



# Stability of SARS-CoV-2 in Phosphate-Buffered Saline for Molecular Detection

Garrett A. Perchetti,<sup>a</sup> Meei-Li Huang,<sup>a</sup> Vikas Peddu,<sup>a</sup> Keith R. Jerome,<sup>a,b</sup> Alexander L. Greninger<sup>a,b</sup>

<sup>a</sup>Department of Laboratory Medicine, Virology Division, University of Washington, Seattle, Washington, USA

<sup>b</sup>Vaccine and Infectious Disease Division, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA

**KEYWORDS** COVID-19, PBS, SARS-CoV-2, coronavirus, molecular detection, refrigeration, specimen stability, transport media

RNA viruses often require “cold chains” of transportation to prevent the breakdown of genetic material. The logistics of getting samples from the patient to a diagnostic laboratory, sometimes thousands of miles away from the original collection site, can be complex and resource intensive (1, 2). Nucleic acid degradation of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) RNA can compromise the accuracy of molecular detection methods. It has been demonstrated that nasopharyngeal specimens containing SARS-CoV-2 can be stored in phosphate-buffered saline (PBS) as a substitute for viral transport medium (VTM) for up to 7 days (3). Here, we evaluate the stability of differing viral loads of SARS-CoV-2 over 28 days stored at room temperature, 4°C, –20°C, or –80°C.

We used the first SARS-CoV-2-positive nasopharyngeal swab detected in our laboratory for spike-in material. PBS spiked with this SARS-CoV-2 specimen was stored in quadruplicates and divided into two concentrations, namely, 5,000 to 10,000 copies/ml (high titer) and 500 to 1,000 copies/ml (low titer), as determined by droplet digital reverse transcriptase PCR (RT-PCR). Nucleic acids were isolated on the Roche MagNA Pure96 system (Basel, Switzerland). Qualitative RT-PCR (qRT-PCR) was performed on 448 samples using our CDC-based laboratory-developed test, as described previously (4, 5).

For the high concentration of SARS-CoV-2, regardless of storage conditions, 100% of samples were detected by qRT-PCR through day 28. At room temperature, median cycle threshold ( $C_T$ ) values for lower titers for both N1 and N2 targets remained consistent through day 28, fluctuating less than 1 median  $C_T$  (Table 1). For lower concentrations of virus, storage at room temperature was associated with reductions of positivity beginning at day 7, and by day 28, 0% of samples were detected for N1. Storage at room temperature was the least stable of all environmental conditions tested, with 54.2% of negative PCR results.

At 4°C, there was minimal change in  $C_T$ s over time at the higher viral concentration. For lower titers,  $C_T$ s increased by 2.1  $C_T$ s for N1 and 2.6  $C_T$ s for N2 over the 28 days. At –20°C, lower titers of virus fluctuated slightly more, increasing by  $\geq 3$   $C_T$ s. Storage of SARS-CoV-2 in PBS at –20°C was the second least stable condition, accounting for 37.5% of negative PCR results. Storage at –80°C showed the greatest stability, with all samples detected throughout the 28 days and  $\leq 1.5$  median  $C_T$ s for both N1 and N2 targets.

Here, we show that the stability of SARS-CoV-2 can be maintained at 4°C for up to a month when –80°C storage is not available (6, 7). At viral loads of  $>5,000$  copies/ml—corresponding to  $>75\%$  of positive samples recovered in our clinical lab to

**Citation** Perchetti GA, Huang M-L, Peddu V, Jerome KR, Greninger AL. 2020. Stability of SARS-CoV-2 in phosphate-buffered saline for molecular detection. *J Clin Microbiol* 58:e01094-20. <https://doi.org/10.1128/JCM.01094-20>.

**Editor** Alexander J. McAdam, Boston Children's Hospital

**Copyright** © 2020 American Society for Microbiology. All Rights Reserved.

Address correspondence to Alexander L. Greninger, [agrening@uw.edu](mailto:agrening@uw.edu).

