagulase-negative *Staphylococcus* sp., a *Lactobacillus* sp., and *C. glabrata*. Cultures of blood samples collected on days +7 and +9 grew a coagulase-negative *Staphylococcus* sp., and blood samples for culture collected on days +9 and +11 grew *C. glabrata*. On hospital day +11 she was transferred to the surgical ICU with mental status changes. The abscess drainage catheters were changed, and a fourth catheter was inserted on day +12. Cultures of hepatic abscess samples collected on day +12 grew *C. glabrata*, a coagulase-negative *Staphylococcus* sp., and *E. faecalis*. CAS was discontinued, and amphotericin B deoxycholate (AMB) at 0.7 mg/kg of body weight/day was started on day +13, with a dose increase to 1.0 mg/kg/day on day +15. On hospital day +20 she was intubated and placed on vasopressors.

On hospital day +21, AMB was replaced by liposomal amphotericin B (L-AMB) at 5 mg/kg/day, and VRC was added at 6 mg/kg i.v. twice daily for two doses, followed by 4 mg/kg i.v. twice daily. She slowly improved, and by hospital day +27 she was extubated. On day +31 she was transferred out of the ICU. Cultures of blood, urine, respiratory secretions, and cerebrospinal fluid remained unremarkable. Fluid collected from the indwelling, percutaneous hepatic drains grew bacteria and *C. glabrata*. An abdominal CT scan on hospital day +35 showed a decrease in the size of one of the liver abscesses, although a 1-cm lesion suspicious of an abscess was noted in the spleen. She returned to the ICU from day +36 to day +45 because of gastrointestinal bleeding. She again stabilized and was transferred out of the ICU. L-AMB was discontinued on hospital day +52. VRC was continued.

On hospital day +53 she deteriorated, with recurrent hypotension and respiratory distress that required intubation and transfer back to the ICU. A repeat abdominal CT scan showed an increase in the size of one of the hepatic abscesses. Cultures of percutaneous drain fluid grew *Candida albicans*, *C. glabrata*, vancomycin-sensitive *Enterococcus faecium*, a viridans group streptococcus, and a coagulase-negative *Staphylococcus* sp. Volume overload and renal failure ensued, necessitating continuous venovenous hemodialysis. VRC administration was changed from intravenous to enteral in the setting of renal insufficiency. Blood cultures grew a vancomycin-resistant *Enterococcus* sp. Despite continued aggressive intensive care, the

insertion of a new hepatic abscess drain, broad-spectrum antibacterial treatment, and the reinitiation of L-AMB treatment, the patient developed progressive multiorgan system failure. Care was withdrawn, and the patient died on hospital day +69.

MATERIALS AND METHODS

Yeast culture, identification, and storage. Clinical specimens were processed according to the standard procedures of the JHH Clinical Mycology Laboratory. Yeasts were identified to the species level by using germ tube formation (Remel, Lenexa, KS), morphology in cornmeal agar with caffeic acid, phenoloxidase and urease tests, carbohydrate fermentation, and carbohydrate assimilation assays (API 20C AUX; bioMérieux, Marcy l'Etoile, France) when necessary. *Candida* isolates from normally sterile body sites, including blood, were stored in water at 4°C and/or in 5% glycerol at -70°C.

Antifungal susceptibility testing and typing of the *C. glabrata* strain isolated from the case patient. The FLC susceptibility testing requested by the clinical team during the patient's hospitalization was performed by the JHH Clinical Mycology Laboratory using a broth microdilution method consistent with the M27-A2 method (8), except for the use of FLC solution for intravenous injection instead of drug compound. Triazole susceptibility testing of the patient's isolates was also performed retrospectively in the JHH Clinical Mycology Laboratory by using the Sensititre YeastOne system, according to the manufacturer's instructions (Trek Diagnostic Systems, Inc., Cleveland, OH).

