

Multiplex Real-Time PCR for Rapid Staphylococcal Cassette Chromosome *mec* Typing[∇]

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Received 15 April 2009/Returned for modification 16 June 2009/Accepted 26 August 2009

Rapid identification and typing of methicillin (meticillin)-resistant *Staphylococcus aureus* (MRSA) is important for understanding the molecular epidemiology and evolution of MRSA and offers many advantages for controlling transmission in both health care and community settings. We developed a rapid molecular beacon real-time PCR (MB-PCR) assay for staphylococcal cassette chromosome *mec* (SCC*mec*) typing. The design of this system is based on the established definition of SCC*mec* types, namely, the combination of the *mec* class complex with the *ccr* allotype. The assay consists of two multiplex panels, the combination of which results in two targets (*mec* class, *ccr*) for each SCC*mec* type. MB-PCR panel I targets *mecA*, *ccrB2*, *mecI*, and the Δ *mecRI-IS1272* junction (*mec* class B); it can definitively identify SCC*mec* types II and IV. MB-PCR panel II detects *ccrC*, *ccrB1*, *ccrB3*, *ccrB4*, and the Δ *mecRI-IS431* junction (*mec* class C2) and is therefore capable of identifying SCC*mec* types I, III, V, and VI in combination with panel I. The method can also detect the recently described novel SCC*mec* type VIII (*ccrAB4* with *mec* class A). Our assay demonstrated 100% concordance when applied to 162 MRSA strains previously characterized by traditional SCC*mec* typing schemes. Four geographically and temporally diverse *S. aureus* collections were also successfully classified by our assay, along with 1,683 clinical isolates comprising both hospital- and community-associated MRSA and methicillin-susceptible *S. aureus* strains. As many as 96 isolates can be classified easily within 3 to 4 h, including DNA isolation, PCR cycling, and analysis. The assay is rapid, robust, sensitive, and cost-effective, allowing for high-throughput SCC*mec* typing of MRSA isolates.

Staphylococcus aureus is a commensal and opportunistic pathogen of significant clinical and veterinary importance, responsible for a wide array of different infections (12, 46). Antibiotic resistance in *S. aureus* poses further challenges, with the global prevalence of methicillin (meticillin)-resistant *Staphylococcus aureus* (MRSA) increasing unabatedly since the isolation of the first strain in 1961 (34). MRSA constitutes a major cause of nosocomial disease worldwide but has also become prevalent in community settings in recent years, further increasing the potential for endemic and zoonotic transmission (6, 54).

MRSA is defined by the presence of a large, heterogeneous genetic element known as “staphylococcal cassette chromosome *mec*” (SCC*mec*), which harbors the gene (*mecA*) coding for methicillin resistance (30, 35). This element is also found in several other staphylococcal species, and it is believed that methicillin-resistant coagulase-negative staphylococci (MR-CoNS) constitute a reservoir for SCC*mec* acquisition by *S. aureus* (36). The SCC*mec* elements discovered thus far are characterized by the presence of (i) terminal inverted and direct repeats, (ii) site-specific cassette chromosome recombi-

nases (*ccr*), (iii) a *mecA* gene complex (*mec*), and (iv) a specific chromosomal insertion site within a conserved open reading frame (*orfX*) of unknown function (5, 29–31, 35, 49, 58, 83). For purposes of molecular epidemiologic analysis, different SCC*mec* “types” are recognized by binary combinations of the *mec* complex and the *ccr* allotype (Fig. 1; see also Table 2), described in detail at <http://www.staphylococcus.net> (Juntendo University). SCC*mec* types can be further classified (subtyped) by differences in regions other than *ccr* and *mec*, designated “joining” (J) regions: J1 (between *ccr* and the right-flanking chromosomal region), J2 (between *mec* and *ccr*), and J3 (between *mec* and *orfX*).

For *S. aureus*, five *ccr* allotypes (*ccrAB1* to *ccrAB4* and *ccrC*) and five classes of *mec* complex (A, B, C1, C2, and E), corresponding to eight SCC*mec* prototypes (I to VIII), have been described to date (5, 29–31, 35, 44, 50, 58, 83). Following the initial description of the SCC*mec* element (30), SCC*mec* types I, II, and III were retrospectively identified in nosocomial MRSA strains isolated in 1961 (NCTC10442), 1982 (N315), and 1985 (85/2082), respectively (29). Subsequently, two different candidates for SCC*mec* IV were proposed by Oliveira et al. (61) (HDE288) and Ma et al. (50) (CA05, 8/6-3P); the former has since been reclassified as SCC*mec* type VI (58). Ito and colleagues later described a strain (WIS) bearing a novel *ccr* element (*ccrC*) and *mec* class (C2); this type was accordingly named SCC*mec* V (31). More recently, various proposals for SCC*mec* type VII have appeared in the literature (5, 11,

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[∇] Published ahead of print on 2 September 2009.

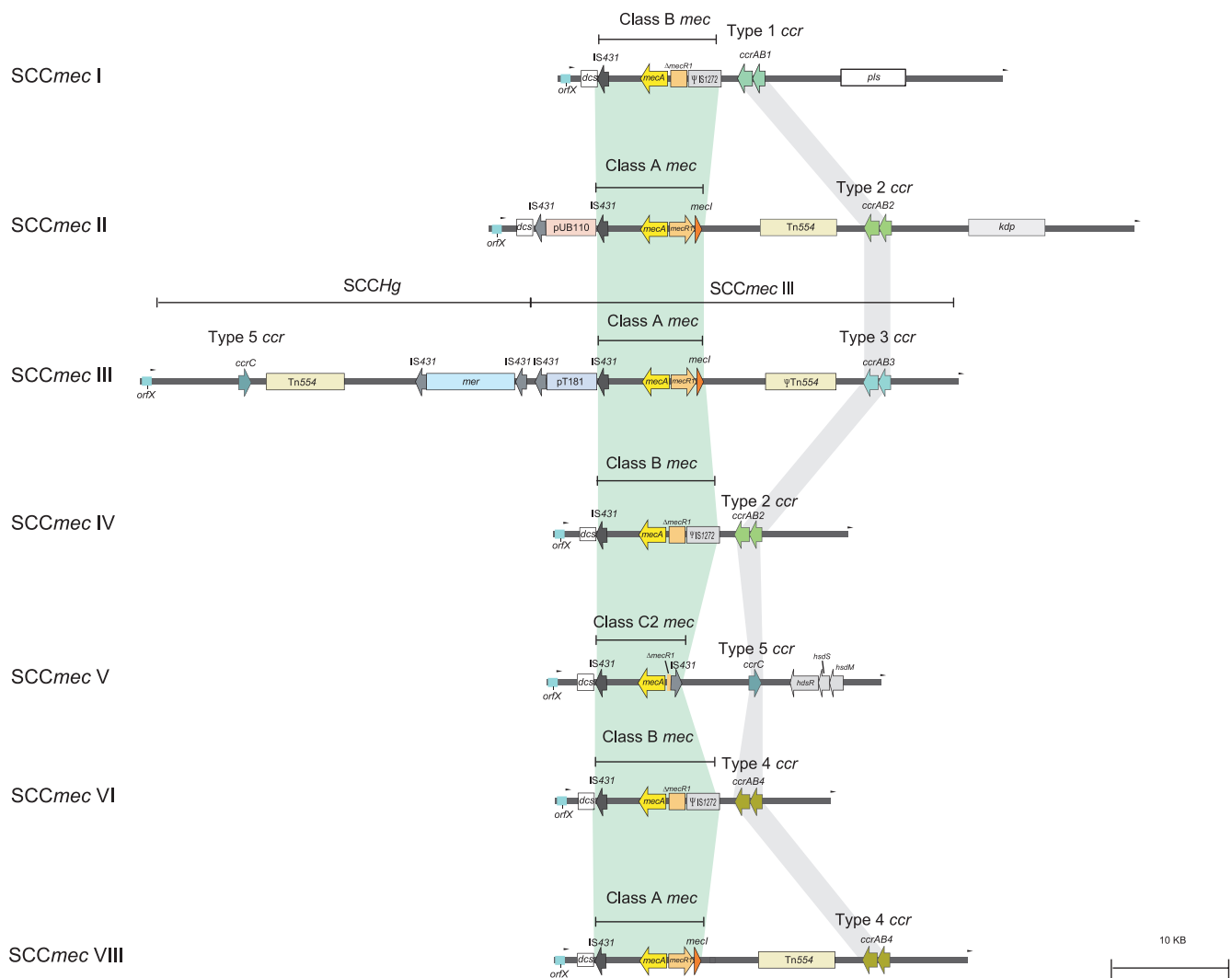


FIG. 1. Schematic representation of SCCmec types I to VI and recently described type VIII in MRSA prototype strains. SCCmec structures are illustrated according to nucleotide sequences deposited in GenBank under the following accession numbers (with strains and SCCmec types given in parentheses): AB033763 (NCTC10442, SCCmec I), D86934 (N315, SCCmec II), AB037671 (85/2082, SCCmec III), AB063172 (CA05, SCCmec IV), AB121219 (WIS, SCCmec V), AF411935 (HDE288, SCCmec VI), and FJ390057 (C10682, SCCmec VIII). The *ccr* and *mec* class complexes grey and utilized for SCCmec type classification are shaded and shown against a grey and green background, respectively; elements outside of these areas constitute “joining” regions (J1, J2, and J3, from right to left).

24); however, the International Working Group on the Classification of Staphylococcal Cassette Chromosome Elements (IWG-SCC) has determined that the combination of *ccrC* and *mec* class C1 (5) should be recognized as the official prototype. Finally, the novel SCCmec type VIII (*ccrAB4* with *mec* class A), in an epidemic Canadian strain, has been described recently by Zhang et al. (83).

Understanding the molecular epidemiology and evolution of MRSA offers many advantages for controlling transmission in both health care and community settings, making rapid identification and typing of MRSA strains an important issue. Along with multilocus sequence typing (MLST), *spa* typing, and pulsed-field gel electrophoresis, SCCmec typing is considered an essential technique for the analysis of MRSA (12, 16). A uniform nomenclature has been proposed (15) in which the multilocus sequence type (ST) and SCCmec type are juxtaposed, e.g., ST5-MRSA-II

(New York/Japan clone) versus ST5-MRSA-IV (Pediatric clone). SCCmec typing is also of potential clinical and epidemiologic interest; SCCmec types I, II, and III, for example, are usually associated with traditional nosocomial strains (hospital-acquired MRSA), whereas types IV, V, and VI are generally characteristic of recently emerging “community” strains (community-acquired MRSA [CA-MRSA]) (10, 29, 31, 79). Lastly, the distribution of SCCmec elements in MR-CoNS is likewise of interest, particularly with regard to the horizontal transmission of SCCmec elements across species boundaries (81, 82).

Various SCCmec typing schemes have been proposed over the past 8 years, most of them based on conventional multiplex PCR (7, 40, 47, 53, 57, 84). The majority of these involve electrophoretic analysis of multiple bands (often greater than 1 kb), thus necessitating DNA isolation methods incompatible with rapid, high-throughput typing. In this study, we describe a

novel procedure for rapid and simple characterization of SCCmec types using multiplex real-time PCR with molecular beacons. Molecular beacons are short oligonucleotide probes possessing a hairpin stem-loop structure, which exhibit fluorescence only when the loop region anneals to a complementary target (77). The presence of the hairpin significantly enhances specificity (76), allowing multiple targets to be distinguished in a single reaction. Real-time PCR assays also offer advantages in that they are highly sensitive, require minimal postamplification analysis, and minimize the possibility of laboratory contamination. The procedure described here uses a two-tiered approach based on the established classification of SCCmec types, i.e., by detecting binary combinations of the *mec* class and the *ccr* allotype, along with detection of the *mecA* gene as a positive control.

MATERIALS AND METHODS

Bacterial strains. Thirteen prototype MRSA strains were used for the derivation and optimization of the real-time PCR SCCmec typing scheme: NCTC10442 (SCCmec type I) (29), N315 (SCCmec type II) (30), BK351 (SCCmec type IIA) (40), JCSC3063 (SCCmec type IIb) (40), 85/2082 (SCCmec type III) (29), CA05 (SCCmec type IVa) (50), 8/6-3P (SCCmec type IVb) (50), JCSC4788 (SCCmec type IVc) (37), JCSC4469 (SCCmec type IVd) (49), AR43/3330.1 (SCCmec type IVE) (68), HAR22 (SCCmec type IVh) (52), WIS (SCCmec type V) (31), and HDE288 (SCCmec type VI) (58). All strains either were kindly provided by T. Ito (Juntendo University, Tokyo, Japan) or came from our own collections (Public Health Research Institute [PHRI], Instituto de Tecnología Química e Biológica, Rockefeller University, Mount Sinai Hospital). Prototype strains for SCCmec types VII (JCSC6082) (5) and VIII (C10682) (83) were not available at the time this study was conducted. Additional strains used for validation, specificity, and applicability are described below.

