



Simple Screening for Carbapenemase-Producing *Enterobacteriaceae* by Moxalactam Susceptibility Testing

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The increase in carbapenemase producing *Enterobacteriaceae* (CPE) is a serious concern worldwide (1–7). However, not all CPE isolates show reduced susceptibility to carbapenems (6, 8–11). Some CPE isolates also produce other beta-lactamases, such as extended-spectrum and/or AmpC-type beta-lactamases (12–14). For these reasons, screening for CPE by antibiotic susceptibility testing is challenging. The specific phenotypic detection methods for CPE currently in use include the carbapenem inactivation method (CIM) (15), the Carba NP test (15, 16), and the Cica-beta test (17). The CIM is based on the disk diffusion method. The Carba NP and Cica-beta tests are able to identify some beta-lactamase classes by using specific inhibitors. However, specific inhibitors that work against OXA-48 group class D carbapenem-hydrolyzing beta-lactamases are not available (18, 19). A screening technique for CPE before a second confirmatory assay by CIM, Carba NP test, Cica-beta test, or genetic detection test by PCR would be useful. Here, we demonstrate the efficiency of a simple screening technique for CPE using moxalactam.

Nonduplicate isolates including CPE and non-CPE were identified and characterized at Toho University (Table 1). The types of beta-lactamase genes were confirmed by PCR amplification and DNA sequencing. All isolates were stored in a freezer at –80°C until use. Antibiotic susceptibility testing was performed by the Clinical and Laboratory Standards Institute-recommended microdilution method (M07-A10) (20). Customized frozen plates for microdilution testing were purchased from Eiken Chemical Co., Ltd. (Tokyo, Japan). The Clinical and Laboratory Standards Institute interpretative criteria in document M100-S25 (21) were applied. *Escherichia coli* ATCC 25922 and *Pseudomonas aeruginosa* ATCC 27853 were used as the quality control strains for antibiotic susceptibility testing.

The positive predictive values (PPVs) of CPE detection by using CLSI resistance criteria for imipenem, meropenem, ceftazidime, and moxalactam were 93.5, 96.3, 74.8, and 93.7%, respectively. The negative predictive values (NPVs) of CPE detection by using the nonsusceptibility criteria for imipenem, meropenem, ceftazidime, and moxalactam were 50.7, 50.0, 80.4, and 72.9%, respectively. The NPV increased from 72.9% to 81.5% when the criterion for moxalactam (≥ 16 mg/liter) was used, but the PPV decreased from 93.7% to 90.4% (Table 2). Five false-positive results were observed in

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TABLE 1 Antibiotic activities of imipenem, meropenem, ceftazidime, and moxalactam against members of the family *Enterobacteriaceae*

Enzyme(s) produced (no. of isolates)	No. of isolates of:								MIC (mg/liter)					
	<i>Escherichia coli</i>	<i>Klebsiella pneumoniae</i>	<i>Klebsiella oxytoca</i>	<i>Salmonella</i> sp.	<i>Enterobacter</i> sp.	<i>Citrobacter</i> sp.	<i>Proteus mirabilis</i>	<i>Morganella morganii</i>	Antibiotic	Range	MIC50	MIC90	%S/R ^a	
Carbapenemases														
IMP type (44)	0	0	1	0	43	0	0	0	0	0	0	0	0	81.8/6.8
														2
														0.5
														2
														77.3/4.5
														32 to >256
														128
														>256
														256
														>256
														0.25
														1
														4
														32
														64
														>256
														256
														>256
														0.0/100
														0.0/100
														0.0/90.9
														64
														16
														>256
														>256
														0.0/100
														>256
														0.0/100
														16.7/58.3
														8
														1
														8
														58.3/25
														16.7/66.7
														2
														32
														75/0.0
														16 to 64
														16 to 64
														4 to >256
														16 to >256
														Moxalactam
														16 to >256
														4
														16
														32
														63.6/27.3
														27.3/72.7
														256
														64
														>256
														8
														>256
														54.5/36.4
														16
														58.0/29.0
														1
														16
														60.0/26.0
														128
														8.0/89.0
														>256
														15.0/74.0
														128
														>256
														Moxalactam
Noncarbapenemases														
CTX-M type (57)	57	0	0	0	0	0	0	0	0	0	0	0	0	0
														Imipenem
														Meropenem
														Ceftazidime
														Moxalactam
														Imipenem
														Meropenem
														Ceftazidime
														Moxalactam
														Imipenem
														Meropenem
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														Moxalactam
														Imipenem
														Meropenem
														Ceftazidime
														Moxalactam

