
1 *Sustained Reduction of Microbial Burden on Common Hospital Surfaces Through The Introduction*
2 *of Copper*

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20 **ABSTRACT**

21 **Background:** The contribution of environmental surface contamination with pathogenic organisms to
22 the development of healthcare-associated infections (HAI) has not been well defined.

23 **Methods:** The microbial burden (MB) associated with commonly touched surfaces in intensive care
24 units (ICUs) was determined by sampling six objects in 16 rooms from ICUs in three hospitals over 43
25 months. At month 23, copper-alloy surfaces, with inherent antimicrobial properties, were installed onto
26 six monitored objects in 8 of 16 rooms and the effect that this application had on the intrinsic MB
27 present on the six objects was assessed. Census continued in rooms with and without copper for an
28 additional 21 months.

29 **Results:** In concert with routine infection control practices, the average MB found for the six objects
30 assessed in the clinical environment during the pre-intervention phase was 28 times higher (6,985
31 cfu/100 cm², n=3977 objects sampled) than levels proposed as benign immediately after terminal
32 cleaning, <250 cfu/100cm². During the intervention phase the MB was found to be significantly lower
33 for both the control and copper surfaced objects. Copper was found to exert a significant 83%
34 reduction to the average MB found on the objects (465 cfu/100cm², n=2714 objects) as compared to the
35 controls (2,674 cfu/100cm², n=2,831 objects, p<0.0001).

36 **Conclusion:** The introduction of copper surfaces to objects formerly covered with plastic, wood,
37 stainless steel and other materials found in the patient care environment significantly reduced the overall
38 MB on a continuous basis thereby providing a potentially safer environment for hospital patients, HCWs
39 and visitors.

40 INTRODUCTION

41 Despite best efforts promoting infection control protocols (28, 33), hospital-acquired infections
42 (HAI) remain a common complication of hospital care occurring at an estimated rate of two-million
43 annually in the US (30). At issue is the source of the microbes responsible for HAI. Much work has
44 focused on the transfer of microbes from patients to healthcare workers (HCWs) and vice versa, and it is
45 likely that commonly touched items serve as significant reservoirs for these microbes.

46 Microbes have an inherent ability to colonize any surface. Studies have shown that microbes can
47 persist for weeks on stainless steel surfaces and polymeric materials used to fabricate touch surfaces in
48 hospitals (22). MRSA may exist on surfaces for as long as 360 days (37, 38) and spore-forming
49 bacteria, including *Clostridium difficile* can survive for months. The longer a nosocomial pathogen
50 persists on a surface, the longer it may be a source for transmission to a susceptible patient or HCW (5,
51 13, 17, 21, 26, 29). Frequently touched surfaces such as doorknobs, push plates, bed rails, faucet
52 handles, and IV poles, have been identified as reservoirs for the spread of pathogenic microbes (3, 27)
53 which can easily contaminate hands and equipment of HCWs who, in turn, can transmit these pathogens
54 to patients during routine care.

55 A concentration of less than 250 aerobic colony-forming units (cfu) per 100 cm² of surface area
56 has been proposed as a standard for being considered benign immediately after terminal cleaning (11,
57 19). When the Microbial Burden (MB) exceeds this level transmission likely increases from the
58 surfaces to healthcare workers and/or patients. To date, while there are multiple protocols for hand-
59 hygiene and room cleaning, there are few strategies that can consistently minimize the MB found in the
60 environment. CDC guidelines for *Disinfection and Sterilization of Healthcare Facilities* (28), describe
61 reducing rates of HAI through appropriate use of disinfection and sterilization of the patient care

62 environment. These guidelines incorporate a disinfection strategy devised more than 40 years ago (34)
63 on the predicted degree of risk involved in the use of inanimate objects: *critical*, which includes items
64 that enter sterile tissue (surgical instruments); *semi-critical*, which includes items that come into contact
65 with mucous membranes or non-intact skin (endoscopes); and *non-critical*, which includes items that
66 only come in contact with skin. Environmental surfaces fall within the non-critical category (16, 31, 34).
67 Increasing evidence suggests that enhanced cleaning/disinfection of environmental surfaces can reduce
68 contamination of HCWs and thus reduce transmission of hospital pathogens (4). However, numerous
69 reports indicate that a high percentage of environmental surfaces are not terminally cleaned well (7-9).
70 When objects from hospital rooms were cultured, 94% of those from rooms housing VRE infected
71 patients and 100% from those housing *C. difficile* patients were widely contaminated with the organisms
72 (12).