Selected *C. glabrata* isolates from the patient were sent by the JHH Clinical Mycology Laboratory to the Fungus Testing Laboratory of the University of Texas Health Science Center (UTHSC) in San Antonio for additional susceptibility testing, including testing for susceptibility to AMB and CAS.

Typing of the patient's *C. glabrata* isolates was performed by pulsed-field gel electrophoresis. DNA was prepared by techniques consistent with published methods (15). Electrophoresis was performed by using the GenePath system and *Candida* parameters (Bio-Rad Laboratories, Hercules, CA). The bands were visualized with the Gel Doc 1000 system (Bio-Rad) by using Quantity One Image Capture software (Bio-Rad) and were analyzed with the Molecular Analyst fingerprinting software (Bio-Rad).

Triazole susceptibility testing of bloodstream *Candida* isolates at JHH. To determine the FLC susceptibility profile for *Candida* isolates causing bloodstream infection at JHH, patients with candidemia between June 2003 and July 2004 (excluding August and September 2003) were identified by using the Johns Hopkins Department of Pathology database. The Johns Hopkins Institutional Review Board reviewed and approved this study of bloodstream isolates as exempt research. One *Candida* isolate per episode of candidemia was selected, except in instances where a patient was infected with more than one *Candida* species. In these cases, given that it was available, one isolate from each species was selected. When possible, an isolate from the patient's first day of candidemia was tested. If an isolate from the first day of candidemia was not available, an isolate from a subsequent day of candidemia was used.

Isolates were passaged at least twice and subcultured on Sabouraud dextrose agar overnight before FLC susceptibility testing was performed by a broth microdilution method (hereafter referred to as the BMD assay) consistent with the M27-A2 method (8), except for the use of FLC solution for intravenous injection instead of drug compound. FLC 96-well plates were prepared by using dilutions of FLC 2-mg/ml solution for intravenous injection (Pfizer Roerig, Inc., New York, NY, and Abbott Laboratories, Abbott Park, IL). Plates stored at -70°C were considered acceptable for use until 6 months following the date of preparation. After inoculation of the wells with the yeast suspension, 100 μ l of the same suspension was used to inoculate Chocolate II Agar plates (Becton Dickinson, Sparks, MD) as growth and purity controls. MIC results were read visually after 48 h of incubation and were considered acceptable for inclusion in the analysis if the colony counts on these plates were between approximately 50 and 250 and if there was no evidence of mold or bacterial contamination.

Isolates with FLC MICs of ≥ 16 μ g/ml in the BMD assay were tested for susceptibilities to FLC, itraconazole (ITC), and VRC by using the Sensititre YeastOne system according to the manufacturer's suggested protocol (Trek Diagnostic Systems, Inc.). MIC results were read visually after 24 to 26 h of incubation and were considered acceptable for inclusion if Sabouraud dextrose agar purity control plates were without contamination and the colony counts on the growth control plate were between 15 and 80.

Reference isolates *C. albicans* ATCC 90028 and ATCC 90029 and quality control strains *C. parapsilosis* ATCC 22019 and *C. krusei* ATCC 6258 were used as controls in the BMD assay. At least two of these organisms were tested for each batch of FLC plates prepared for the BMD assay. *C. parapsilosis* ATCC