Molecular beacons and primers. Molecular beacon and oligonucleotide primer sequences are listed in Table 1. Molecular beacon probes were designed according to guidelines presented at <http://www.molecular-beacons.org/>; oligonucleotide primers were designed by Primer3 online software (65). Potential cross-reactivity among beacons and primers was tested using the Autodimer online tool (78). Molecular beacon probes were obtained from Biosearch Technologies (Novato, CA) and purified by high-performance liquid chromatography as described elsewhere (80); oligonucleotide primers were obtained from Integrated DNA Technologies (Coralville, IA). Single-probe (simplex) reactions were conducted with each primer-probe set to evaluate specificity and sensitivity for individual targets. Two panels consisting of four molecular beacon targets each were then designed based on the established classification of SCCmec types by *mec* class and *ccr* allotype; a minimum of two targets were chosen for each SCCmec type, not including *mecA* (see Table 2).

The following considerations were incorporated into the design of individual targets. All *S. aureus* type 1 to 4 *ccrB* and *ccrC* gene sequences available in NCBI GenBank prior to 2009 were downloaded and aligned with ClustalW2 (43) in order to identify divergent and conserved regions. For strains possessing the *ccrAB* allotype, *ccrB* was used as a typing target, since it is more discriminatory than *ccrA* for purposes of SCCmec type assignment (59). The *ccrC* target was designed to detect the *ccrC* alleles found in SCCmec V (*ccrC1*) (accession number AB121219) and SCCmec III strains containing SCCHg (formerly "SCCmercury") (*ccrC3*) (AB037671); of the *ccrC* alleles described to date, this target will also detect *ccrC2* (AY894416), *ccrC6* (EF190467), and *ccrC7* (EF190468), but it may not detect *ccrC4* (U10927), *ccrC5* (AP006716), *ccrC8* (AB353125), or *ccrC9* (NC_007350), due to the presence of one or more mismatches.

For strains possessing a class A *mec* complex (IS431*mecA-mecRI-mecI*), a region in which both *mecI* (intact) and Δ *mecI* (truncated) were conserved was selected for target design. For strains possessing a class B *mec* complex (IS431*mecA-ΔmecRI-ISI272*), the Δ *mecRI-ISI272* junction region was targeted, since it has been reported that the junction (ISI272J) between Δ *mecRI* and ISI272 is conserved in class B *mec* complex sequences (39), whereas ISI272 alone may occur in other regions of the chromosome (75). Similarly, for the class C2 *mec* complex (IS431*mecA-ΔmecRI-IS431*), found in SCCmec V and V_T (8, 31, 74), the region overlapping Δ *mecRI-IS431* was chosen as a target. Finally, a universally conserved region of *mecA* was selected as an internal control for SCCmec, as well as to identify examples of SCC elements lacking *mecA*. Addi-

tional primers and molecular beacons used in this study included (i) a primer-probe set targeting the J1 region of SCCmec subtype IVa and (ii) a primer-probe set specific for the protein A (*spa*) gene of *S. aureus* (70), used to differentiate *S. aureus* from CoNS and to confirm MB-PCR results for methicillin-susceptible *S. aureus* (MSSA) strains.

DNA template isolation. Strains were streaked onto nutrient agar and grown overnight at 37°C. A loopful of bacterial growth was suspended in 100 μ l of sterile distilled water containing 20 μ g/ml lysostaphin (Sigma-Aldrich, St. Louis, MO), incubated at room temperature for 5 min, and then boiled for 10 min. After centrifugation at 10,000 \times g for 5 min, the supernatant was diluted 40-fold into sterile water, and 1 μ l of the diluted supernatant was used as a template in a 10- μ l PCR mixture. For high-throughput applications, the DNA isolation procedure was scaled up using 96-well raised PCR plates as follows. One hundred microliters of the 20- μ g/ml lysostaphin mixture was dispensed into each well, and as many as 96 strains were inoculated using sterile pipette tips. The plate was sealed using PCR strip caps overlaid by an aluminum plate sealer; then it was incubated and boiled as described above. Centrifugation was performed at 10,000 \times g for 5 min in a centrifuge with microtiter plate adapters. Then the supernatant was diluted 40-fold as described above into a new 96-well plate. Diluted DNA lysates were used directly for PCR or stored at -20°C.

PCR amplification. Two multiplex real-time PCR panels were used in this assay (molecular beacon real-time PCR [MB-PCR] panels I and II). Each MB-PCR panel includes four sets of molecular beacons (four dyes), which are accommodated by most commercial real-time PCR instruments. The combination of the two panels results in a minimum of two specific targets for each SCCmec prototype (Table 2); for the purposes of this study, detection of both targets (*mec* class and *ccr* allotype) is considered "definitive" identification. MB-PCR panel I consists of primers and beacons targeting *mecA* (all SCCmec types), *ccrB2* (SCCmec types II and IV), *mecI* (SCCmec types II, III, and VIII), and ISI272J (SCCmec types I, IV, and VI) and is therefore capable of definitively identifying SCCmec types II and IV in a single reaction, as well as presumptively identifying the remaining SCCmec prototypes. MB-PCR panel II is designed to identify *ccrC* (SCCHg [SCCmec type III], SCCmec V), *ccrB1/3* (SCCmec type I or III), *ccrB4* (SCCmec types VI and VIII), and the *mecC2* complex (SCCmec type V), thereby definitively identifying SCCmec type V and confirming SCCmec types I, III, VI, and VIII. In MB-PCR panel II, *ccrB1* and *ccrB3* are detected by different primer sets but share the same molecular beacon (*ccrB1/3*); this hybrid target is a perfect match for *ccrB1* but exhibits a single-nucleotide mismatch relative to *ccrB3* (Table 1).

All amplifications were performed using the Mx4000 multiplex quantitative PCR system (Stratagene, La Jolla, CA). For both MB-PCR panels, each 10- μ l reaction mixture consisted of 0.5 U of AmpliTaq Gold DNA polymerase (Applied Biosystems, Foster City, CA), 4 mM MgCl₂, 250 μ M each deoxynucleoside triphosphate, and 0.4 μ M each primer (Table 1) in 1 \times PCR buffer (Applied Biosystems). Optimized molecular beacon probe concentrations for each panel were as follows: for panel I, 0.2 μ M for *ccrB2*, 0.1 μ M for *mecA* and ISI272J, and 0.05 μ M for *mecI*; for panel II, 0.1 μ M for *ccrC* and *ccrB4* and 0.2 μ M for *ccrB1/3* and *mecC2*. Optimal cycling conditions included an initial denaturation step of 95°C for 10 min, followed by 40 cycles of 95°C for 30 s (denaturation), 50°C for 30 s (annealing), and 72°C for 30 s (extension). The entire PCR can be performed in less than 2 h on the Mx4000 instrument.

MB-PCR sensitivity. The analytical sensitivity of MB-PCR panels I and II was estimated by performing limiting dilution experiments using both purified genomic DNA and bacterial cultures. Six prototype strains (NCTC10442, N315, 85/2082, CA05, WIS, and HDE288) were grown overnight in Trypticase soy broth at 37°C with shaking and were used for both of the following procedures. To estimate sensitivity based on the genomic DNA copy number, 1 ml of each culture was subjected to DNA isolation using a Promega (Madison, WI) Wizard genomic DNA purification kit. Each eluate was diluted serially in sterile double-distilled water (ddH₂O), and the DNA concentration was measured using the Quant-iT PicoGreen dsDNA assay kit from Invitrogen (Molecular Probes, Eugene, OR). One microliter of each dilution was then used for PCR as described above, and limits of detection for each MB-PCR target were calculated using a previously described approach (23). To estimate sensitivity based on bacterial CFU, the remainder of the overnight cultures were pelleted by centrifugation and resuspended in sterile saline to an optical density corresponding to an A₆₀₀ of 1.0. Each suspension was then used to prepare serial 10-fold dilutions using sterile ddH₂O. DNA template isolation was performed on 100 μ l of each dilution using the boiling lysis procedure described above, and 1 μ l of each lysate was used for PCR. The number of CFU per suspension (CFU per milliliter) was estimated by standard plating procedures and was then used to calculate limits of detection for the individual targets in both MB-PCR panels.

TABLE 1. Molecular beacons and primers used in this study

Primer/probe ^a	Target	Sequence (5'–3') ^b	Size of amplicon(bp)	5' fluorescent dye ^c	3' quencher ^c
MB-PCR panel I					
mecA-F	<i>mecA</i>	GCAATACAATCGCACATACATT	148		
mecA-R		CCTGTTTGAGGGTGGATAGC			
mecA-MB		cgcg GATGGCAAAGATATTCAACTAACTA		FAM	BHQ-1
ccrB2-F	Type 2 <i>ccr</i>	CTCATGTTACARATACTTGCG	107		
ccrB2-R		CCTTGATAATAGCCTTCTTGG			
ccrB2-MB		cgcgat TCTCGACAATGTTATTACGTTTCG		HEX	DABCYL
mecI-F	Class A	CGTTATAAGTGTACGAATGGTTTTTG	126		
mecI-R	<i>mec</i>	TCATCTGCAGAATGGGAAGTT			
mecI-MB		cgcg ACTCCAGTCTTTTTGCATTTTGTA		CAL Fluor Red 610	BHQ-2
IS1272J-F	Class B	GAAGCTTTGGCGGATAAAGA	98		
IS1272J-R	<i>mec</i>	GCACTGTCTCGTTTAGACCAATC			
IS1272J-MB		cgcg AGCCATTGCACATGAGTTAA		Quasar 670	BHQ-2
MB-PCR panel II					
ccrC-F	Type 5 <i>ccr</i>	TCCAGTCTATAAAGGSTATGTCAG	124		
ccrC-R		ACTTATAATGGCTTCATGCTTACC			
ccrC-MB		cgcgat GCTGTTACGCGAAGAAGTGGTAA		FAM	DABCYL
ccrB1-F	Type 1 and 3 <i>ccr</i>	ACCACAAACACACTTAAAGATG	150		
ccrB1-R		CAATTTCAAGTATTTGGTCCATAAC			
ccrB3-F		AACACAACGAACACATTGAAAG	130		
ccrB3-R		CGTATTTCTCAATCACATCAGC			
ccrB1/3-MB ^d		cgcgat AAAGGCTCAAAGGTATGTTCTGC		HEX	DABCYL
ccrB4-F	Type 4 <i>ccr</i>	CGAAGTATAGACACTGGAGCGATA	134		
ccrB4-R		GCGACTCTCTTGGCGTTTA			
ccrB4-MB		cgcg GACGAAAAGGACTCAATGAGG		CAL Fluor Red 610	BHQ-2
mecC2-F	Class C2	TCAGTTCATTGCTCACGATATG	174		
mecC2-R	<i>mec</i>	GCCAACGGCTACAGTGATAA			
mecC2-MB		cgcg ACCAAAAGGAGTCTTCTGTATG		Quasar 670	BHQ-2
SCCmec IVa subtyping					
IVa-J1-F	IVa-J1 region	CAGGATATTTTTCAAACCTTCA	131		
IVa-J1-R		TGGAGGACCAAGGATATTCG			
IVa-J1-MB		cgcg CATTAGTGTCCATGTCCGTT		CAL Fluor Red 610	BHQ-2
<i>spa</i> (<i>S. aureus</i>)^e					
<i>spa</i> -F	Protein A	CATTACTTATATCTGGTGGCG	98		
<i>spa</i> -R		GTTAGGCATATTTAAGACTTG			
<i>spa</i> -MB		cgcg TTGTTGAGCTTCATCGTGTG		Quasar 670	BHQ-2

^a F, forward primer; R, reverse primer; MB, molecular beacon.

^b Complementary arm sequences (hairpins) in each molecular beacon are shown in lowercase.

^c FAM, fluorescein; HEX, hexachlorofluorescein; BHQ, Black Hole Quencher; DABCYL, 4-(4'-dimethylaminophenylazo)benzoic acid.

^d The mismatch site between *ccrB1* and *ccrB3* is underlined; a 150-bp amplicon is generated with the *ccrB1* primers, versus a 130-bp amplicon with the *ccrB3* primers.

^e *S. aureus*-specific primer-probe set (70) used to distinguish *S. aureus* from CoNS and to confirm MB-PCR results for MSSA.

TABLE 2. SCCmec classification by MB-PCR panels and targets used in this study

SCCmec type	<i>ccr</i> type and <i>mec</i> class ^a	Typing result by:							
		MB-PCR panel I				MB-PCR panel II			
		<i>mecA</i>	<i>ccrB2</i>	<i>mecI</i>	IS1272J	<i>ccrC</i>	<i>ccrB1/3</i>	<i>ccrB4</i>	<i>mecC2</i>
I	1B	+			+				
II	2A	+	+	+					
III	3A	+		+		(+) ^b	+		
IV	2B	+	+		+				
V	5C2	+				+			+
VI	4B	+			+			+	
VIII ^c	4A	+		+				+	

^a According to the nomenclature system proposed by Chongtrakool et al. (9).

^b (+), *ccrC* is also present in SCCmec III strains containing SCCHg.

^c Novel SCCmec type recently described by Zhang et al. (83).

MB-PCR specificity. The analytical specificities of all primer-probe sets in both MB-PCR panels were tested using a variety of nonstaphylococcal species (50 strains), selected for phylogenetic relatedness as well as nosocomial importance. Gram-positive bacterial strains included *Enterococcus* spp. (*n* = 12), *Streptococcus* spp. (*n* = 8), and *Micrococcus* spp. (*n* = 2). Gram-negative bacterial strains included *Enterobacter* spp. (*n* = 4), *Klebsiella* spp. (*n* = 4), *Acinetobacter* spp. (*n* = 4), *Pseudomonas* spp. (*n* = 4), *Serratia* spp. (*n* = 4), and *Escherichia coli* (*n* = 4). Four fungal strains (*Candida albicans*) were also included. All bacterial strains were obtained from the American Type Culture Collection (ATCC) or else from our own collections; *C. albicans* strains were generously provided by D. Perlin (PHRI).