73 *In vitro* (24, 25, 39) and *in vivo* (10, 15, 20) studies have established the effectiveness of metallic
74 copper surfaces as an antimicrobial material for its ability to reduce the concentration of bacteria on hard
75 surfaces. In this study we have expanded on these observations by characterizing the MB associated
76 with commonly touched objects surfaced with and without copper in the intensive care unit (ICU) in
77 order to understand the risk that the MB might represent and the benefit that a perpetually active copper
78 material might offer in continuously reducing the MB in the built hospital environment.

79 MATERIALS AND METHODS

80 **SETTING:** A multi-site study was conducted within the ICUs of three separate US hospitals. The study
81 was approved by the institutional review boards for all sites as well as by the Office of Risk Protection
82 of the United States Army, the sponsor of the work. The Medical University of South Carolina (hospital
83 1) located in Charleston, South Carolina is a 660-bed academic facility with 17 medical ICU beds.
84 Memorial Sloan Kettering Cancer Center (hospital 2) located in New York, New York is a 432-bed

85 cancer hospital with 20 medical-surgical ICU beds. The Ralph H. Johnson Veterans Administration
86 Medical Center (hospital 3), also in Charleston, is a 98-bed hospital with eight medical ICU beds.

87 **STUDY DESIGN:** The MB associated with six common, high touch objects with which patients, HCWs,
88 and visitors routinely interact between daily routine cleanings (Table 1) were measured weekly for 43
89 months using six rooms each from hospitals 1 & 2 and four rooms from hospital 3 for a total of 16
90 rooms. Commencing during month 23 (intervention phase) the six objects associated with one-half of
91 the study rooms were surfaced with a continuously active antimicrobial material, metallic copper, in
92 order to determine the effect on the MB.

93 **FABRICATION OF ITEMS SURFACED WITH COPPER ALLOYS:** The objects surfaced with copper were
94 fabricated using copper alloys registered with the U.S. EPA for their inherent ability to kill bacteria
95 (36). Four items were common to all hospitals; the side rails of the patient bed, the over-bed tray table,
96 the IV pole, and the contact surface of the arm rests of the visitor's chair (Figure 1). Two other high
97 touch objects were also sampled from each site (Table 1). For additional details on fabrication please
98 see the supplemental material.

99 **ENVIRONMENTAL CLEANING REGIMES:** Each of the study sites followed routine standards of
100 environmental cleaning and disinfection as prescribed by their respective infection control programs.
101 This required that all objects and surfaces be cleaned at least once each day using a prescribed hospital
102 grade disinfectant and upon patient discharge. Three US-EPA registered disinfectants were used during
103 the intervention. Virex256[®] was used for routine and terminal cleaning. Dispatch[®] was used to clean
104 rooms with a confirmed case of *Clostridium difficile* and Cavicide[®] was used for spot cleaning.
105 Additionally, during the pre-intervention phase one site, Hospital 2, used the disinfectant
106 ElimstaphNo2[®] (Walter G. Legge Company, Inc., Peekskill, NY) rather than Virex256[®] for its routine

107 and terminal cleaning. The products were all used according to the label instructions and were
108 consistently applied.”

109 **SAMPLE COLLECTION PROCEDURE:** Surfaces were sampled once each week at approximately 9 AM,
110 excluding weeks with US federal holidays using either a 10 cm x 10 cm or 4 cm x 25 cm sterile template
111 placed over each surface. The exposed area was vigorously wiped using uniform pressure and motion,
112 five strokes horizontally and vertically for a total of ten strokes in each direction. Samples were
113 transported to Medical University of South Carolina and processed as previously described (1). MB was
114 reported as colony forming units (cfu) per 100cm². For greater detail see supplemental materials and
115 methods.