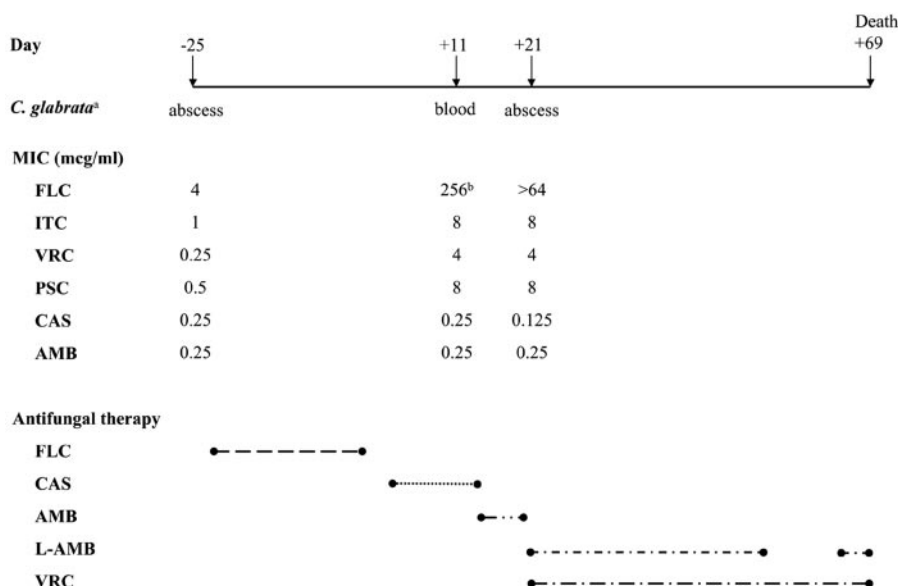


FIG. 1. Chronological presentation of *C. glabrata* isolates from case patient, triazole MICs, and antifungal therapy. Superscript letters: a, Only isolates sent to UTHSC for susceptibility testing are presented; b, Sensititre YeastOne MIC; all other results are from UTHSC.

22019 and *Candida krusei* ATCC 6258 were tested with each batch of Sensititre YeastOne testing that was performed. The MIC results were considered acceptable for inclusion in the analysis if the MICs were within the accepted reference ranges (8; Trek Diagnostic Systems, Inc., YeastOne package insert). The acceptable microdilution FLC MIC range for *C. albicans* ATCC 90028 and ATCC 90029 was considered to be $\leq 0.125 \mu\text{g/ml}$ to $0.5 \mu\text{g/ml}$.

Discrepant isolates were defined as isolates with FLC MIC results that differed by >2 dilutions in the BMD assay and the Sensititre YeastOne assay. For analysis purposes, a BMD assay MIC of $>64 \mu\text{g/ml}$ was considered to be within 2 dilutions of a Sensititre YeastOne MIC of $\geq 32 \mu\text{g/ml}$. Discrepant isolates were retested, and the final FLC MIC was determined by examining all MIC results. For an isolate's MIC results to be considered interpretable, two of three (or, in some cases, three of four) MIC results had to be within 2 dilutions of one another. The final recorded FLC MIC was the highest BMD MIC of those MICs within 2 dilutions of one another. If repeated testing could not resolve the MIC difference, the isolate was excluded.

Definitions. A candidemia episode was defined as the isolation of a *Candida* strain from at least one blood culture. Episodes of candidemia in a single patient were regarded as distinct if they were separated by at least a 30-day period during which time no blood cultures were positive for *Candida*.

Susceptibility to FLC was defined by the use of established interpretive breakpoints (8); specifically, susceptibility was defined by an MIC of $\leq 8 \mu\text{g/ml}$, dose-dependent susceptibility was defined by an MIC of 16 to $32 \mu\text{g/ml}$, and resistance was defined by an MIC of $\geq 64 \mu\text{g/ml}$. Susceptibility to ITC was also defined by the use of established interpretive breakpoints (8): susceptible, an MIC of $\leq 0.125 \mu\text{g/ml}$; dose-dependent susceptible, an MIC of 0.25 to $0.5 \mu\text{g/ml}$; and resistant, an MIC of $\geq 1 \mu\text{g/ml}$. Reduced susceptibility or resistance to VRC and posaconazole (PSC) was defined by an MIC of $\geq 2 \mu\text{g/ml}$ (16, 19).

Statistical analysis. The proportions of *Candida* spp. isolates with reduced susceptibilities to FLC and VRC and with uninterpretable FLC MICs were compared by using Fisher's exact test. The Spearman rank correlation coefficient was used to measure the association between VRC and FLC MICs. In addition, simple linear regression was used to describe the correlation between \log_2 -transformed VRC and FLC MICs. A slope of 1 from this analysis indicates that as the FLC MIC doubles, the VRC MIC doubles.