Validation and application of MB-PCR for SCCmec typing. The MB-PCR scheme was initially optimized using the 13 prototype MRSA strains listed above. Then it was validated using a well-studied MRSA collection (59 strains, including the 13 prototypes) consisting of reference strains previously characterized with respect to genetic background and SCCmec type and chosen to maximize geographical and temporal diversity (Table 3). All strains were from our collections (Instituto de Tecnologia Química e Biológica, Rockefeller University, PHRI), except for the strains from T. Ito mentioned above and five MRSA carriage isolates from swine (*n* = 3) and swine workers (*n* = 2), generously provided by T. Smith of the University of Iowa (71). An additional 68 strains from our own

TABLE 3. Representative collection of MRSA strains used for MB-PCR validation

Isolate ID ^a	Geographic origin	Yr of isolation	Genotypic background			MB-PCR SCCmec typing result ^b										Reference		
			CC	ST	Allelic profile (MLST)	<i>spa</i> -type (Ridom)	<i>spa</i> repeat pattern (eGenomics)	SCCmec type (previous)	<i>mecA</i> (control)	<i>ccrB2</i> (type 2 <i>ccr</i>)	<i>mecI</i> (class A <i>mec</i>)	ISI272J (class B <i>mec</i>)	<i>ccrC</i> (type 5 <i>ccr</i>)	<i>ccrBI/3</i> (type 1/3 <i>ccr</i>)	<i>ccrB4</i> (type 4 <i>ccr</i>)		<i>mecC2</i> (class C2 <i>mec</i>)	SCCmec (this study)
NCTC10442	United Kingdom	1961	8	250	3-3-1-1-4-4-16	1008	YHGFMBQBLO	I	+	-	+	-	-	-	-	-	I	29
HAR21	Finland	2002	5	5	1-4-1-4-12-1-10	1001	TO2MBMDMGMK	I	+	-	+	-	-	-	-	-	I	59
HAR40	Belgium	1995	5	228	1-4-1-4-12-24-29	1001	TO2MBMDMGMK	I	+	-	+	-	-	-	-	-	I	59
E2125	Denmark	1964	8	247	3-3-1-1-2-4-4-16	1051	YHGFMBQBLO	I	+	-	+	-	-	-	-	-	I	61
COL	United Kingdom	1960	8	250	3-3-1-1-4-4-16	1051	YHGFMBQBLO	I	+	-	+	-	-	-	-	-	I	61
UK13136	United Kingdom	1965	8	250	3-3-1-1-4-4-16	12481	YHGFMBQBLO	I	+	-	+	-	-	-	-	-	I	61
BK793	Egypt	1961	8	250	3-3-1-1-4-4-16	1008	YHGFMBQBLO	I	+	-	+	-	-	-	-	-	I	61
HPV107	Portugal	1992	8	247	3-3-1-1-2-4-4-16	1051	YHGFMBQBLO	IA	+	-	+	-	-	-	-	-	I	61
BK1953	United States	1995	8	247	3-3-1-1-2-4-4-16	1051	YHGFMBQBLO	IA	+	-	+	-	-	-	-	-	I	61
PER88	Spain	1992	8	247	3-3-1-1-2-4-4-16	1051	YHGFMBQBLO	IA	+	-	+	-	-	-	-	-	I	61
PER184	Spain	1991	8	247	3-3-1-1-2-4-4-16	1121	YHGFMBQBLO	IA	+	-	+	-	-	-	-	-	I	61
PER_34	Spain	1989	8	250	3-3-1-1-4-4-16	1051	YHGFMBQBLO	IA	+	-	+	-	-	-	-	-	I	61
PL72	Poland	1991	5	5	1-4-1-4-12-1-10	1053	TJMBMDMGMB	I-var	+	-	+	-	-	-	-	-	I	59
POL3	Poland	1992	5	5	1-4-1-4-12-1-10	1053	TJMBMDMGMB	I-var	+	-	+	-	-	-	-	-	I	59
N315	Japan	1982	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	II	+	-	+	-	-	-	-	-	II	30
Mu3	Japan	1997	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	II	+	-	+	-	-	-	-	-	II	25
Mu50	Japan	1997	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	II	+	-	+	-	-	-	-	-	II	41
BK2464	United States	1996	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	II	+	-	+	-	-	-	-	-	II	57
MRSA252	United Kingdom	1997	30	36	2-2-2-2-3-2	1018	WGKAKAO	II	+	-	+	-	-	-	-	-	II	27
BK351	Ireland	1991	8	8	3-3-1-1-4-4-3	1190	YMBQBL0	IIA	+	-	+	-	-	-	-	-	II	40
JCSC3063	Japan	2001-2002	91 ^c	89	1-26-28-18-18-33-30	1375	Y2EJCMBPB	IIb	+	-	+	-	-	-	-	-	II	26
RN7170	Canada	1989	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	IIIe	+	-	+	-	-	-	-	-	II	40
85/2082	New Zealand	1985	8	239	2-3-1-1-4-4-3	1037	WGKAOMO	III	+	-	+	-	-	-	-	-	III	29
ANS46	Australia	1982	8	239	2-3-1-1-4-4-3	1037	WGKAOMO	III	+	-	+	-	-	-	-	-	III	61
R35	United States	1987	8	239	2-3-1-1-4-4-3	1037	WGKAOMO	III	+	-	+	-	-	-	-	-	III	61
LHH1	United States	1994	8	239	2-3-1-1-4-4-3	1037	WGKAOMO	III	+	-	+	-	-	-	-	-	III	61
BK2421	United States	1996	8	239	2-3-1-1-4-4-3	1037	WGKAOMO	III	+	-	+	-	-	-	-	-	III	61
HS1216	Portugal	1997	8	239	2-3-1-1-4-4-3	1037	WGKAOMO	III	+	-	+	-	-	-	-	-	III	61
HU106	Hungary	1996	8	239	2-3-1-1-4-4-3	1538	WGKAOKAOK	III	+	-	+	-	-	-	-	-	III	61
HUSA304	Hungary	1993	8	239	2-3-1-1-4-4-3	^d	WKAKAOKAOMO	III	+	-	+	-	-	-	-	-	III	61
HU25	Brazil	1993	8	239	2-3-1-1-4-4-3	1138	XKAOMO	IIIa	+	-	+	-	-	-	-	-	III	61
HDC2	Portugal	1993	8	239	2-3-1-1-4-4-3	1421	WGKAOM	IIIb	+	-	+	-	-	-	-	-	III	61
DEN907	Denmark	2001	8	239	2-3-1-1-4-4-3	1037	WGKAOMO	NT ^e	+	-	+	-	-	-	-	-	III	59
CA05	United States	1999	45	256	10-14-8-6-10-37-2	1040	A2AKEMBKB	IVa	+	-	+	-	-	-	-	-	IV	50
MW2	United States	1998	1	1	1-1-1-1-1-1	1128	UIJFKPPE	IVa	+	-	+	-	-	-	-	-	IV	4
HSA74	Portugal	1993	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	IVa	+	-	+	-	-	-	-	-	IV	58
FFP311	Portugal	1996	5	5	1-4-1-4-12-1-10	1311	TJMBMDMGMK	IVa	+	-	+	-	-	-	-	-	IV	58
FFR3757	United States	2003	8	8	3-3-1-1-4-4-3	1008	YHGFMBQBLO	IVa	+	-	+	-	-	-	-	-	IV	13
8/6-3P	United States	1996	8	770	1-3-1-1-4-4-3	1008	YHGFMBQBLO	IVb	+	-	+	-	-	-	-	-	IV	50
JCSC4788	Japan	2001	8	451	3-4-1-4-2-4-4-3	11852	YGFMBQBLO	IVc	+	-	+	-	-	-	-	-	IV	37
ARG9	Argentina	1996	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	IVc	+	-	+	-	-	-	-	-	IV	58
Q2314	United States	1996	5	5	1-4-1-4-12-1-10	1088	TJMBMDMGMK	IVc	+	-	+	-	-	-	-	-	IV	1
JCSC4469	Japan	1982	5	5	1-4-1-4-12-1-10	1002	TJMBMDMGMK	IVd	+	-	+	-	-	-	-	-	IV	49
AR43/3330.1	Ireland	1988-2002	8	8	3-3-1-1-4-4-3	1190	YMBQBL0	IVe	+	-	+	-	-	-	-	-	IV	68
HAR22	Finland	2002	22	22	7-6-1-5-8-8-6	1022	TJEINF2MNF2MO	IVh	+	-	+	-	-	-	-	-	IV	52
HAR36	Greece	2002	8	254	3-12-1-1-4-4-3	1009	YGFMBQBLO	IVh	+	-	+	-	-	-	-	-	IV	52
GRE120	Greece	1993	8	8	3-3-1-1-4-4-3	1036	YGFMBQBLO	IV	+	-	+	-	-	-	-	-	IV	2
BARG II 17	United States	1996	8	8	3-3-1-1-4-4-3	1064	YHGFMBQBLO	IV	+	-	+	-	-	-	-	-	IV	61
WBG10049	Australia	1999	30	30	2-2-2-2-6-3-2	1019	XKAKAOMO	IV	+	-	+	-	-	-	-	-	IV	64
COB3	Colombia	1996	5	5	1-4-1-4-12-1-10	11107	TMDMGK	IV ^{NT}	+	-	+	-	-	-	-	-	IV	61
WIS	Australia	1996	45	45	10-14-8-6-10-37-2	1123	A2AKBEKKB	V	+	-	+	-	-	-	-	-	V	31

HT0826	France	2003	152	377	46-75-49-50-13-68-60	t355	UJ2GMKKPNSG	V	+	-	-	+	-	+	V	53
SW31.1	United States	2008	398	398	3-35-19-2-20-26-39	t034	XKAOAOBQ	V	+	-	-	+	-	+	V	71
SW62.1	United States	2008	398	398	3-35-19-2-20-26-39	t034	XKAOAOBQ	V	+	-	-	+	-	+	V	71
SW181.1	United States	2008	398	398	3-35-19-2-20-26-39	t034	XKAOAOBQ	V	+	-	-	+	-	+	V	71
HU01010T	United States	2008	398	398	3-35-19-2-20-26-39	t034	XKAOAOBQ	V	+	-	-	+	-	+	V	71
HU01010IN	United States	2008	398	398	3-35-19-2-20-26-39	t034	XKAOAOBQ	V	+	-	-	+	-	+	V	71
HDE288	Portugal	1996	5	5	1-4-1-4-12-1-10	t311	TJMBDMGMK	VI	+	-	-	-	+	+	VI	58
IPO92	Portugal	2001	5	5	1-4-1-4-12-1-10	t311	TJMBDMGMK	VI	+	-	-	-	+	+	VI	58

^a Prototype strains used to optimize the MB-PCR assay are in boldface.
^b +, detected; -, not detected.
^c ST89 is currently assigned to CC509 by eBURST, version 3 (17).
^d The spa repeat pattern WKAKAOKAOMQ is not recognized by the Ridom SpaServer as of this writing.
^e NT, nontypeable.

collections, previously SCCmec typed by conventional multiplex PCR (40), were also tested using our MB-PCR assay, and the results were compared with those of Kondo et al. (40) (Table 4). A blinded collection of clinical MRSA strains from Canada (n = 35), selected to include diverse SCCmec types and subtypes previously characterized by traditional SCCmec typing methods (29, 31, 56, 57) and antibiotic susceptibility testing, was also tested (Table 4).

The applicability of the MB-PCR scheme was investigated further by performing SCCmec typing on a variety of clinical *S. aureus* isolates (Table 4), including (i) 66 MRSA and 135 MSSA strains (range, NRS1 to NRS407) from the collection of the Network on Antimicrobial Resistance in *Staphylococcus aureus* (NARSA), representing both geographically and temporally diverse isolates for which the genetic background has been determined by *spa* typing and MLST (see below); (ii) a well-characterized collection of historical MRSA isolates (n = 270) from a 1996 study (Bacterial Antibiotic Resistance Group [BARG] study) encompassing 12 New York City (NYC) hospitals (62); (iii) a large collection (n = 639) of isolates from community-associated MRSA infections, collected from 2004 to 2006 in outpatient settings throughout the NYC area; and (iv) a similarly sized collection (n = 573) of carriage and invasive isolates collected from 2007 to 2008 in several NYC area hospitals.

Lastly, our MB-PCR assay was tentatively used to type 71 staphylococcal strains representing the following species: *Staphylococcus aureus* subsp. *anaerobius* (n = 1), *S. capitis* (n = 6), *S. epidermidis* (n = 25), *S. gallinarum* (n = 1), *S. haemolyticus* (n = 7), *S. hominis* (n = 4), *S. lentus* (n = 2), *S. lugdunensis* (n = 4), *S. pseudintermedius* (n = 2), *S. saprophyticus* (n = 5), *S. sciuri* (n = 2), *S. simulans* (n = 4), *S. succinus* (n = 1), *S. xylosus* (n = 5), and *S. warneri* (n = 2) (Table 5). Staphylococcal strains were obtained from ATCC, NARSA, and our own collections; one *S. lentus* strain was provided by T. Smith of the University of Iowa.