116 **CALCULATIONS AND STATISTICAL ANALYSIS:** The average MB of each item was calculated and the
117 MB of each room was determined as the sum of the MB of the six objects within that room. The
118 Kruskal-Wallis Test was used to compare the average MB associated with objects and rooms (EpiInfo,
119 CDC, Atlanta GA) between pre-intervention and intervention phases as well between copper surfaced
120 rooms and control-surfaced rooms. A p-value of ≤ 0.05 was considered statistically significant. The
121 antimicrobial efficacy of copper was calculated as the difference in average MB between copper and
122 non-copper objects and rooms and was expressed as the percentage with which copper reduced the MB.

123 **RESULTS**

124 **INTRINSIC MICROBIAL BURDEN FOUND ON COMMON HIGH TOUCH OBJECTS**

125 Over the 43 months of the study, samples were recovered from 9,522 objects in 1,587 rooms
126 across three study sites. The average MB found for the six objects assessed in the clinical environment
127 during the pre-intervention phase was 28 times higher (6,985 cfu/100 cm², n=3977 objects sampled)
128 than levels commonly accepted as benign, <250 cfu/100cm² (Figure 2) (11, 18, 19, 23, 40). This value

129 was exceeded for each object sampled. Bed rails were the most heavily burdened of the objects
130 averaging a concentration 69 times greater than the level proposed as benign immediately after terminal
131 cleaning or 17,336 cfu/100cm² with a standard error of sampling of $\pm 2,896$ cfu/100cm² (For additional
132 detail please see Supplemental, Table S1). The majority of microorganisms (64%) were staphylococci
133 where approximately 90% of the population recovered was coagulase-negative.

134 MRSA and VRE were also frequently recovered from objects. Bed rails had an average
135 concentration of 151 cfu of MRSA/100cm² and of 667 VRE/100cm² and nurses call buttons had
136 averages of 146 cfu of MRSA/100cm² and 16 cfu of VRE/100cm² (Figure 2). The average
137 concentration of gram-negative bacteria on bed rails and call buttons was 57 and 109 cfu/100cm²
138 respectively and the average concentration of gram-negative bacteria resident on the monitor and tray
139 table was higher at 5,914 and 8,572 cfu/100cm² respectively, reflecting a small number of outliers from
140 samples collected from hospital 2.

141 **COPPER LOWERED THE MB FOUND ON COMMON HIGH TOUCH OBJECTS**

142 During the 21 month intervention the antimicrobial effect exerted by metallic copper surfaces
143 was immediate and consistently evident. A significant, 83% reduction to the average MB recovered
144 from the copper surfaced objects was seen. Collectively, the average MB likely to be encountered from
145 one of the six copper surfaced objects was 465 cfu/100cm², n=2714 objects, while the average burden
146 recovered from the control items was 2,674 cfu/100cm², n=2,831 objects (p<0.0001). The summative
147 average for the six objects surfaced in copper was also approximately 83% lower, than the burden
148 recovered from the control objects (2,521 vs. 14,813 cfu/100cm² (Supplemental, Table S2)). When
149 considered individually, 5 of the 6 objects surfaced in copper also saw significantly lower burdens
150 (Figure 3, Panel A, Supplemental Table S2). Polypropylene bed rails were again the most heavily
151 burdened of the control objects sampled with an average MB of 6,456 cfu/100cm². In contrast, the MB

152 recovered from copper bed rails was 94% lower (366 cfu/100cm²) and this difference was significant
153 (p<0.0001). Finally, the antimicrobial activity of copper was found to be universal in its ability to kill
154 many types of microbes (Figure 3, Panels B & C).