RESULTS

Case report patient. Seven *C. glabrata* isolates from the case report patient were available: an abscess isolate from day -25, an abscess isolate from day +4, two blood isolates from day +9, two

blood isolates from day +11, and an abscess isolate from day +21. The antifungal susceptibility data from UTHSC for three isolates (day -25, pre-FLC; day +11, post-FLC; and day +21, post-FLC), as well as the antifungal therapy that the patient received, are presented in Fig. 1. The day +11 isolate was sent to UTHSC on day +26 of the patient's hospitalization, with the results reported on day +34; and the day -25 and day +21 isolates were sent retrospectively. These isolates were indistinguishable by pulsed-field gel electrophoresis performed in the JHH Clinical Microbiology Laboratory (data not shown).

The day -25 isolate was susceptible to FLC (MIC, $4 \mu\text{g/ml}$). This isolate was resistant to ITC but was susceptible to both VRC (MIC, $0.25 \mu\text{g/ml}$) and PSC (MIC, $0.5 \mu\text{g/ml}$). The AMB MIC was $0.25 \mu\text{g/ml}$, and the CAS MIC was $0.25 \mu\text{g/ml}$. A bloodstream isolate obtained 2 weeks after FLC therapy, prior to AMB therapy, and during CAS therapy was resistant to FLC (testing was performed at JHH by using the Sensititre YeastOne system [Trek Diagnostic Systems, Inc.]), as well as ITC, VRC, and PSC. The AMB and CAS MICs were unchanged. The hepatic abscess isolate obtained on day +21 also demonstrated in vitro resistance to FLC, ITC, VRC, and PSC. The AMB MIC remained $0.25 \mu\text{g/ml}$, and the CAS MIC was $0.125 \mu\text{g/ml}$.

Species distribution and FLC susceptibilities of *Candida* spp. causing bloodstream infections at JHH. To put this case in perspective, data from candidemia episodes occurring during the 12 months from June 2003 through July 2004 (excluding August and September 2003) at JHH were collected. A total of 147 separate episodes of candidemia occurred in 144 patients, and 153 *Candida* isolates were identified from these 147 episodes of candidemia. Six patients were infected with two different *Candida* species during the same episode of candidemia, and three patients had two separate episodes of candidemia. *C. albicans* was the most commonly isolated species (56 of 153 iso-

TABLE 1. *Candida* spp. recovered from blood cultures over a 12-month period and in vitro FLC susceptibility profiles

<i>Candida</i> sp.	No. of isolates		No. (%) of isolates with the following FLC susceptibility (MIC [μ g/ml]):		
	Total	Tested	Susceptible (≤ 8)	Dose-dependent susceptible (16–32)	Resistant (≥ 64)
<i>C. albicans</i>	56	45	43 (96)	1 (2)	1 (2)
<i>C. glabrata</i>	52	44	25 (57)	11 (25)	8 (18)
<i>C. parapsilosis</i>	22	21	20 (95)	0	1 (5)
<i>C. tropicalis</i>	15	9	6 (67)	1 (11)	2 (22)
<i>C. krusei</i>	7	5	0	3 (60)	2 (40)
<i>C. lusitanae</i>	1	1	1 (100)	0	0
Total	153	125	95 (76)	16 (13)	14 (11)

lates; 37%), followed by *C. glabrata* (52 of 153 isolates; 34%), *C. parapsilosis* (22 of 153 isolates; 14%), *C. tropicalis* (15 of 153 isolates; 9.8%), *C. krusei* (7 of 153 isolates; 5%), and *C. lusitanae* (1 of 153 isolates; 0.7%).