Molecular typing. All *S. aureus* isolates used in the study were characterized genotypically by *spa* typing using eGenomics software, as described previously (51, 67); Ridom *spa* types were subsequently assigned using the *spa* typing website (<http://www.spaserver.ridom.de/>) developed by Ridom GmbH and curated by SeqNet.org (<http://www.SeqNet.org/>) (22). MLST was performed on all NARSA strains (data not shown) as described previously (14); MLST data for all other strains in Table 3 were obtained directly from the literature. For all strains in Table 4, the genotypic background is given in terms of the MLST “clonal complex” (CC), as determined by eBURST analysis (17); except for the NARSA strains, all CCs in Table 4 were inferred by *spa* repeat pattern analysis (51, 73).

DNA sequencing. During the development of this assay, four isolates with the combination of *ccrAB4* and the class A *mec* complex were found. The entire SCCmec region of one isolate (BK20781) was amplified using four sets of primers ranging from the right chromosomal junction to *orfX* (Table 6) and was then sequenced by primer walking.

Nucleotide sequence accession number. The sequence of the SCCmec region of BK20781 was submitted to GenBank as accession number FJ670542.

RESULTS

MB-PCR targets. In order to design the real-time PCR SCCmec typing platform, an extensive sequence similarity search was conducted using NCBI BLAST (3), followed by comprehensive alignment of all the *S. aureus* SCCmec sequences available in GenBank prior to 2009. For each SCCmec type available during the design phase (types I to VI), two loci were selected for target design, based on the combination of *mec* and *ccr* complex. A conserved region of *mecA* was selected as an internal control for methicillin resistance (Table 1). Two four-color multiplex PCR panels were then optimized and validated for the differentiation of SCCmec types I to VI.

Four targets (*mecA*, *ccrB2*, *mecI*, and *IS1272J*) were included in MB-PCR panel I, which was designed to identify methicillin resistance and definitively classify SCCmec types II (*mecA*, *ccrB2*, *mecI*) and IV (*mecA*, *ccrB2*, *IS1272J*) in a single reaction, with each type exhibiting three positive amplification curves (Fig. 2C and G). SCCmec types I and VI also possess the *IS1272J* target, resulting in two indistinguishable amplification curves (Fig. 2A and K), while SCCmec III possesses *mecI* (Fig. 2E) and SCCmec V is positive only for *mecA* (Fig.

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TABLE 4. *SCCmec* MB-PCR results for MRSA and MSSA strains from various collections

Source or study	Geographic origin	Yr of isolation	Genotypic background ^a	MB-PCR result ^b				No. (%) of isolates
				<i>mecA</i>	<i>ccr</i> type	<i>mec</i> class	<i>SCCmec</i>	
Kondo et al., 2007 (40) (<i>n</i> = 68) ^d	Canada, Denmark, Egypt, Ireland, Switzerland, United Kingdom, United States	1960–1993	CC5	+	1	B	I	1 (1.5)
				+	2	A	II	22 (32.4)
				+	2	B	IV	2 (2.9)
			CC8	+	1	B	I	13 (19.1)
				+	1, C	B	I ^c	1 (1.5)
				+	2	A	II	5 (7.4)
				+	3	A	III	2 (2.9)
				+	3, C	A	III	6 (8.8)
				+	2	B	IV	2 (2.9)
				+	2	A	II	1 (1.5)
			CC25	+	2	A	II	1 (1.5)
				+	2	A	II	3 (4.4)
			CC30	+	2	A	II	1 (1.5)
				+	3, C	A	III	1 (1.5)
			CC45	+	–	–	NT	1 (1.5)
				+	2	A	II	1 (1.5)
			CC59	+	3	A	III	1 (1.5)
			CC88	+	2	A	II	1 (1.5)
			CC121	+	1	B	I	1 (1.5)
Other	+	3	A	III	1 (1.5)			
	+	3, C	A	III	1 (1.5)			
	+	2	B	IV	1 (1.5)			
			+	–	A	NT	1 (1.5)	
Canada (<i>n</i> = 35)	Canada	2007–2009	CC5	+	2	A	II	5 (14.3)
				+	2	B	IV	5 (14.3)
				+	4	B	VI	1 (2.9)
			CC8	+	3, C	A	III	1 (2.9)
				+	2	B	IV	12 (34.3)
				+	C	C2	V	1 (2.9)
				+	4	A	VIII	1 (2.9)
			CC12	+	C	C2	V	1 (2.9)
				+	2	B	IV	1 (2.9)
			CC22	+	2	B	IV	1 (2.9)
			CC30	+	2	B	IV	1 (2.9)
			CC45	+	C	C2	V	1 (2.9)
				+	2	B	IV	2 (5.7)
			CC59	+	C	C2	V	2 (5.7)
			CC80	+	2	B	IV	1 (2.9)
NARSA MRSA (<i>n</i> = 66) ^d	Belgium, Brazil, France, Japan, Oman, South Korea, Switzerland, United Kingdom, United States	1935–2004	CC1	+	2	B	IV	5 (7.6)
				+	2	A	II	21 (31.8)
			CC5	+	2	B	IV	1 (1.5)
				+	2	B	I	11 (16.7)
				+	1	B	I	1 (1.5)
			CC8	+	2	A	II	1 (1.5)
				+	3, C	A	III	4 (6.1)
				+	2	B	IV	14 (21.2)
				+	2	B	IV	1 (1.5)
			CC22	+	2	B	IV	1 (1.5)
			CC30	+	2	A	II	3 (4.5)
CC45	+	2	A	II	2 (3.0)			
CC59	+	2	B	IV	1 (1.5)			
CC80	+	2	B	IV	1 (1.5)			
CC88	+	2	B	IV	1 (1.5)			
NARSA MSSA (<i>n</i> = 135)				–	1 ^e	–	–	2 (1.5)
				–	–	–	–	2 (1.5)
				–	–	–	–	14 (10.4)
				–	–	–	–	34 (25.2)
				–	–	–	–	5 (3.7)
				–	–	–	–	8 (5.9)
				–	–	–	–	28 (20.7)
				–	–	–	–	5 (3.7)
				–	–	–	–	1 (0.7)
				–	2	–	–	2 (1.5)
				–	–	–	–	1 (0.7)
				–	–	–	–	17 (12.6)
				–	–	–	–	16 (11.9)
BARG (<i>n</i> = 270)	United States (NYC)	1996	CC5	+	2	A	II	130 (48.1)
				+	2	B	IV	10 (3.7)
				+	2, 4	A	NT	1 (0.4)
			CC8	+	2	B	IV	44 (16.3)
				+	3, C	A	III	28 (10.4)
				+	3	A	III	1 (0.4)
				+	1	B	I	16 (5.9)
			CC30	+	–	–	NT	1 (0.4)
				+	2	A	II	20 (7.4)
			CC45	+	2, 4	A	NT	1 (0.4)
				+	2	A	II	4 (1.5)
			Other	+	2	A	II	2 (0.7)
				+	3	A	III	1 (0.4)

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TABLE 4—Continued

Source or study	Geographic origin	Yr of isolation	Genotypic background ^a	MB-PCR result ^b				No. (%) of isolates
				<i>mecA</i>	<i>ccr</i> type	<i>mec</i> class	SCC _{mec}	
				+	2	B	IV	5 (1.9)
				+	—	—	NT	1 (0.4)
				—	—	—	MSSA	5 (1.9)
NYC outpatients (<i>n</i> = 639)	United States (NYC)	2004–2006	CC1	+	2	B	IV	23 (3.6)
			CC8	+	2	B	IV	480 (75.1)
				+	2	A	II	1 (0.2)
				+	4	A	VIII	2 (0.3)
			CC5	+	2	A	II	91 (14.2)
				+	2	B	IV	23 (3.6)
			CC30	+	2	A	II	4 (0.6)
				+	2	B	IV	2 (0.3)
			CC45	+	2	A	II	1 (0.2)
				+	2	B	IV	2 (0.3)
			CC59	+	2	B	IV	8 (1.3)
			CC80	+	2	B	IV	1 (0.2)
CC88	+	2	B	IV	1 (0.2)			
NYC inpatients (<i>n</i> = 573)	United States (NYC)	2007–2008	CC5	+	2	B	IV	111 (19.4)
				+	2	A	II	361 (63.0)
				+	—	—	NT	2 (0.3)
			CC8	+	2	B	IV	61 (10.6)
				+	2	A	II	1 (0.2)
				+	C	C2	V	1 (0.2)
				+	4	A	VIII	1 (0.2)
			CC22	+	2	B	IV	1 (0.2)
			CC30	+	2	A	II	13 (2.3)
			CC45	+	2	A	II	18 (3.1)
			CC59	+	2	B	IV	1 (0.2)
				+	—	—	NT	1 (0.2)
			CC398	+	—	—	NT	1 (0.2)

^a Expressed in terms of the MLST CC; except for the NARSA strains, all CCs were inferred by *spa* repeat pattern analysis (51, 73).

^b The *ccr* type and *mec* class are given according to the nomenclature proposed by Chongtrakool et al. (9). NT, nontypeable; +, *mecA* positive; —, not detected.

^c The structure is similar to that of BK793; this isolate possesses an SCC_{Hg} region, as reported previously (40).

^d Not including strains already listed in Table 3.

^e To prevent misclassification, singleplex PCRs using *ccrB1*- and *ccrB3*-specific primers in combination with the *ccrB1/3* beacon were conducted separately to confirm the multiplex MB-PCR results.

^f “Other” NARSA MSSA strains include STs 10, 12, 25, 109, 395, 398, 707, 846, and 890.

2I); classification of these prototypes requires the second panel.

MB-PCR panel II targets *ccrC*, *ccrB1/3*, *ccrB4*, and the *mecC2* complex and can identify SCC_{mec} types I, III, V, and VI in combination with the results from MB-PCR panel I. As described in Materials and Methods, *ccrB1* and *ccrB3* utilize the same molecular beacon (*ccrB1/3*); this is a compromise measure adopted in order to accommodate two targets for each prototype by using only four dyes per panel. The amplification curve of *ccrB3* exhibits approximately half the magnitude of the amplification curve of *ccrB1* (Fig. 2B and F), due to the presence of a single-nucleotide mismatch between the *ccrB3* target and the *ccrB1/3* beacon (Table 1). SCC_{mec} type V displays two amplification curves (*ccrC* and *mecC2*), while SCC_{mec} types I (*ccrB1*) and VI (*ccrB4*) each exhibit only one curve (Fig. 2B and L), in MB-PCR panel II. The *ccrC* target is also present in SCC_{mec} III elements that harbor SCC_{Hg} (Fig. 2F) and by itself is not indicative of SCC_{mec} type III or V.

Sensitivity of MB-PCR. The sensitivity of the MB-PCR platform was evaluated utilizing the following six SCC_{mec} reference strains: NCTC10442 (type I), N315 (type II), 85/2082 (type III), CA05 (type IV), WIS (type V), and HDE288 (type VI). Limits of detection based on DNA copy numbers were obtained using purified genomic DNA as described in Materials and Methods, and the following lower limits were reliably detected within 40 cycles: 10 to 25 copies per PCR for *mecA*,

IS1272J, and *mecI*; and 25 to 40 copies per PCR for *ccrB2*, *ccrB1/3*, *ccrB4*, *ccrC*, and *mecC2*. Limits of detection based on bacterial CFU were also estimated using the rapid DNA boiling lysis procedure described above; the assay was capable of reproducibly detecting as few as 0.5×10^3 CFU per PCR for *mecA* and *mecI*; 1.5×10^3 CFU for IS1272J, *ccrB1/3*, and *ccrB2*; and 4×10^3 CFU for *ccrB4*, *ccrC*, and *mecC2*.

Specificity of MB-PCR. As shown in Fig. 2, all molecular beacon probe-primer sets displayed good specificity for their respective SCC_{mec} targets; no cross-reactivity was observed between the various *ccrB* and *ccrC* beacons, or among class A, B, and C2 *mec* complex beacons. Additionally, no amplification was observed for any of the molecular beacon targets with respect to the 50 nonstaphylococcal strains described in Materials and Methods, in agreement with the BLAST search results obtained during the assay design phase.