155 Copper surfaces were also found to attenuate the inherent variability associated with the MB
156 resident on surfaces within the patient care areas. Eighty-three percent of samples recovered from
157 copper bed rails were found below 250 cfu/100cm² whereas only 20% of the samples recovered from the
158 plastic rails were found below this level (Figure 4) suggesting that copper might limit the heterogeneity
159 of risk to the patient attributed to the variation to MB by limiting the range of MB resident on commonly
160 touched surfaces. The dynamic nature of the MB resident on the objects sampled, attributed to
161 stochastic processes, was evident throughout the trial (Supplemental Figures S1 & S2). A comparison
162 of the summative MB from the pre-intervention and intervention phases found that the MB was 64.4%
163 lower during the intervention period of the trial (41,586 cfu/100 cm², n=668 rooms, vs. 16,188 cfu/100
164 cm², n=511 rooms; p<0.0001).

165 **EFFECT OF COPPER INTERVENTION ON ANTIBIOTIC-RESISTANT BACTERIA**

166 When considering the frequency with which MRSA and/or VRE were encountered over the
167 study period, 169 (2.4%) of 7,005 control objects were found to harbor MRSA, while 239 (3.4%) were
168 found to harbor VRE. During the intervention phase, MRSA and VRE were recovered with greater
169 frequency from objects in rooms without copper surfaces. MRSA was recovered eight times (0.3%;
170 n=2,781 copper objects) compared to 19 times (0.63%; n=3,004; p=0.0804, control) while VRE was
171 recovered 9 times from rooms with copper surfaces (0.3%) compared to 91 times (3%) from control
172 rooms (p<0.0001). On a per sample basis, copper surfaces were approximately six-times less likely to
173 harbor one of these organisms. Based on the summative MB measured for each of the surfaces sampled
174 over the intervention period, the combined MRSA and VRE burdens were 96.8% lower on copper

175 surfaces than on comparable plastic, wood, metal and painted surfaces and were 98.8%, less on the bed
176 rails, the most heavily burdened object.

177 **DISCUSSION**

178 This study suggests that six common high-touch objects with which HCWs, patients and visitors
179 routinely interact carry a substantial MB, and thus present a risk to patients. These data underscore the
180 need to insure that cleaning is completed in an effective manner as bacterial concentrations resident on
181 items sampled were well above values recommended immediately after terminal cleaning (11, 18, 19,
182 23, 40). Concentrations of bacteria on objects varied substantially (Figure 4). The stochastic behavior of
183 the MB distributed across the three ICUs is likely attributed to the inherent dynamics of patient care,
184 cleaning, patient characteristics as well as other unknown factors.

185 Incorporation of inherently and continuously active antimicrobial copper onto high-touch
186 surfaces in the ICU offered an enhanced effect in combination with regular cleaning and infection
187 control practices resulting in significantly lower MB and potentially safer surfaces. Bed rails were the
188 most heavily burdened control objects with a maximum MB of 306,000 cfu/100 cm², seventeen times
189 higher than the maximum value observed from a copper surfaced rail. In fact 80% of plastic bed rails
190 had bacterial concentrations above the risk threshold for transferring infectious bacteria (Figure 4). In
191 contrast, 83% of copper bed rails had levels below this threshold. Thus, generally, during the conduct of
192 patient care, objects surfaced with copper carried concentrations of bacteria at or below the threshold
193 recommended immediately after terminal cleaning (11, 18, 19, 23, 40). The antimicrobial activity of
194 the metallic copper surfaces was equivalent throughout the course of the trial. This was evident from
195 the observation that over the course of sampling, 46% of copper bedrails had no recoverable bacteria
196 (Figure 4). In contrast only 3% of bed rails sampled from control rooms failed to yield viable bacteria.