FLC MICs were determined for 125 of 153 isolates (81.7%) (Table 1), most of which (85.6%) were from the first day of candidemia. Seventeen isolates (11%) were not found, and three (2%) did not grow when they were subcultured from storage vials. Three isolates (one isolate each of *C. albicans*, *C. tropicalis*, and *C. parapsilosis*) had low colony counts on the growth control plate, and the purity control plate for one *C. glabrata* isolate had contamination with mold.

Four isolates (3%; three *C. tropicalis* isolates and one *C. albicans* isolate) had MIC discrepancies that could not be resolved with repeated testing. Isolates of *C. tropicalis* were significantly more likely to have uninterpretable FLC MIC results than isolates of other *Candida* species (4 of 13 *C. tropicalis* isolates tested versus 4 of 120 non-*C. tropicalis* isolates tested; $P = 0.003$, Fisher's exact test).

Thirty of 125 isolates (24%) had FLC MICs of $\geq 16 \mu$ g/ml (Table 1); 14 (11%) had FLC MICs of $\geq 64 \mu$ g/ml. Of the 30 isolates with higher FLC MICs, 19 (63%) were *C. glabrata*, 5 (17%) were *C. krusei*, 3 (10%) were *C. tropicalis*, 2 (7%) were *C. albicans*, and 1 (3%) was *C. parapsilosis*. Of the 14 isolates with FLC MICs $\geq 64 \mu$ g/ml, 8 (57%) were *C. glabrata*, 2 were *C. krusei* (14%), 2 were *C. tropicalis* (14%), 1 was *C. albicans* (7%), and 1 was *C. parapsilosis* (7%).

Eleven of 153 *Candida* isolates (7%) were grown from cultures of blood collected in the JHH surgical ICUs, where FLC prophylaxis is routinely administered to patients with expected

ICU stays of ≥ 3 days. *Candida* spp. known to have reduced FLC susceptibilities (*C. glabrata* and *C. krusei*) accounted for a greater proportion of bloodstream *Candida* isolates in the surgical ICUs than in other patient locations (8 of 11 isolates versus 51 of 142 isolates; $P = 0.02$, Fisher's exact test), but reduced FLC susceptibility was not more prevalent among the surgical ICU isolates tested than among isolates from other locations (3 of 7 isolates versus 27 of 118 isolates; $P = 0.36$, Fisher's exact test). In this analysis we were not able to distinguish infections incident to the surgical ICUs from prevalent infections.

ITC and VRC MICs for bloodstream *Candida* isolates with reduced FLC susceptibilities. ITC MICs were determined for 28 of 30 isolates (93%) with FLC MICs $\geq 16 \mu$ g/ml. One *C. tropicalis* isolate tested in the BMD assay did not grow when it was subcultured for ITC and VRC susceptibility testing, and one *C. glabrata* isolate could not be located for ITC and VRC susceptibility testing. Twenty-one of 28 isolates (75%; including 17 of 18 *C. glabrata* isolates tested) were ITC resistant, with MICs $\geq 1 \mu$ g/ml. The remaining seven isolates (four *C. krusei* isolates, one *C. glabrata* isolate, one *C. albicans* isolate, and one *C. parapsilosis* isolate) demonstrated dose-dependent ITC susceptibilities, with MICs of 0.25 to 0.5 μ g/ml.

VRC MICs were also determined for 28 isolates with FLC MICs $\geq 16 \mu$ g/ml (Table 2). Nine of 28 isolates (32%) had VRC MICs $\geq 2 \mu$ g/ml. Five of nine isolates with reduced VRC susceptibilities (56%) were *C. glabrata*. Two were *C. tropicalis*, one was *C. albicans*, and one was *C. parapsilosis*. No isolate of *C. krusei*, a species considered intrinsically FLC resistant, had reduced VRC susceptibility. FLC-resistant *Candida* species other than *C. krusei* were significantly more likely to have VRC MICs $\geq 2 \mu$ g/ml than *C. krusei* (9 of 11 isolates tested versus 0 of 5 isolates tested; $P = 0.005$, Fisher's exact test). All 11 non-*C. krusei*, FLC-resistant isolates had VRC MICs $\geq 1 \mu$ g/ml, 9 of 11 (82%) had VRC MICs $\geq 2 \mu$ g/ml, and 5 of 11 (45%) had VRC MICs $\geq 4 \mu$ g/ml.