Validation of the MB-PCR assay. In order to validate the MB-PCR assay, a geographically, temporally, and genotypically diverse MRSA collection (*n* = 59) that had been characterized previously was tested using both panels (Table 3). We obtained 100% concordance for all isolates of SCC_{mec} types I to VI, except for a type III strain (DEN907) previously described as nontypeable for *ccrAB* by conventional PCR and not assigned to a specific SCC_{mec} type (57, 59). This strain was subsequently reanalyzed by *ccrB* sequence typing and was shown to cluster with *ccrAB3* (59, 60), in agreement with our

TABLE 5. MB-PCR SCCmec typing results for staphylococcal isolates (n = 71)

Species	MB-PCR result ^a				No. of isolates
	<i>mecA</i>	<i>ccr</i> type ^b	<i>mec</i> class	SCCmec	
<i>S. epidermidis</i>	+	3, C	A	III	6
	+	3	A	III	4
	+	2	B	IV	4
	+	2	A	II	1
	+	2, 3, 4	A	NT	2
	+	2, 4	A	NT	1
	+	—	—	NT	1
	—	2, 4	—	Unk	1
	—	2, 4, C	—	Unk	1
	—	—	—	—	4
	—	—	—	—	4
<i>S. haemolyticus</i>	+	C	C2	V	4
	+	2	B	IV	1
	+	—	—	NT	1
	—	4	—	Unk	1
<i>S. capitis</i>	+	2	A	II	4
	+	3, 4, C	A	NT	1
	—	—	—	—	1
<i>S. hominis</i>	+	3, 4, C	A	NT	1
	+	3, 4	A	NT	3
<i>S. saprophyticus</i>	—	C	—	Unk	1
	—	—	B	Unk	1
	—	—	—	—	3
<i>S. lugdunensis</i>	—	3	—	Unk	1
	—	—	—	—	3
<i>S. simulans</i>	—	4	—	Unk	2
	—	—	—	—	2
<i>S. lentus</i>	+	—	A	NT	1
	—	3	—	Unk	1
<i>S. xylosum</i>	—	—	—	—	5
<i>S. pseudintermedius</i>	—	—	—	—	2
<i>S. sciuri</i>	—	—	—	—	2
<i>S. warneri</i>	—	—	—	—	2
<i>S. gallinarum</i>	—	—	—	—	1
<i>S. succinus</i>	—	—	—	—	1
<i>S. aureus</i> subsp. <i>anaerobius</i>	—	—	—	—	1

^a The *ccr* type and *mec* class are given according to the nomenclature proposed by Chongtrakool et al. (9). NT, nontypeable SCCmec; Unk, unknown SCC structure; +, *mecA* positive; —, not detected.

^b To prevent misclassification, singleplex PCRs using *ccrB1*- and *ccrB3*-specific primers in combination with the *ccrB1/3* beacon were conducted separately to confirm multiplex MB-PCR results.

MB-PCR result of SCCmec III (Table 3). Several mosaic/composite staphylococcal cassettes with more than one *ccr* locus (not including SCCHg) were also identified. For example, strains BK793 (SCCmec I) and HAR36 (SCCmec IV) both possess *ccrC* in addition to their “native” *ccr* elements; similarly, strain AR43/3330.1 (SCCmec IVE) harbors *ccrAB4* in addition to *ccrB2*.

The MB-PCR assay was validated further using 68 MRSA isolates previously characterized by the SCCmec typing strategy described by Kondo et al. (40). Once again, our MB-PCR displayed 100% concordance with previously reported results; however, two strains classified as nontypeable in the study of Kondo et al. could not be successfully classified by our method, either (Table 4). Additionally, 35 blinded MRSA isolates from a Canadian laboratory (Table 4) displayed complete agreement with previous results obtained using various traditional SCCmec typing methods (29, 31, 56, 57), except for 1 isolate deemed nontypeable by conventional PCR. This isolate (BK23684) was positive by our MB-PCR method for both *mecI* and *ccrB4*, with a structure similar to that described recently for strain C10682 (GenBank accession number FJ390057), an epidemic Canadian clone (CMRSA-9) that has been classified by the IWG-SCC as SCCmec VIII (83).

Application of the multiplex PCR strategy. Several additional *S. aureus* collections (1,683 strains) were also tested in order to investigate the high-throughput applicability of the MB-PCR assay for SCCmec typing. Aggregate results are shown in Table 4, grouped according to source/study, genotypic background, and SCCmec type, and representing both historical and contemporary MRSA and MSSA isolates from nosocomial as well as outpatient settings. As shown in Table 4, our MB-PCR method was able to successfully type nearly all of the MRSA isolates from each of these collections, except for 10 strains classified as nontypeable. Several nontypeable MRSA strains were positive only for *mecA*, suggesting the possibility of novel SCCmec elements or structures. Addition-

TABLE 6. Primers used for long-range PCR and DNA sequencing

Procedure	Primer	Nucleotide sequence (5'–3')	Region amplified	Size	Reference
Long-range PCR for BK20781	Left-F	ATTCGGTACACGCAATGAAA	Right chromosome junction to <i>ccrB4</i>	8.5 kb	This study
	B4-R1	CITTATGATTCAATGCGTTC			
	A4-F1	TATTGTGTTGCTATCGCTTG	<i>ccrA4</i> to Tn554	9.5 kb	This study
	Tn554-R	CTAAAAACCAATTCCGACAG			
	Tn554-F	CGGAAAAATACCAAATCAAG	Tn554 to <i>mecA</i>	11.5 kb	This study
	MA-R	AGTTTTTCGAGTCCCTTTTT			
	MA-F	AACAAATGGATCAAATTTGG	<i>mecA</i> to left chromosome junction	7.3 kb	This study
	CR2	AAACGACATGAAAATCACCAT			
Long-range PCR for BK793	<i>ccrC</i> -2 Xsau325	ACTTATAATGGCTTCATGCTTACC GGATCAAACGGCCTGCACA	<i>ccrC</i> to <i>orfX</i>	10 kb	This study 28
Partial <i>ccrC</i> amplification and sequencing for BK793 and HAR36	<i>ccrC</i> -F2 <i>ccrC</i> -R1	CGAAATGGTRTTAAGTTGGAAA TGGCTTCATGYTTWCCTTTG	Within <i>ccrC</i>	587 bp	This study
Long-range PCR for AR43/3330.1	B4-F4 J IVc-R	CGGCAAAGGAACAAACCTAC ATAGATTCTACTGCAAGTCC	<i>ccrB4</i> to SCCmec J1	9.5 kb	This study 52
Partial <i>ccrAB4</i> sequencing for AR43/3330.1, BK2391, and BK2539	A4-F1 B4-R3	TATTGTGTTGCTATCGCTTG GTGCTAGGGAGCACTTCGTC	<i>ccrA4</i> to <i>ccrB4</i>	2.7 kb	This study

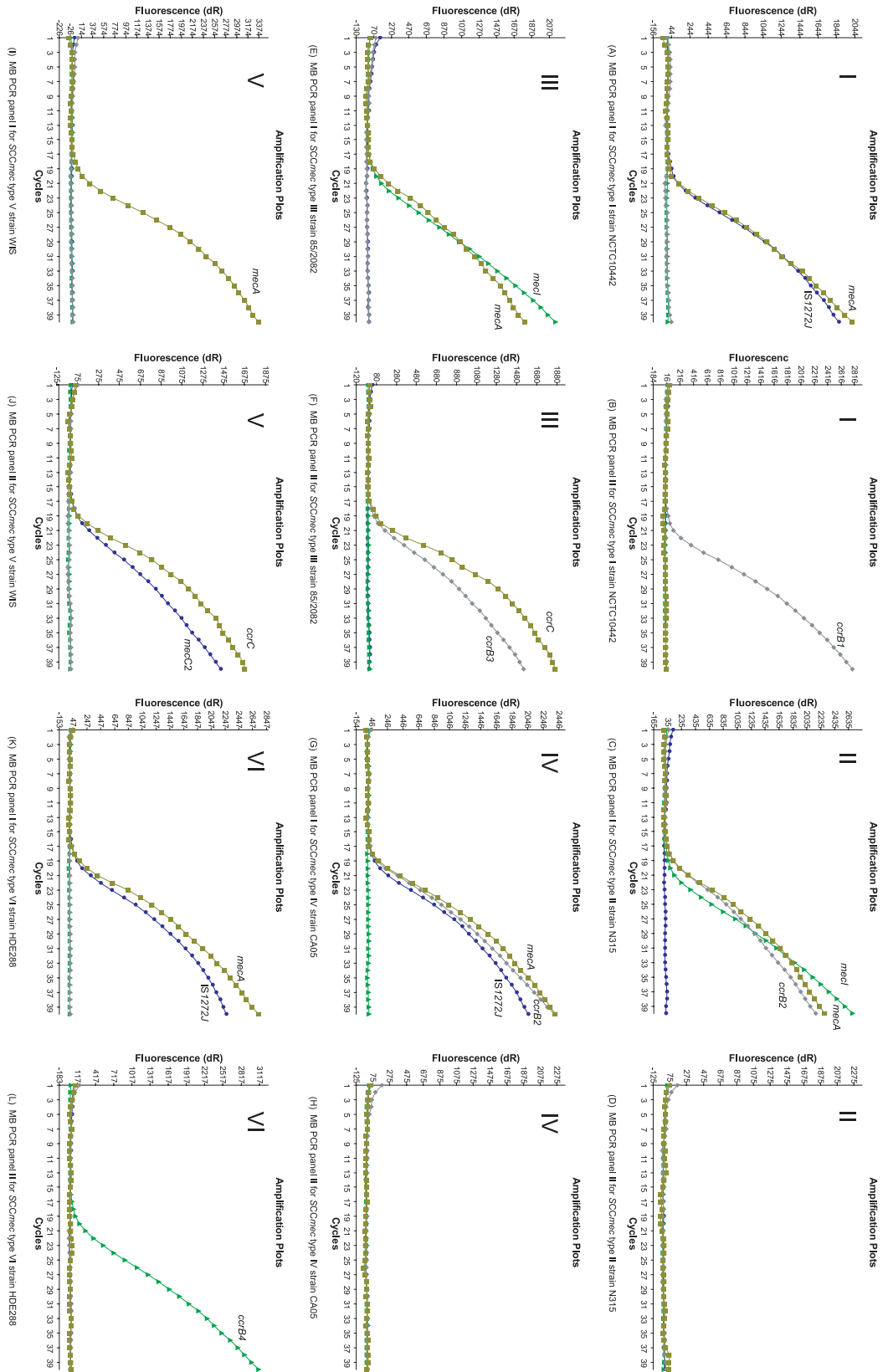


FIG. 2. Stratagene Mx4000 real-time PCR amplification curves for SCCmec type I to VI prototype strains (NCTC10442, N315, 85/2082, CA05, WIS, and HDE288). (Left) MB-PCR panel I results; (right) MB-PCR panel II results. Each MB-PCR panel consists of four molecular beacon probes labeled with different fluorescent dyes, as indicated in the keys at the bottom.

ally, two isolates from the BARG study with different genetic backgrounds, BK2391 (CC30) and BK2539 (CC5), possessed two *ccr* loci (*ccrB2*, *ccrB4*) in combination with the class A *mec* complex, a structure similar to that reported recently for two ST8-MRSA-II isolates from Ireland (M06/0075 and M06/0179) (69). Finally, five isolates from the same study, previously described by *mecA*-Tn554 hybridization typing as “NH:NH” or “NH:unique” (NH stands for “no hybridization”) (62), were confirmed as MSSA by our assay (Table 4).

In striking contrast to the BARG collection, which was obtained from 12 NYC area hospitals in 1996, no examples of SCC*mec* type I or III were found among a total of 1,212 MRSA isolates collected from the same region during 2004 to 2008. The majority of the latter isolates were either SCC*mec* II (40.4%) or SCC*mec* IV (58.9%), with the relative prevalence of each type differing according to setting; among nosocomial isolates, SCC*mec* type II was observed at a higher frequency (68.6%), whereas SCC*mec* IV was the predominant type among outpatient isolates (84.5%). Only one SCC*mec* V isolate was discovered in this collection, while no strains bearing SCC*mec* VI were observed. In contrast, three additional SCC*mec* VIII isolates (BK20015, BK20781, and BK25150) were discovered, two of which were found among outpatient CA-MRSA isolates. The entire SCC*mec* region from one of these strains (BK20781) has been sequenced and submitted to GenBank (accession number FJ670542).

An additional 71 staphylococcal strains, representing 15 different species, were also tested by the MB-PCR assay (Table 5); all were negative for the presence of *S. aureus* DNA by use of the *spa* molecular beacon, except for *S. aureus* subsp. *anaerobius*. Among 34 *mecA*-positive, coagulase-negative strains (MR-CoNS), 24 were successfully assigned SCC*mec* types; the rest were considered nontypeable, since they either possessed more than one *ccr* allotype or else were *mecA* positive with no detectable *ccr* or *mec* complex sequence (Table 5). Conversely, several strains positive for *ccr*, and one strain positive for *IS1272J* (*mec* complex B), were observed among *mecA*-negative strains. The *ccrB4* allotype was detected in five CoNS species (13 strains), usually accompanied by other types of *ccr* locus (e.g., *ccrB2*, *ccrB3*).

In contrast, no SCC elements were detected among 135 diverse MSSA strains from the NARSA collection mentioned above, except for 2 ST97 strains positive for *ccrB2* and 2 ST1 strains harboring *ccrB1*, as described previously for the SCC*far* element in MSSA476 (27) (Table 4). All MSSA isolates exhibited positive amplification using the *spa* molecular beacon.

DISCUSSION

SCC*mec* typing is one of the most important molecular tools for exploring the epidemiology and evolution of MRSA (12, 16). Knowledge about the nature of MRSA clones disseminating globally is necessary for implementing strategies to control the transmission of MRSA within nosocomial as well as community settings (6). Consequently, rapid characterization of MRSA strains by SCC*mec* typing is an important issue. A number of multiplex PCR-based SCC*mec* typing methods have been developed over the past several years (7, 40, 47, 53, 57, 84) and are reviewed extensively elsewhere (12). Several of these methods depend on targets located within J regions, in

order to characterize both prototypes and subtypes simultaneously; however, since J regions are likely to be less conserved than *ccr* and *mec* complexes, these methods risk sacrificing discriminatory power for convenience. The method described by Zhang and colleagues (84), for example, uses a single J-region target to identify SCC*mec* III and has recently been reported to misclassify SCC*mec* V isolates as type III (33). Another disadvantage shared by these methods is that they are based on electrophoretic analysis of multiple bands, which can be time-consuming, laborious, and potentially subjective. Furthermore, reproducible amplification of all target bands can depend on template quality, thereby compromising high-throughput studies in which DNA isolation is often a rate-limiting step.