197 Similarly, the five other copper items had remarkably lower burdens. The call button was the
198 most heavily burdened of the copper-surfaced objects evaluated, however 71% of the samples were
199 below the proposed terminal cleaning threshold. Seventy-five percent of chair arms sampled; 90% of
200 tray tables; 91% of IV poles and 90% of data input devices were below the proposed standard of <250
201 cfu/100cm². In total, 45% of control objects from the 511 rooms sampled exceeded an average MB
202 considered to represent a risk to patients, compared to just 16% of copper-clad objects. The most
203 surprising finding was the 64% decrease in MB between the pre-intervention and intervention phases in
204 the control rooms. This might be accounted for as a consequence of a number of independent and
205 uncontrolled variables: First, the presence of copper on the unit may have resulted in better cleaning by
206 the environmental services staff; 2) the presence of copper may have resulted in an antimicrobial halo
207 limiting the transfer of microbes between control rooms as staff were common to both rooms; 3) or
208 variations in compliance with other infection control measures such as hand hygiene might account for
209 the differences seen.

210 Unlike programs for better compliance with infection control such as hand-hygiene or barrier
211 precautions, the antimicrobial activity of copper-surfaced objects was not dependent on additional
212 training or supervision. It did not require alterations to existing cleaning practices or add to the annual
213 environmental cleaning costs, as does the application of ultraviolet light and/or hydrogen peroxide vapor
214 deposition for reduction in MB. Additionally reductions to the MB manifested by the copper objects
215 during active patient care approached the reduction level of 99.9% observed in tests conducted for
216 registration of copper-based surfaces with the U.S. EPA.

217 Recent literature provides increased evidence that contaminated hospital surfaces may be a
218 source of transmission of pathogens. Kramer reported that, in hospitals, surfaces with hand contact are
219 often contaminated with nosocomial pathogens and may serve as vectors for cross transmission (17).

220 Steifeland found that in patient rooms with MRSA carriers, HCWs are just as likely to contaminate their
221 hands or gloves from commonly-touched environmental surfaces as from direct contact with colonized
222 patients (35). Boyce and colleagues (6) demonstrated that nurses frequently acquired MRSA on their
223 gloves after touching surfaces near colonized patients and a report by Bhalla and others found that 53%
224 of hand-imprint cultures were positive for one or more pathogens after contact with surfaces near
225 hospitalized patients (2). Other studies have found that patients treated in rooms previously occupied by
226 individuals with colonization or infection with MRSA, VRE, and *C. difficile* are at a higher risk of
227 acquiring the organism than patients admitted to rooms where the previous occupant did not have
228 colonization or infection (14, 32).

229 The use of copper to control or reduce the MB on surfaces in healthcare has been previously
230 reported (10, 15, 20). In a South African community healthcare facility, copper surfaces (desks,
231 trolleys) were associated with a 71% reduction in MB compared to control surfaces when sampled every
232 6 weeks for a period of 6 months (20). A recent crossover study in a 19 bed acute medical ward found
233 that many copper surfaces were associated with significantly decreased MB compared to control
234 surfaces when sampled weekly for 24 weeks with reductions ranging from -0.4 cfu/cm² to -80.3 cfu/cm²
235 (15). Also, like our study, copper surfaces were significantly less likely to be contaminated with
236 indicator organisms such as VRE and coliforms. Our study differs from these in several respects.
237 Sampling in our study was performed over a substantially longer period of time (21 months), the objects
238 surfaced with copper were often medical devices within close proximity to the patient and used routinely
239 during direct patient care. Additionally, the populations cared for in the rooms involved in our study
240 were critically ill and generally not ambulatory which reduced the influence of their interactions with
241 other environmental surfaces within and outside the room.

242 Reducing the overall MB on a continuous basis, as evidenced in this study and others with the
243 introduction of continuously active antimicrobial copper surfaces, may provide a safer environment for
244 hospital patients, HCWs and visitors.