VRC MICs were correlated with FLC MICs (Fig. 2) when the *C. krusei* isolates were included (coefficient for \log_2 [FLC MIC] = 1.56; 95% confidence interval = 1.13 to 2.00; Spearman correlation coefficient = 0.83) and when all five *C. krusei* isolates were excluded (coefficient for \log_2 [FLC MIC] = 1.58; 95% confidence interval = 1.14 to 2.01; Spearman correlation coefficient = 0.88). Specifically, we estimated that as the FLC MIC doubles, the VRC MIC will double to quadruple.

TABLE 2. VRC susceptibilities of *Candida* bloodstream isolates with reduced susceptibility to FLC (MIC $\geq 16 \mu$ g/ml)

<i>Candida</i> sp.	No. of isolates		No. (%) of isolates with the following VRC susceptibility (MIC [μ g/ml]):										
	Total	Tested	Susceptible ^a					Resistant ^a					
			≤ 0.06	0.12	0.25	0.5	1.0	2.0	4.0	8.0	16	>16	
<i>C. albicans</i>	2	2	0	1	0	0	0	0	0	0	0	0	1
<i>C. glabrata</i>	19	18	0	2 (11)	3 (17)	5 (28)	3 (17)	3 (17)	2 (11)	0	0	0	0
<i>C. parapsilosis</i>	1	1	0	0	0	0	0	1	0	0	0	0	0
<i>C. tropicalis</i>	3	2	0	0	0	0	0	0	1	0	0	0	1
<i>C. krusei</i>	5	5	0	0	3 (60)	2 (40)	0	0	0	0	0	0	0
Total	30	28	0	3 (11)	6 (21)	7 (25)	3 (11)	4 (14)	3 (11)	0	0	0	2 (7)

^a Total percentages may exceed 100 due to rounding.

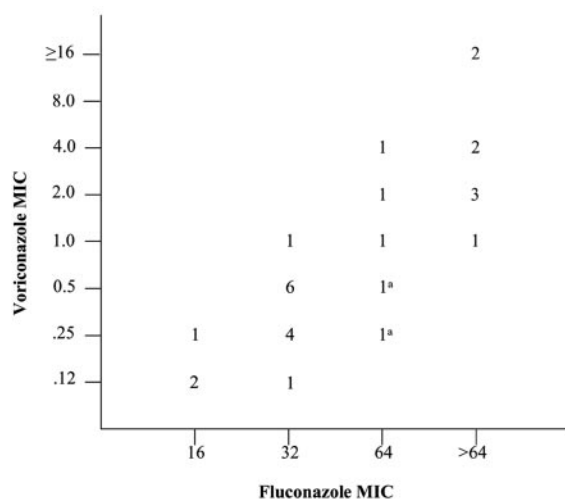


FIG. 2. Distribution of FLC and VRC MICs for *Candida* isolates with FLC MICs ≥ 16 $\mu\text{g/ml}$, including *C. krusei*. VRC and FLC MICs were correlated ($\log_2[\text{VRC MIC}] = -9.18 + 1.56 \log_2[\text{FLC MIC}]$; Spearman correlation coefficient = 0.83). ^a, Isolates of *C. krusei* with an FLC MIC of 64 $\mu\text{g/ml}$ but a VRC MIC of < 1 $\mu\text{g/ml}$.