Various multiplex schemes not based on conventional PCR have also been reported. A real-time PCR assay utilizing a mixture of TaqMan and minor groove binding probes, and targeting the *ccrB* regions of SCC*mec* types I to IV, was published in 2004 by François et al. (18). More recently, a novel approach utilizing *ccr*-specific padlock probes and tag microarray analysis for identification of SCC*mec* types I to VI has been described (42). A shortcoming of both methods is that they detect only one locus (*ccr*) and ignore the *mec* complex altogether, which may result in misclassification, given the growing number of reports describing composite SCC*mec* elements with multiple *ccr* loci, *ccr* loci not linked to the *mecA* gene, and novel combinations of the *mec* complex and *ccr* in SCC*mec* elements (5, 69, 83). SCC*mec* typing approaches based on DNA sequencing have also been described (45, 59, 60). However, these methods are not suitable for rapid typing and may not work for strains with multiple *ccr* elements (e.g., *ccrAB2* plus *ccrAB4*) or particular *ccr* allotypes (e.g., *ccrC*). Finally, an approach utilizing single-nucleotide polymorphisms has been described, in which a minimum set of 22 markers can be used to identify all presently known SCC*mec* types and subtypes (72). Although no specific detection procedure is described, the authors propose that the single-nucleotide polymorphism marker sets be used to guide the design of real-time PCR or oligoarray hybridization assays. While such an approach would undoubtedly be comprehensive and may be of interest for evolutionary studies, it is inherently impractical for clinical applications or routine genotyping.

SCC*mec* types are defined by binary combinations of the various *mec* class complexes and *ccr* allotypes described to date (Fig. 1). Accordingly, a novel method of nomenclature has been proposed by Chongtrakool et al. (9), in which a given SCC*mec* type is denoted by the *ccr* allotype (Arabic numeral) and *mec* complex (capital letter), as shown in Table 2. It is therefore recommended by the IWG-SCC that SCC*mec* elements be initially assigned using this combination of *mec* class and *ccr* type, followed by differences in J regions for individual subtypes (32). In this study, we developed an MB-PCR platform for rapid typing of SCC*mec* prototypes in MRSA strains based on the recommended system of nomenclature. Our MB-PCR assay targets all five known *ccr* allotypes (*ccrAB1* to *ccrAB4* and *ccrC*), along with the three principal *mec* classes (A, B, and C2) identified thus far in *S. aureus*; the emphasis is on identification of SCC*mec* prototypes, and no attempt is made to subtype.

Our MB-PCR assay demonstrated 100% concordance with

previous characterizations of two MRSA reference collections ($n = 127$), as well as with conventional SCC_{mec} typing of a blinded panel of isolates ($n = 35$). In this study, our assay identified only 10 (0.6%) nontypeable isolates out of 1,818 geographically and temporally diverse *S. aureus* strains. Moreover, we successfully identified a previously nontypeable strain (DEN907) as SCC_{mec} III, in agreement with previous results based on *ccrB* sequence typing (59). The assay is also capable of readily identifying novel or composite SCC_{mec} elements by virtue of the discrete target design. In this study, for instance, we discovered four strains possessing a novel combination of *ccrB4* and the class A *mec* complex, one of which was shown to be nearly identical (>99.9%) by DNA sequencing to a Canadian strain (GenBank accession number FJ390057) that has recently been classified as SCC_{mec} type VIII (83). Interestingly, whereas the initial description of SCC_{mec} VIII is based on a nosocomial CMRSA-9 strain (C10628) (83), two of the strains described above (BK20015 and BK20781) represent outpatient CA-MRSA isolates from the NYC area. To our knowledge, this is the first report of SCC_{mec} VIII outside of Canada.

As described in Results, additional *ccrC* and *ccrB4* elements were also found in several historic *S. aureus* strains (AR43/3330.1, HAR36, BK793, BK2391, and BK2539). An insertion of at least 8 kb between the *dcs* and *orfX* loci in strain BK793 has been reported previously (61). Long-range PCR and sequencing using primers spanning *ccrC* and *orfX* (Table 6) were therefore performed, confirming the presence of a *ccrC5*-like sequence similar to that in *S. haemolyticus* strain JCSC1435 (GenBank accession number AP006716) and located approximately 10 kb upstream of *orfX*. Similarly, strain HAR36 is characterized as SCC_{mec} IV but possesses a J3 region locus associated with SCC_{mec} type III (SCCHg) (53); *ccrC* universal primers (Table 6) were therefore designed and used to confirm the presence of a *ccrC3* element in this strain by DNA sequencing. Long-range PCR and sequencing of a *ccrB4* element detected in prototype SCC_{mec} IVE strain AR43/3330.1 likewise demonstrated close identity with *S. epidermidis* strain ATCC 12228 (data not shown), and two class A *mec* strains from the 1996 BARG study (BK2391 and BK2539) were also found to possess both *ccrB4* and *ccrB2*. Such examples of multiple or composite SCC_{mec} elements argue strongly against the reliability of using a single locus (e.g., *ccrC* or *ccrB4*) for assignment of SCC_{mec} types (18, 42, 83), due to the heterogeneity and mobility of SCC elements (20, 63). For instance, novel SCC_{mec} types involving combinations of *ccrC* with class B, E, and C1 *mec* complexes (5B, 5B1, 5E, and 5C1, respectively) have been reported (5, 55), but several existing SCC_{mec} typing methods may misclassify these as SCC_{mec} type V, based on the use of *ccrC* alone for typing (42, 47).

Our real-time MB-PCR platform offers several advantages over existing methods. Since the specificity of the assay is dependent on both oligonucleotide primer and molecular beacon design, the discriminatory power of target identification exceeds that afforded by conventional PCR. Although the assay is subject to target abrogation resulting from sequence heterogeneity or mosaic structure, the results are not sensitive to changes in fragment size as a result of insertions or deletions. Furthermore, the requirement for template quality is minimal, allowing for rapid typing of large numbers of isolates

using a simple boiling lysis method. All PCR amplicons in our multiplex platform are ≤ 174 bp long (Table 1), thereby contributing directly to increased amplification efficiency and sensitivity of detection using minimal reaction volumes, as well as facilitating analysis of low-quality/fragmented DNA. The sensitivity afforded by our method may also allow for SCC_{mec} typing directly from blood cultures (38), which is of potential interest in light of recent reports describing differences in clinical outcome between *S. aureus* bacteremias associated with particular SCC_{mec} types (19).

The procedure described in this study is also particularly well-suited for large-scale MRSA analyses. Given the relaxed requirement for template quality, DNA isolation need not be a rate-limiting step, and strains can be typed easily in batches of 96 using the high-throughput method mentioned above. Data interpretation is also relatively straightforward compared to electrophoretic analysis of banding patterns, since it involves simple “yes/no” interrogation of each individual target (Fig. 2). In our experience, a single panel (MB-PCR panel I or II) can be performed on 96 strains in approximately 3 to 4 h, including DNA isolation, PCR cycling, and data analysis. Moreover, where routine typing of contemporary isolates is involved, a single panel may be sufficient for definitive SCC_{mec} assignment of most strains. In our geographic region, for example, SCC_{mec} types II and IV account for nearly all nosocomial and community MRSA isolates (Table 4), such that the results of MB-PCR panel I are usually definitive.

Our assay can also be used to investigate SCC elements in other staphylococcal species, as demonstrated by analysis of 71 strains from 15 different species (Table 5). Our results suggest that SCC structure is very diverse among CoNS, with various strains possessing more than one *ccr* allotype, or else possessing SCC elements without *mecA*, in agreement with previous studies (21, 66). The *ccrB4* allotype in particular appears to have been transmitted from *S. epidermidis* to *S. aureus* on more than one occasion, and *ccrB4* elements derived from *S. epidermidis* strain ATCC 12228 have been reported in MRSA strains of various genetic backgrounds (69, 83). Given the apparent prevalence of the *ccrB4* allotype among CoNS species, it is reasonable to expect that further transmission will occur in the future, leading to novel SCC_{mec} variants or prototypes in MRSA (20).

Several limitations are associated with our assay. First, despite a concerted effort to design all molecular beacon targets using conserved regions, the universality of any given target cannot be guaranteed, due to the heterogeneity of SCC_{mec} elements. For example, the class B *mec* complex target (IS1272J) selected for this study was not detected within the class B *mec* complexes from two *S. haemolyticus* strains, due to a previously published deletion (39); although no such deletion has been reported in MRSA, the potential for inability to detect class B *mec* in variant strains exists. Second, since a hybrid beacon is used to detect both *ccrB1* and *ccrB3*, the potential exists for misclassification of isolates possessing *mec* class A in combination with *ccrAB1*, or *mec* class B in combination with *ccrAB3*. However, since there have been no reports to date of *S. aureus* containing SCC_{mec} elements possessing (i) both *ccrAB1* and *ccrAB3*, (ii) *ccrAB1* with class A *mec*, or (iii) *ccrAB3* with class B *mec*, the *ccrB1/3* beacon in conjunction with IS1272J (class B *mec*) or *mecI* (class A *mec*) can theoret-

ically be used to distinguish SCCmec types I and III, respectively. Although no such combinations have been observed in *S. aureus*, an example of the former observed in a *Staphylococcus cohnii* strain has been described previously (66). Third, as described in Materials and Methods, the *ccrC* molecular beacon may not reliably detect certain variants of this allotype; among these, however, only *ccrC4* and *ccrC8* have been described in *S. aureus* (48, 74). Finally, our assay is not capable of detecting the recently described *mec* class C1 element associated with SCCmec type VII (5).

An additional limitation of our assay is the inability to detect SCCmec subtypes. In this study our goal was to design a multiplex MB-PCR platform for rapid typing of SCCmec prototypes, and J-region targets used for subtyping were not included. However, depending on the needs of potential users, subtyping probes can be designed and substituted in either of the two MB-PCR panels. As an example, a primer-probe set specific to the J1 region of SCCmec IVa was designed (Table 1) and substituted for *mecI* in panel I, in order to further characterize SCCmec IV isolates from a large-scale CA-MRSA study. Only SCCmec IV isolates bearing the IVa-specific J1-region target were amplified using this approach (data not shown). Similar probes can be designed for other SCCmec subtypes of interest, as has been done using conventional PCR; alternatively, dedicated subtyping panels can be created using combinations of different J-region targets.

As the availability of real-time PCR instruments with 5-dye capability becomes commonplace, it will be possible to expand each panel to include an additional target. For example, a *ccrB3*-specific target can be added to panel I, thereby allowing for the definitive identification of SCCmec types II, III, and IV in a single reaction and eliminating the need to use a hybrid target for the identification of *ccrB1* and *ccrB3*. Similarly, an additional target can be added to panel II in order to detect the *mecC1* complex mentioned above (5), thereby allowing the assay to definitively classify SCCmec type VII as well. With projected developments in real-time PCR spectrofluorometric thermal cyclers, more than seven fluorophore targets may be detected in a single reaction.

Finally, although the initial investment for the full complement of molecular beacons is significant (approximately \$400 per beacon, including purification), it is offset by the reduced requirements for template quality, reaction volume, and time. A detailed cost analysis of all reagents and consumables (not including labor) indicates that the cost of running both panels is approximately USD 2.6 per sample. Moreover, as described above, one panel may suffice for routine typing, in which case the cost per sample is only USD 1.3, significantly less than that reported for conventional PCR (USD 2.4 to 4.9) (47). Consequently, for laboratories with access to real-time PCR instrumentation, our assay represents an economically feasible approach for high-throughput SCCmec typing, especially considering the additional savings in labor and processing time.

We have developed a multiplex MB-PCR platform for rapid, high-throughput SCCmec typing of MRSA. This novel assay is faster, more robust, and significantly more sensitive than previously published schemes and is based on unambiguous classification of SCCmec types according to established nomenclature. Nevertheless, as molecular characterization of MRSA

and MR-CoNS continues to widen in scope, it is likely that numerous SCC elements and/or novel SCCmec types will be discovered. Therefore, the typing system described here should be seen as reflecting the current situation and must necessarily be developed further to accommodate novel developments.

ACKNOWLEDGMENTS

We thank T. Ito and T. Smith for the generous gift of control strains, as well as assistance with comparison of SCCmec typing results. We also acknowledge B. Mathema for editorial assistance with reading the manuscript.