**Accepted manuscript posted online** 15 May 2020

**Published** 23 July 2020

**TABLE 1** Median  $C_T$  values for N1 and N2 targets<sup>a</sup>

Storage condition by target and titer	Median $C_T$ value						
	Day 0	Day 1	Day 2	Day 7	Day 14	Day 21	Day 28
<b>N1</b>							
5,000–10,000 copies/ml							
Room temp	31.4	31.0	31.5	31.2	31.6	31.4	31.3
4°C	30.8	30.9	31.3	31.5	31.7	31.5	31.4
–20°C	30.9	31.4	31.7	31.2	31.7	31.8	31.8
–80°C	30.7	31.1	31.4	31.0	31.0	30.9	31.8
500–1,000 copies/ml							
Room temp	33.4	36.8	36.3	36.1 (3/4) <sup>b</sup>	35.8 (3/4)	35.2 (1/4)	NDET (0/4)
4°C	34.4	34.9	35.1	35.9	35.9	35.8 (3/4)	36.5
–20°C	34.2	35.2	36.0	36.5	37.5 (1/4)	37.4	37.3 (3/4)
–80°C	35.1	34.4	35.3	34.9	36.2	36.4	35.3
<b>N2</b>							
5,000–10,000 copies/ml							
Room temp	30.6	30.4	30.9	30.9	31.1	31.1	31.2
4°C	30.4	30.2	30.5	30.6	31.3	30.6	31.1
–20°C	30.6	30.5	30.9	30.6	31.6	30.9	31.2
–80°C	30.6	30.5	30.6	30.6	30.6	30.5	31.2
500–1,000 copies/ml							
Room temp	32.8	35.0	35.8	36.6 (3/4)	37.0	38.3 (2/4)	38.3 (3/4)
4°C	33.8	34.2	34.7	34.9 (3/4)	36.0	35.4	36.4
–20°C	33.7	35.3	36.0	37.8 (3/4)	36.7	36.3 (3/4)	37.0 (3/4)
–80°C	34.4	33.7	34.8	35.7	35.8	34.7	35.9

<sup>a</sup> $C_T$ , cycle threshold; NDET, not detected. N1 and N2 are amplicons within the nucleocapsid gene of SARS-CoV-2.

<sup>b</sup>Parentheticals denote the number of samples that were detected at a condition only if the 4 replicates were not all detected.

date—different storage temperatures did not have a substantial impact on our ability to detect SARS-CoV-2 when stored in PBS.

## REFERENCES

- Nybo M, Cadamuro J, Cornes MP, Gómez Rioja R, Grankvist K. 2019. Sample transportation—an overview. *Diagnosis (Berl)* 26:39–43. <https://doi.org/10.1515/dx-2018-0051>.
- Opitz L, Salinas-Riester G, Grade M, Jung K, Jo P, Emons G, Ghadimi BM, Beissbarth T, Gaedcke J. 2010. Impact of RNA degradation on gene expression profiling. *BMC Med Genomics* 3:36. <https://doi.org/10.1186/1755-8794-3-36>.
- Rodino KG, Espy MJ, Buckwalter SP, Walchak RC, Germer JJ, Fernholz E, Boerger A, Schuetz AN, Yao JD, Binnicker MJ. 2020. Evaluation of saline, phosphate buffered saline and minimum essential medium as potential alternatives to viral transport media for SARS-CoV-2 testing. *J Clin Microbiol* <https://doi.org/10.1128/JCM.00590-20>.
- Lieberman JA, Pepper G, Naccache SN, Huang M-L, Jerome KR, Greninger AL. 2020. Comparison of commercially available and laboratory developed assays for in vitro detection of SARS-CoV-2 in clinical laboratories. *J Clin Microbiol* <https://doi.org/10.1128/JCM.00821-20>.
- Nalla AK, Casto AM, Huang M-L, Perchetti GA, Sampoleo R, Shrestha L, Wei Y, Zhu H, Jerome KR, Greninger AL. 2020. Comparative performance of SARS-CoV-2 detection assays using seven different primer/probe sets and one assay kit. *J Clin Microbiol* <https://doi.org/10.1128/JCM.00557-20>.
- Relova D, Rios L, Acevedo AM, Coronado L, Perera CL, Pérez LJ. 2018. Impact of RNA degradation on viral diagnosis: an understated but essential step for the successful establishment of a diagnosis network. *Vet Sci* 5:19. <https://doi.org/10.3390/vetsci5010019>.
- Ren S-Y, Wang W-B, Hao Y-G, Zhang H-R, Wang Z-C, Chen Y-L, Gao R-D. 2020. Stability and infectivity of coronaviruses in inanimate environments. *World J Clin Cases* 8:1391–1399. <https://doi.org/10.12998/wjcc.v8.i8.1391>.