DISCUSSION

We describe here a patient with a complex illness who developed polymicrobial sepsis, including intra-abdominal and bloodstream infections due to *C. glabrata*. This patient's *C. glabrata* isolate, which was initially susceptible in vitro to FLC, VRC, and PSC, developed cross-resistance to all currently available triazole antifungals after a course of FLC therapy and with no known prior exposure to expanded-spectrum triazoles. This case is similar to another reported recently, involving a patient with prosthetic valve endocarditis due to *C. parapsilosis* (7). Despite infection with an initially FLC- and VRC-susceptible organism, the patient in that report (7) went on to develop recurrent candidemia caused by an apparently identical strain of *C. parapsilosis* that was FLC and VRC resistant. This patient had been treated with a prolonged course of FLC but had never received therapy with VRC (7).

The rapidity with which our patient's *C. glabrata* isolate developed resistance to all currently available triazoles is supported by in vitro work by Borst and colleagues (2). They demonstrated that *C. glabrata* isolates obtained prior to the introduction of FLC developed stable FLC, ITC, and VRC resistance within only 4 days of exposure to FLC (2). They concluded that the rapid acquisition of azole resistance in *C. glabrata* can occur in the absence of prior exposure to these drugs and is associated with the increased expression of ABC transporter genes (2). Another study of azole resistance mechanisms among *C. glabrata* isolates from hematopoietic stem cell transplant recipients receiving FLC prophylaxis showed that MICs doubled, on average, every 31 days (1).

Although some studies have shown that *C. albicans* continues to account for more than half of all cases of candidemia (4, 14, 22), others have demonstrated that *Candida* species other than *C. albicans* in some areas of the United States now account for the majority of *Candida* bloodstream infections (5, 11). In our study, *Candida* species other than *C. albicans* accounted for almost two-thirds of the candidemia episodes, and

C. glabrata alone accounted for just over one-third. Even more striking is the finding that 24% of the evaluable bloodstream *Candida* isolates during this period had in vitro FLC MICs ≥ 16 $\mu\text{g/ml}$, indicating at least reduced susceptibility, and 11% were resistant, with MICs ≥ 64 $\mu\text{g/ml}$. A limitation of our data is that approximately 13% of isolates were not found or did not grow and that susceptibility testing problems were encountered for another 6% of the isolates, which precluded their inclusion in the analyses. Most notably, we had difficulty with a subset of *C. tropicalis* isolates that had disparate MICs on repeated testing, possibly as a result of the trailing phenomenon, which may occur frequently with this species (10).

Our data suggest that with the exception of *C. krusei*, *Candida* isolates with FLC MICs ≥ 64 $\mu\text{g/ml}$ are likely to be resistant in vitro to VRC. Although our study is limited by the small numbers of bloodstream *Candida* isolates from a single institution, our findings support the results obtained by Pfaller and colleagues, who showed that among 12,796 clinical *Candida* isolates, FLC MICs of ≥ 64 $\mu\text{g/ml}$ predicted ravuconazole resistance, defined as a MIC of ≥ 2 $\mu\text{g/ml}$ (16). Another study by Pfaller and colleagues demonstrated that while FLC resistance was uncommon among 3,932 clinical *Candida* isolates (2.9%), approximately half of the FLC-resistant isolates had VRC and PSC MICs of ≥ 2 $\mu\text{g/ml}$ (17). This correlation between in vitro FLC and VRC resistance among *Candida* species highlights the clinical issues raised by our case presentation and may have important implications for antifungal therapy selection in patients with invasive candidiasis.

While FLC resistance and VRC resistance were relatively uncommon in the studies cited above, with approximately 3% of isolates having FLC MICs of ≥ 64 $\mu\text{g/ml}$ and less than 2% of isolates having VRC MICs of ≥ 2 $\mu\text{g/ml}$ (17), resistance to these drugs was more common in our study. At least 9 of the 125 bloodstream isolates in our study (approximately 7%) had VRC MICs of ≥ 2 $\mu\text{g/ml}$, highlighting the geographic differences in azole resistance that exist nationally and internationally (14, 18). Most of the resistant isolates in our study were *C. glabrata*; but a number of *C. albicans*, *C. tropicalis*, and *C. parapsilosis* isolates were also found to have reduced susceptibility or resistance to both drugs. By contrast, it is interesting that the few isolates of *C. krusei* in our study were susceptible to VRC, a finding which has been noted by others (17).