TABLE 2 Results of screening of carbapenemase-producing members of the family *Enterobacteriaceae* by interpretation criteria^a

Antibiotic	% PPV ^b	% NPV ^c
Imipenem	93.5 (29/31)	50.7 (73/144)
Meropenem	96.3 (26/27)	50.0 (74/148)
Ceftazidime	74.8 (89/119)	80.4 (45/56)
Moxalactam	93.7 (74/79)	72.9 (70/96)
Moxalactam (≥ 16 mg/liter)	90.4 (85/94)	81.5 (66/81)
Ceftazidime ^d	67.0 (61/91)	95.7 (45/47)

^aThe interpretation criteria used were those in reference 21, except for moxalactam (≥ 16 mg/liter).

^bThe values in parentheses are the number of carbapenem producers/number of resistant isolates.

^cThe values in parentheses are the number of non-carbapenem producers/number of susceptible and nonsusceptible isolates.

^dIMP-type enzyme producers, $n = 63$.

AmpC producers, and 26 false-negative results were observed in 12 KPC-type, 7 OXA-type, 6 IMP-type, and 1 GES-4-like enzyme-producing members of the family *Enterobacteriaceae*.

A limitation of this study is that we were unable to test a comprehensive range of CPE isolates because of a limited number of KPC-type, OXA-48, OXA-181, NDM-type, VIM-type, and VEB-type enzyme-producing CPE isolates. In Japan, the major carbapenemase is of the IMP type. Further testing to assess performance with chromosomal or acquired AmpC-producing *Enterobacteriaceae* isolates is in progress.

In conclusion, moxalactam at ≥ 16 mg/liter may be a useful, cheap, and simple primary screening method for detecting CPE in the clinical laboratory but requires follow-up confirmatory testing.

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We have no conflicts of interest to declare.

REFERENCES

- Abraham S, Wong HS, Turnidge J, Johnson JR, Trott DJ. 2014. Carbapenemase-producing bacteria in companion animals: a public health concern on the horizon. *J Antimicrob Chemother* 69:1155–1157. <https://doi.org/10.1093/jac/dkt518>.
- Falagas ME, Tansarli GS, Karageorgopoulos DE, Vardakas KZ. 2014. Deaths attributable to carbapenem-resistant *Enterobacteriaceae* infections. *Emerg Infect Dis* 20:1170–1175. <https://doi.org/10.3201/eid2007.121004>.
- Nordmann P. 2014. Carbapenemase-producing *Enterobacteriaceae*: overview of a major public health challenge. *Med Mal Infect* 44:51–56. <https://doi.org/10.1016/j.medmal.2013.11.007>.
- Savard P, Perl RM. 2014. Combating the spread of carbapenemases in *Enterobacteriaceae*: a battle that infection prevention should not lose. *Clin Microbiol Infect* 20:854–861. <https://doi.org/10.1111/1469-0691.12748>.
- Tängdén T, Giske CG. 2015. Global dissemination of extensively drug-resistant carbapenemase-producing *Enterobacteriaceae*: clinical perspectives on detection, treatment and infection control. *J Intern Med* 277:501–512. <https://doi.org/10.1111/joim.12342>.
- Voulgari E, Poulou A, Koumaki V, Tsakris A. 2013. Carbapenemase-producing *Enterobacteriaceae*: now that the storm is finally here, how will timely detection help us fight back? *Future Microbiol* 8:27–39. <https://doi.org/10.2217/fmb.12.130>.
- Woodford N, Wareham DW, Guerra B, Teale C. 2014. Carbapenemase-producing *Enterobacteriaceae* and non-*Enterobacteriaceae* from animals and the environment: an emerging public health risk of our own making? *J Antimicrob Chemother* 69:287–291. <https://doi.org/10.1093/jac/dkt392>.
- Birgy A, Bidet P, Genel N, Doit C, Decre D, Arlet G, Bingen E. 2012. Phenotypic screening of carbapenemases and associated beta-lactamases in