REFERENCES

- Adcock, P. M., P. Pastor, F. Medley, J. E. Patterson, and T. V. Murphy. 1998. Methicillin-resistant *Staphylococcus aureus* in two child care centers. *J. Infect. Dis.* **178**:577–580.
- Aires de Sousa, M., and H. de Lencastre. 2003. Evolution of sporadic isolates of methicillin-resistant *Staphylococcus aureus* (MRSA) in hospitals and their similarities to isolates of community-acquired MRSA. *J. Clin. Microbiol.* **41**:3806–3815.
- Altschul, S. F., W. Gish, W. Miller, E. W. Myers, and D. J. Lipman. 1990. Basic local alignment search tool. *J. Mol. Biol.* **215**:403–410.
- Baba, T., F. Takeuchi, M. Kuroda, H. Yuzawa, K. Aoki, A. Oguchi, Y. Nagai, N. Iwama, K. Asano, T. Naimi, H. Kuroda, L. Cui, K. Yamamoto, and K. Hiramatsu. 2002. Genome and virulence determinants of high virulence community-acquired MRSA. *Lancet* **359**:1819–1827.
- Berglund, C., T. Ito, M. Ikeda, X. X. Ma, B. Soderquist, and K. Hiramatsu. 2008. Novel type of staphylococcal cassette chromosome *mec* in a methicillin-resistant *Staphylococcus aureus* strain isolated in Sweden. *Antimicrob. Agents Chemother.* **52**:3512–3516.
- Boucher, H. W., and G. R. Corey. 2008. Epidemiology of methicillin-resistant *Staphylococcus aureus*. *Clin. Infect. Dis.* **46**(Suppl. 5):S344–S349.
- Boye, K., M. D. Bartels, I. S. Andersen, J. A. Moller, and H. Westh. 2007. A new multiplex PCR for easy screening of methicillin-resistant *Staphylococcus aureus* SCCmec types I–V. *Clin. Microbiol. Infect.* **13**:725–727.
- Boyle-Vavra, S., B. Ereshefsky, C. C. Wang, and R. S. Daum. 2005. Successful multiresistant community-associated methicillin-resistant *Staphylococcus aureus* lineage from Taipei, Taiwan, that carries either the novel staphylococcal chromosome cassette *mec* (SCCmec) type VT or SCCmec type IV. *J. Clin. Microbiol.* **43**:4719–4730.
- Chongtrakool, P., T. Ito, X. X. Ma, Y. Kondo, S. Trakulsomboon, C. Tien-sasitorn, M. Jamklang, T. Chavalit, J. H. Song, and K. Hiramatsu. 2006. Staphylococcal cassette chromosome *mec* (SCCmec) typing of methicillin-resistant *Staphylococcus aureus* strains isolated in 11 Asian countries: a proposal for a new nomenclature for SCCmec elements. *Antimicrob. Agents Chemother.* **50**:1001–1012.
- Daum, R. S., T. Ito, K. Hiramatsu, F. Hussain, K. Mongkolrattanothai, M. Jamklang, and S. Boyle-Vavra. 2002. A novel methicillin-resistance cassette in community-acquired methicillin-resistant *Staphylococcus aureus* isolates of diverse genetic backgrounds. *J. Infect. Dis.* **186**:1344–1347.
- Descloux, S., A. Rossano, and V. Perreten. 2008. Characterization of new staphylococcal cassette chromosome *mec* (SCCmec) and topoisomerase genes in fluoroquinolone- and methicillin-resistant *Staphylococcus pseudintermedius*. *J. Clin. Microbiol.* **46**:1818–1823.
- Deurenberg, R. H., and E. E. Stobberingh. 2008. The evolution of *Staphylococcus aureus*. *Infect. Genet. Evol.* **8**:747–763.
- Diep, B. A., S. R. Gill, R. F. Chang, T. H. Phan, J. H. Chen, M. G. Davidson, F. Lin, J. Lin, H. A. Carleton, E. F. Mongodin, G. F. Sensabaugh, and F. Perdreau-Remington. 2006. Complete genome sequence of USA300, an epidemic clone of community-acquired methicillin-resistant *Staphylococcus aureus*. *Lancet* **367**:731–739.
- Enright, M. C., N. P. Day, C. E. Davies, S. J. Peacock, and B. G. Spratt. 2000. Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J. Clin. Microbiol.* **38**:1008–1015.
- Enright, M. C., D. A. Robinson, G. Randle, E. J. Feil, H. Grundmann, and B. G. Spratt. 2002. The evolutionary history of methicillin-resistant *Staphylococcus aureus* (MRSA). *Proc. Natl. Acad. Sci. USA* **99**:7687–7692.
- Faria, N. A., J. A. Carrico, D. C. Oliveira, M. Ramirez, and H. de Lencastre. 2008. Analysis of typing methods for epidemiological surveillance of both methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* strains. *J. Clin. Microbiol.* **46**:136–144.
- Feil, E. J., B. C. Li, D. M. Aanensen, W. P. Hanage, and B. G. Spratt. 2004. eBURST: inferring patterns of evolutionary descent among clusters of related bacterial genotypes from multilocus sequence typing data. *J. Bacteriol.* **186**:1518–1530.
- François, P., G. Renzi, D. Pittet, M. Bento, D. Lew, S. Harbarth, P. Vaudaux, and J. Schrenzel. 2004. A novel multiplex real-time PCR assay for rapid

- typing of major staphylococcal cassette chromosome *mec* elements. *J. Clin. Microbiol.* **42**:3309–3312.
19. Ganga, R., K. Riederer, M. Sharma, M. G. Fakih, L. B. Johnson, S. Shemes, and R. Khatib. 2009. Role of SCC_{mec} type in outcome of *Staphylococcus aureus* bacteremia in a single medical center. *J. Clin. Microbiol.* **47**:590–595.
 20. Hanssen, A. M., and J. U. Ericson Sollid. 2006. SCC_{mec} in staphylococci: genes on the move. *FEMS Immunol. Med. Microbiol.* **46**:8–20.
 21. Hanssen, A. M., G. Kjeldsen, and J. U. Sollid. 2004. Local variants of staphylococcal cassette chromosome *mec* in sporadic methicillin-resistant *Staphylococcus aureus* and methicillin-resistant coagulase-negative staphylococci: evidence of horizontal gene transfer? *Antimicrob. Agents Chemother.* **48**:285–296.
 22. Harmsen, D., H. Claus, W. Witte, J. Rothganger, D. Turnwald, and U. Vogel. 2003. Typing of methicillin-resistant *Staphylococcus aureus* in a university hospital setting by using novel software for *spa* repeat determination and database management. *J. Clin. Microbiol.* **41**:5442–5448.
 23. Hein, L., A. Lehner, P. Rieck, K. Klein, E. Brandl, and M. Wagner. 2001. Comparison of different approaches to quantify *Staphylococcus aureus* cells by real-time quantitative PCR and application of this technique for examination of cheese. *Appl. Environ. Microbiol.* **67**:3122–3126.
 24. Higuchi, W., T. Takano, L. J. Teng, and T. Yamamoto. 2008. Structure and specific detection of staphylococcal cassette chromosome *mec* type VII. *Biochem. Biophys. Res. Commun.* **377**:752–756.
 25. Hiramatsu, K., N. Aritaka, H. Hanaki, S. Kawasaki, Y. Hosoda, S. Hori, Y. Fukuchi, and I. Kobayashi. 1997. Dissemination in Japanese hospitals of strains of *Staphylococcus aureus* heterogeneously resistant to vancomycin. *Lancet* **350**:1670–1673.
 26. Hisata, K., K. Kuwahara-Arai, M. Yamanoto, T. Ito, Y. Nakatomi, L. Cui, T. Baba, M. Terasawa, C. Sotozono, S. Kinoshita, Y. Yamashiro, and K. Hiramatsu. 2005. Dissemination of methicillin-resistant staphylococci among healthy Japanese children. *J. Clin. Microbiol.* **43**:3364–3372.
 27. Holden, M. T., E. J. Feil, J. A. Lindsay, S. J. Peacock, N. P. Day, M. C. Enright, T. J. Foster, C. E. Moore, L. Hurst, R. Atkin, A. Barron, N. Bason, S. D. Bentley, C. Chillingworth, T. Chillingworth, C. Churcher, L. Clark, C. Corton, A. Cronin, J. Doggett, L. Dowd, T. Feltwell, Z. Hance, B. Harris, H. Hauser, S. Holroyd, K. Jagels, K. D. James, N. Lennard, A. Line, R. Mayes, S. Moule, K. Mungall, D. Ormond, M. A. Quail, E. Rabinowitz, K. Rutherford, M. Sanders, S. Sharp, M. Simmonds, K. Stevens, S. Whitehead, B. G. Barrell, B. G. Spratt, and J. Parkhill. 2004. Complete genomes of two clinical *Staphylococcus aureus* strains: evidence for the rapid evolution of virulence and drug resistance. *Proc. Natl. Acad. Sci. USA* **101**:9786–9791.
 28. Huletsky, A., R. Giroux, V. Rossbach, M. Gagnon, M. Vaillancourt, M. Bernier, F. Gagnon, K. Truchon, M. Bastien, F. J. Picard, A. van Belkum, M. Ouellette, P. H. Roy, and M. G. Bergeron. 2004. New real-time PCR assay for rapid detection of methicillin-resistant *Staphylococcus aureus* directly from specimens containing a mixture of staphylococci. *J. Clin. Microbiol.* **42**:1875–1884.
 29. Ito, T., Y. Katayama, K. Asada, N. Mori, K. Tsutsumimoto, C. Tiensasitorn, and K. Hiramatsu. 2001. Structural comparison of three types of staphylococcal cassette chromosome *mec* integrated in the chromosome in methicillin-resistant *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* **45**:1323–1336.
 30. Ito, T., Y. Katayama, and K. Hiramatsu. 1999. Cloning and nucleotide sequence determination of the entire *mec* DNA of pre-methicillin-resistant *Staphylococcus aureus* N315. *Antimicrob. Agents Chemother.* **43**:1449–1458.
 31. Ito, T., X. X. Ma, F. Takeuchi, K. Okuma, H. Yuzawa, and K. Hiramatsu. 2004. Novel type V staphylococcal cassette chromosome *mec* driven by a novel cassette chromosome recombinase, *ccrC*. *Antimicrob. Agents Chemother.* **48**:2637–2651.
 32. IWG-SCC. 31 August 2009. Classification of staphylococcal cassette chromosome *mec* (SCC_{mec}): guidelines for reporting novel SCC_{mec} elements. *Antimicrob. Agents Chemother.* doi:10.1128/AAC.00579-09.
 33. Jansen, M. D., A. T. Box, and A. C. Fluit. 2009. SCC_{mec} typing in methicillin-resistant *Staphylococcus aureus* strains of animal origin. *Emerg. Infect. Dis.* **15**:136–137.
 34. Jevons, M. P. 1961. “Celbenin”-resistant staphylococci. *Br. Med. J.* **1**:124–125.
 35. Katayama, Y., T. Ito, and K. Hiramatsu. 2000. A new class of genetic element, staphylococcal cassette chromosome *mec*, encodes methicillin resistance in *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* **44**:1549–1555.
 36. Katayama, Y., F. Takeuchi, T. Ito, X. X. Ma, Y. Ui-Mizutani, I. Kobayashi, and K. Hiramatsu. 2003. Identification in methicillin-susceptible *Staphylococcus hominis* of an active primordial mobile genetic element for the staphylococcal cassette chromosome *mec* of methicillin-resistant *Staphylococcus aureus*. *J. Bacteriol.* **185**:2711–2722.
 37. Kishii, K., T. Ito, S. Watanabe, K. Okuzumi, and K. Hiramatsu. 2008. Recurrence of heterogeneous methicillin-resistant *Staphylococcus aureus* (MRSA) among the MRSA clinical isolates in a Japanese university hospital. *J. Antimicrob. Chemother.* **62**:324–328.
 38. Klouche, M., and U. Schroder. 2008. Rapid methods for diagnosis of blood-stream infections. *Clin. Chem. Lab. Med.* **46**:888–908.
 39. Kobayashi, N., S. Urasawa, N. Uehara, and N. Watanabe. 1999. Distribution of insertion sequence-like element IS1272 and its position relative to methicillin resistance genes in clinically important staphylococci. *Antimicrob. Agents Chemother.* **43**:2780–2782.
 40. Kondo, Y., T. Ito, X. X. Ma, S. Watanabe, B. N. Kreiswirth, J. Etienne, and K. Hiramatsu. 2007. Combination of multiplex PCRs for staphylococcal cassette chromosome *mec* type assignment: rapid identification system for *mec*, *ccr*, and major differences in junkyard regions. *Antimicrob. Agents Chemother.* **51**:264–274.
 41. Kuroda, M., T. Ohta, I. Uchiyama, T. Baba, H. Yuzawa, I. Kobayashi, L. Cui, A. Oguchi, K. Aoki, Y. Nagai, J. Lian, T. Ito, M. Kanamori, H. Matsumaru, A. Maruyama, H. Murakami, A. Hosoyama, Y. Mizutani-Ui, N. K. Takahashi, T. Sawano, R. Inoue, C. Kaito, K. Sekimizu, H. Hirakawa, S. Kuhara, S. Goto, J. Yabuzaki, M. Kanehisa, A. Yamashita, K. Oshima, K. Furuya, C. Yoshino, T. Shiba, M. Hattori, N. Ogasawara, H. Hayashi, and K. Hiramatsu. 2001. Whole genome sequencing of methicillin-resistant *Staphylococcus aureus*. *Lancet* **357**:1225–1240.
 42. Kurt, K., A. Alderborn, M. Nilsson, B. Strommenger, W. Witte, and U. Nubel. 2009. Multiplexed genotyping of methicillin-resistant *Staphylococcus aureus* isolates by use of padlock probes and tag microarrays. *J. Clin. Microbiol.* **47**:577–585.
 43. Larkin, M. A., G. Blackshields, N. P. Brown, R. Chenna, P. A. McGettigan, H. McWilliam, F. Valentin, I. M. Wallace, A. Wilm, R. Lopez, J. D. Thompson, T. J. Gibson, and D. G. Higgins. 2007. Clustal W and Clustal X version 2.0. *Bioinformatics* **23**:2947–2948.
 44. Lim, T. T., F. N. Chong, F. G. O'Brien, and W. B. Grubb. 2003. Are all community methicillin-resistant *Staphylococcus aureus* related? A comparison of their *mec* regions. *Pathology (Philadelphia)* **35**:336–343.
 45. Lina, G., G. Durand, C. Berchich, B. Short, H. Meugnier, F. Vandenesch, J. Etienne, and M. C. Enright. 2006. Staphylococcal chromosome cassette evolution in *Staphylococcus aureus* inferred from *ccr* gene complex sequence typing analysis. *Clin. Microbiol. Infect.* **12**:1175–1184.
 46. Lowy, F. D. 1998. *Staphylococcus aureus* infections. *N. Engl. J. Med.* **339**:520–532.
 47. Lu, P. L., J. C. Chang, H. T. Hsu, J. H. Chen, F. J. Chen, S. F. Lin, and L. K. Siu. 2008. One tube multiplex PCR for simple screening of SCC_{mec} I-V types of methicillin-resistant *Staphylococcus aureus*. *J. Chemother.* **20**:690–696.
 48. Luong, T. T., S. Ouyang, K. Bush, and C. Y. Lee. 2002. Type 1 capsule genes of *Staphylococcus aureus* are carried in a staphylococcal cassette chromosome genetic element. *J. Bacteriol.* **184**:3623–3629.
 49. Ma, X. X., T. Ito, P. Chongtrakool, and K. Hiramatsu. 2006. Predominance of clones carrying Pantone-Valentine leukocidin genes among methicillin-resistant *Staphylococcus aureus* strains isolated in Japanese hospitals from 1979 to 1985. *J. Clin. Microbiol.* **44**:4515–4527.
 50. Ma, X. X., T. Ito, C. Tiensasitorn, M. Jamklang, P. Chongtrakool, S. Boyle-Vavra, R. S. Daum, and K. Hiramatsu. 2002. Novel type of staphylococcal cassette chromosome *mec* identified in community-acquired methicillin-resistant *Staphylococcus aureus* strains. *Antimicrob. Agents Chemother.* **46**:1147–1152.
 51. Mathema, B., J. Mediavilla, and B. N. Kreiswirth. 2008. Sequence analysis of the variable number tandem repeat in *Staphylococcus aureus* protein A gene: *spa* typing. *Methods Mol. Biol.* **431**:285–305.
 52. Milheiro, C., D. C. Oliveira, and H. de Lencastre. 2007. Multiplex PCR strategy for subtyping the staphylococcal cassette chromosome *mec* type IV in methicillin-resistant *Staphylococcus aureus*: ‘SCC_{mec} IV multiplex.’ *J. Antimicrob. Chemother.* **60**:42–48.
 53. Milheiro, C., D. C. Oliveira, and H. de Lencastre. 2007. Update to the multiplex PCR strategy for assignment of *mec* element types in *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* **51**:3374–3377.
 54. Morgan, M. 2008. Methicillin-resistant *Staphylococcus aureus* and animals: zoonosis or humanosis? *J. Antimicrob. Chemother.* **62**:1181–1187.
 55. O'Brien, F. G., G. W. Coombs, J. C. Pearson, K. J. Christiansen, and W. B. Grubb. 2005. Type V staphylococcal cassette chromosome *mec* in community staphylococci from Australia. *Antimicrob. Agents Chemother.* **49**:5129–5132.
 56. Okuma, K., K. Iwakawa, J. D. Turnidge, W. B. Grubb, J. M. Bell, F. G. O'Brien, G. W. Coombs, J. W. Pearman, F. C. Tenover, M. Kapi, C. Tiensasitorn, T. Ito, and K. Hiramatsu. 2002. Dissemination of new methicillin-resistant *Staphylococcus aureus* clones in the community. *J. Clin. Microbiol.* **40**:4289–4294.
 57. Oliveira, D. C., and H. de Lencastre. 2002. Multiplex PCR strategy for rapid identification of structural types and variants of the *mec* element in methicillin-resistant *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* **46**:2155–2161.
 58. Oliveira, D. C., C. Milheiro, and H. de Lencastre. 2006. Redefining a structural variant of staphylococcal cassette chromosome *mec*, SCC_{mec} type VI. *Antimicrob. Agents Chemother.* **50**:3457–3459.
 59. Oliveira, D. C., C. Milheiro, S. Vinga, and H. de Lencastre. 2006. Assessment of allelic variation in the *ccrAB* locus in methicillin-resistant *Staphylococcus aureus* clones. *J. Antimicrob. Chemother.* **58**:23–30.
 60. Oliveira, D. C., M. Santos, C. Milheiro, J. A. Carrico, S. Vinga, A. L.