Because we did not collect clinical data, we were unable to ascertain whether extensive triazole experience among patients with candidemia at JHH could explain the prevalence of *C. glabrata* or the degree of reduced triazole susceptibility observed. FLC prophylaxis is routinely administered to JHH surgical ICU patients with an expected ICU stay of at least 3 days (13); it is also administered to liver and hematopoietic stem cell transplant recipients. A retrospective study of candidemia in the JHH surgical ICU before and after the institution of FLC prophylaxis showed that the incidence of candidemia was significantly lower in the period after FLC prophylaxis became routine, although *C. glabrata* was a more common cause of incident and prevalent candidemia in that period than in the period prior to prophylaxis implementation (21). In the current study, *C. glabrata* and *C. krusei* accounted for a greater proportion of bloodstream *Candida* isolates in the surgical ICUs than in other patient locations, but reduced FLC susceptibility was not more common among a small number of surgical ICU

Candida isolates. Overall, only 7% of bloodstream *Candida* sp. isolates and 13% of *C. glabrata* isolates were obtained from cultures of blood drawn in the surgical ICUs. By contrast, 16% of all isolates and 21% of *C. glabrata* isolates were grown from cultures of blood collected in the medical ICU, where FLC prophylaxis is not routinely used. The prevalence of *C. glabrata* and the reduced triazole susceptibilities of isolates at JHH are likely due to a multitude of factors, including triazole exposure in different clinical settings, rather than solely due to a prophylaxis practice applied to a select group of patients in the two surgical ICUs in the hospital.

Our lack of clinical data also prevented us from evaluating the relationship between in vitro susceptibility and clinical outcome. Ostrosky-Zeichner and colleagues reported an overall response rate of 56% in patients with refractory invasive candidiasis who received VRC as salvage therapy (9). This response rate, as the authors note, is similar to the rates seen in studies of other antifungal drugs used as primary therapy, even though many patients had prior azole exposure (9). However, a large proportion of patients in this study (almost 20%) were infected with *C. krusei*, which may have lower VRC MICs than FLC-resistant isolates of other *Candida* spp. The response rate was lower (38%) in patients infected with *C. glabrata*, a species known to acquire azole resistance during therapy (9). Some patients included in that study were participants in studies conducted outside of the United States, where FLC resistance among clinical *Candida* isolates may be less prevalent (3, 14). In vitro susceptibility data from these patients' isolates were not available (9).

Current guidelines suggest that clinicians consider antifungals other than FLC as therapy for invasive candidiasis in patients who have had prior azole exposure and who are unstable or infected with species known to have reduced susceptibilities to FLC (12). Based on the frequency with which triazole resistance was detected among the *Candida* isolates at our institution, reflexive FLC susceptibility testing of patients' initial blood isolates is being instituted. With the wider availability of easy-to-use antifungal susceptibility testing formats that include expanded-spectrum triazoles, further studies evaluating the relationship between in vitro antifungal resistance and clinical outcome are critical to determine the most effective treatment approach for patients with invasive disease. Until such data are available, we believe that it may be prudent to avoid VRC as initial therapy for invasive candidiasis in unstable patients, particularly in the setting of *C. glabrata* infection and prior triazole exposure.

ACKNOWLEDGMENTS

We thank Elizabeth Johnson, Department of Biostatistics, Johns Hopkins Bloomberg School of Public Health, for statistical support.

S.S.M. is supported by a K23 Mentored Patient-Oriented Research Career Development Award from NIAID (grant AI53601) and has received grant support from Pfizer. W.G.M. has received grant support from Pfizer and Merck. Merck provided financial support to W.G.M. for the FLC susceptibility testing of the JHH bloodstream isolates included in this study.

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