- carbapenem-resistant Enterobacteriaceae. *J Clin Microbiol* 50: 1295–1302. <https://doi.org/10.1128/JCM.06131-11>.
9. Cohen Stuart J, Leverstein-Van Hall MA, Dutch Working Party on the Detection of Highly Resistant Microorganisms. 2010. Guideline for phenotypic screening and confirmation of carbapenemases in Enterobacteriaceae. *Int J Antimicrob Agents* 36:205–210. <https://doi.org/10.1016/j.ijantimicag.2010.05.014>.
 10. Nordmann P, Poirel L. 2002. Emerging carbapenemases in Gram-negative aerobes. *Clin Microbiol Infect* 8:321–331. <https://doi.org/10.1046/j.1469-0691.2002.00401.x>.
 11. Pasteran F, Mendez T, Guerriero L, Rapoport M, Corso A. 2009. Sensitive screening tests for suspected class A carbapenemase production in species of Enterobacteriaceae. *J Clin Microbiol* 47:1631–1639. <https://doi.org/10.1128/JCM.00130-09>.
 12. Carvalho-Assef AP, Pereira PS, Albano RM, Beriao GC, Tavares CP, Chagas TP, Marques EA, Timm LN, Da Silva RC, Falci DR, Asensi MD. 2014. Detection of NDM-1-, CTX-M-15-, and qnrB4-producing *Enterobacter hormaechei* isolates in Brazil. *Antimicrob Agents Chemother* 58: 2475–2476. <https://doi.org/10.1128/AAC.02804-13>.
 13. Seiffert SN, Marschall J, Perreten V, Carattoli A, Furrer H, Endimiani A. 2014. Emergence of *Klebsiella pneumoniae* co-producing NDM-1, OXA-48, CTX-M-15, CMY-16, QnrA and ArmA in Switzerland. *Int J Antimicrob Agents* 44:260–262. <https://doi.org/10.1016/j.ijantimicag.2014.05.008>.
 14. Yong D, Toleman MA, Giske CG, Cho HS, Sundman K, Lee K, Walsh TR. 2009. Characterization of a new metallo-beta-lactamase gene, bla_{NDM-1}, and a novel erythromycin esterase gene carried on a unique genetic structure in *Klebsiella pneumoniae* sequence type 14 from India. *Antimicrob Agents Chemother* 53:5046–5054. <https://doi.org/10.1128/AAC.00774-09>.
 15. van der Zwaluw K, de Haan A, Pluister GN, Bootsma HJ, de Neeling AJ, Schouls LM. 2015. The carbapenem inactivation method (CIM), a simple and low-cost alternative for the Carba NP test to assess phenotypic carbapenemase activity in Gram-negative rods. *PLoS One* 10:e0123690. <https://doi.org/10.1371/journal.pone.0123690>.
 16. Poirel L, Nordmann P. 2015. Rapidec Carba NP test for rapid detection of carbapenemase producers. *J Clin Microbiol* 53:3003–3008. <https://doi.org/10.1128/JCM.00977-15>.
 17. Lavigne JP, Pfeiffer C, Vidal L, Sotto A. 2011. Rapid detection of multi-drug resistant Gram-negative bacilli by Cica-Beta-test strips. *Pathol Biol (Paris)* 59:e7–e11. <https://doi.org/10.1016/j.patbio.2010.08.004>.
 18. Harris PN, Tambyah PA, Paterson DL. 2015. Beta-lactam and beta-lactamase inhibitor combinations in the treatment of extended-spectrum beta-lactamase producing Enterobacteriaceae: time for a reappraisal in the era of few antibiotic options? *Lancet Infect Dis* 15:475–485. [https://doi.org/10.1016/S1473-3099\(14\)70950-8](https://doi.org/10.1016/S1473-3099(14)70950-8).
 19. Shlaes DM. 2013. New beta-lactam–beta-lactamase inhibitor combinations in clinical development. *Ann N Y Acad Sci* 1277:105–114. <https://doi.org/10.1111/nyas.12010>.
 20. Clinical and Laboratory Standards Institute. 2015. Performance standards for antimicrobial susceptibility testing; twenty-fifth informational supplement. M100-S25. CLSI, Wayne, PA.
 21. Clinical and Laboratory Standards Institute. 2015. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard tenth edition. M07-A10. CLSI, Wayne, PA.