- Oliveira, and H. de Lencastre. 2008. *ccrB* typing tool: an online resource for staphylococci *ccrB* sequence typing. *J. Antimicrob. Chemother.* **61**:959–960.
61. Oliveira, D. C., A. Tomasz, and H. de Lencastre. 2001. The evolution of pandemic clones of methicillin-resistant *Staphylococcus aureus*: identification of two ancestral genetic backgrounds and the associated *mec* elements. *Microb. Drug Resist.* **7**:349–361.
 62. Roberts, R. B., A. de Lencastre, W. Eisner, E. P. Severina, B. Shopsin, B. N. Kreiswirth, A. Tomasz, et al. 1998. Molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in 12 New York hospitals. *J. Infect. Dis.* **178**:164–171.
 63. Robinson, D. A., and M. C. Enright. 2003. Evolutionary models of the emergence of methicillin-resistant *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* **47**:3926–3934.
 64. Robinson, D. A., A. M. Kearns, A. Holmes, D. Morrison, H. Grundmann, G. Edwards, F. G. O'Brien, F. C. Tenover, L. K. McDougal, A. B. Monk, and M. C. Enright. 2005. Re-emergence of early pandemic *Staphylococcus aureus* as a community-acquired methicillin-resistant clone. *Lancet* **365**:1256–1258.
 65. Rozen, S., and H. Skaletsky. 2000. Primer3 on the WWW for general users and for biologist programmers. *Methods Mol. Biol.* **132**:365–386.
 66. Ruppé, E., F. Barbier, Y. Mesli, A. Maiga, R. Cojocar, M. Benkhalfat, S. Benchouk, H. Hassaine, I. Maiga, A. Diallo, A. K. Koumare, K. Ouattara, S. Soumare, J. B. Dufourcq, C. Nareth, J. L. Sarthou, A. Andremon, and R. Ruimy. 2009. Diversity of staphylococcal cassette chromosome *mec* structures in methicillin-resistant *Staphylococcus epidermidis* and *Staphylococcus haemolyticus* strains among outpatients from four countries. *Antimicrob. Agents Chemother.* **53**:442–449.
 67. Shopsin, B., M. Gomez, S. O. Montgomery, D. H. Smith, M. Waddington, D. E. Dodge, D. A. Bost, M. Riehm, S. Naidich, and B. N. Kreiswirth. 1999. Evaluation of protein A gene polymorphic region DNA sequencing for typing of *Staphylococcus aureus* strains. *J. Clin. Microbiol.* **37**:3556–3563.
 68. Shore, A., A. S. Rossney, C. T. Keane, M. C. Enright, and D. C. Coleman. 2005. Seven novel variants of the staphylococcal chromosomal cassette *mec* in methicillin-resistant *Staphylococcus aureus* isolates from Ireland. *Antimicrob. Agents Chemother.* **49**:2070–2083.
 69. Shore, A. C., A. S. Rossney, B. O'Connell, C. M. Herra, D. J. Sullivan, H. Humphreys, and D. C. Coleman. 2008. Detection of staphylococcal cassette chromosome *mec*-associated DNA segments in multiresistant methicillin-susceptible *Staphylococcus aureus* (MSSA) and identification of *Staphylococcus epidermidis ccrAB4* in both methicillin-resistant *S. aureus* and MSSA. *Antimicrob. Agents Chemother.* **52**:4407–4419.
 70. Sinsimer, D., S. Leekha, S. Park, S. A. Marras, L. Koreen, B. Willey, S. Naidich, K. A. Musser, and B. N. Kreiswirth. 2005. Use of a multiplex molecular beacon platform for rapid detection of methicillin and vancomycin resistance in *Staphylococcus aureus*. *J. Clin. Microbiol.* **43**:4585–4591.
 71. Smith, T. C., M. J. Male, A. L. Harper, J. S. Kroeger, G. P. Tinkler, E. D. Moritz, A. W. Capuano, L. A. Herwaldt, and D. J. Diekema. 2008. Methicillin-resistant *Staphylococcus aureus* (MRSA) strain ST398 is present in Midwestern U.S. swine and swine workers. *PLoS ONE* **4**:e4258.
 72. Stephens, A. J., F. Huygens, J. Inman-Bamber, E. P. Price, G. R. Nimmo, J. Schooneveldt, W. Munckhof, and P. M. Giffard. 2006. Methicillin-resistant *Staphylococcus aureus* genotyping using a small set of polymorphisms. *J. Med. Microbiol.* **55**:43–51.
 73. Strommenger, B., C. Kettlitz, T. Weniger, D. Harmsen, A. W. Friedrich, and W. Witte. 2006. Assignment of *Staphylococcus* isolates to groups by *spa* typing, SmaI macrorestriction analysis, and multilocus sequence typing. *J. Clin. Microbiol.* **44**:2533–2540.
 74. Takano, T., W. Higuchi, T. Otsuka, T. Baranovich, S. Enany, K. Saito, H. Isobe, S. Dohmae, K. Ozaki, M. Takano, Y. Iwao, M. Shibuya, T. Okubo, S. Yabe, D. Shi, I. Reva, L. J. Teng, and T. Yamamoto. 2008. Novel characteristics of community-acquired methicillin-resistant *Staphylococcus aureus* strains belonging to multilocus sequence type 59 in Taiwan. *Antimicrob. Agents Chemother.* **52**:837–845.
 75. Takeuchi, F., S. Watanabe, T. Baba, H. Yuzawa, T. Ito, Y. Morimoto, M. Kuroda, L. Cui, M. Takahashi, A. Ankai, S. Baba, S. Fukui, J. C. Lee, and K. Hiramatsu. 2005. Whole-genome sequencing of *Staphylococcus haemolyticus* uncovers the extreme plasticity of its genome and the evolution of human-colonizing staphylococcal species. *J. Bacteriol.* **187**:7292–7308.
 76. Tyagi, S., D. P. Bratu, and F. R. Kramer. 1998. Multicolor molecular beacons for allele discrimination. *Nat. Biotechnol.* **16**:49–53.
 77. Tyagi, S., and F. R. Kramer. 1996. Molecular beacons: probes that fluoresce upon hybridization. *Nat. Biotechnol.* **14**:303–308.
 78. Vallone, P. M., and J. M. Butler. 2004. AutoDimer: a screening tool for primer-dimer and hairpin structures. *BioTechniques* **37**:226–231.
 79. Vandenesch, F., T. Naimi, M. C. Enright, G. Lina, G. R. Nimmo, H. Heffernan, N. Liassine, M. Bes, T. Greenland, M. E. Reverdy, and J. Etienne. 2003. Community-acquired methicillin-resistant *Staphylococcus aureus* carrying Panton-Valentine leukocidin genes: worldwide emergence. *Emerg. Infect. Dis.* **9**:978–984.
 80. Vet, J. A., and S. A. Marras. 2005. Design and optimization of molecular beacon real-time polymerase chain reaction assays. *Methods Mol. Biol.* **288**:273–290.
 81. Wielders, C. L., M. R. Vriens, S. Brisse, L. A. de Graaf-Miltenburg, A. Troelstra, A. Fleer, F. J. Schmitz, J. Verhoef, and A. C. Fluit. 2001. In-vivo transfer of *mecA* DNA to *Staphylococcus aureus*. *Lancet* **357**:1674–1675. (Corrected.)
 82. Wu, S. W., H. de Lencastre, and A. Tomasz. 2001. Recruitment of the *mecA* gene homologue of *Staphylococcus sciuri* into a resistance determinant and expression of the resistant phenotype in *Staphylococcus aureus*. *J. Bacteriol.* **183**:2417–2424.
 83. Zhang, K., J. A. McClure, S. Elsayed, and J. M. Conly. 2009. Novel staphylococcal cassette chromosome *mec* type, tentatively designated type VIII, harboring class A *mec* and type 4 *ccr* gene complexes in a Canadian epidemic strain of methicillin-resistant *Staphylococcus aureus*. *Antimicrob. Agents Chemother.* **53**:531–540.
 84. Zhang, K., J. A. McClure, S. Elsayed, T. Louie, and J. M. Conly. 2005. Novel multiplex PCR assay for characterization and concomitant subtyping of staphylococcal cassette chromosome *mec* types I to V in methicillin-resistant *Staphylococcus aureus*. *J. Clin. Microbiol.* **43**:5026–5033.