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250 REFERENCES

251

- 252 1. **Attaway, H. H., S. Fairey, L. L. Steed, C. D. Salgado, H. T. Michels, and M. G.**
253 **Schmidt.** Intrinsic bacterial burden associated with intensive care unit hospital beds: Effects of
254 disinfection on population recovery and mitigation of potential infection risk. *American Journal*
255 *of Infection Control.* (<http://dx.doi.org/10.1016/j.ajic.2011.11.019>)
- 256 2. **Bhalla, A., N. J. Pultz, D. M. Gries, A. J. Ray, E. C. Eckstein, D. C. Aron, and C. J.**
257 **Donskey.** 2004. Acquisition of nosocomial pathogens on hands after contact with environmental
258 surfaces near hospitalized patients. *Infect Control Hosp Epidemiol* **25**:164-7.
- 259 3. **Blythe, D., D. Keenlyside, S. J. Dawson, and A. Galloway.** 1998. Environmental
260 contamination due to methicillin-resistant *Staphylococcus aureus* (MRSA). *J. Hosp. Infect.*
261 **38**:67-69.
- 262 4. **Boyce, J. M.** 2007. Environmental contamination makes an important contribution to hospital
263 infection. *Journal of Hospital Infection* **65**:50-54.
- 264 5. **Boyce, J. M., N. L. Havill, J. A. Otter, and N. M. Adams.** 2007. Widespread environmental
265 contamination associated with patients with diarrhea and methicillin-resistant *Staphylococcus*
266 *aureus* colonization of the gastrointestinal tract. *Infect Control Hosp Epidemiol* **28**:1142-7.
- 267 6. **Boyce, J. M., G. Potter-Bynoe, C. Chenevert, and T. King.** 1997. Environmental
268 contamination due to methicillin-resistant *Staphylococcus aureus*: possible infection control
269 implications. *Infect Control Hosp Epidemiol* **18**:622-7.
- 270 7. **Carling, P. C., M. F. Parry, and S. M. Von Beheren.** 2008. Identifying opportunities to
271 enhance environmental cleaning in 23 acute care hospitals. *Infect Control Hosp Epidemiol* **29**:1-
272 7.
- 273 8. **Carling, P. C., M. M. Parry, M. E. Rupp, J. L. Po, B. Dick, and S. Von Beheren.** 2008.
274 Improving cleaning of the environment surrounding patients in 36 acute care hospitals. *Infect*
275 *Control Hosp Epidemiol* **29**:1035-41.
- 276 9. **Carling, P. C., S. Von Beheren, P. Kim, and C. Woods.** 2008. Intensive care unit
277 environmental cleaning: an evaluation in sixteen hospitals using a novel assessment tool. *J Hosp*
278 *Infect* **68**:39-44.
- 279 10. **Casey, A. L., D. Adams, T. J. Karpanen, P. A. Lambert, B. D. Cookson, P. Nightingale, L.**
280 **Miruszenko, R. Shillam, P. Christian, and T. S. Elliott.** 2010. Role of copper in reducing
281 hospital environment contamination. *J Hosp Infect* **74**:72-7.
- 282 11. **Dancer, S. J.** 2004. How do we assess hospital cleaning? A proposal for microbiological
283 standards for surface hygiene in hospitals. *J Hosp Infect* **56**:10-5.
- 284 12. **Eckstein, B. C., D. A. Adams, E. C. Eckstein, A. Rao, A. K. Sethi, G. K. Yadavalli, and C. J.**
285 **Donskey.** 2007. Reduction of *Clostridium Difficile* and vancomycin-resistant *Enterococcus*
286 contamination of environmental surfaces after an intervention to improve cleaning methods.
287 *BMC Infect Dis* **7**:61.
- 288 13. **Falk, P. S., J. Winnike, C. Woodmansee, M. Desai, and C. G. Mayhall.** 2000. Outbreak of
289 vancomycin-resistant enterococci in a burn unit. *Infect Control Hosp Epidemiol* **21**:575-82.
- 290 14. **Huang, S. S., and R. Platt.** 2003. Risk of methicillin-resistant *Staphylococcus aureus* infection
291 after previous infection or colonization. *Clin Infect Dis* **36**:281-5.
-

- 292 15. **Karpanen, T. J., A. L. Casey, P. A. Lambert, B. D. Cookson, P. Nightingale, L.**
293 **Miruszenko, and T. S. J. Elliott.** 2012. The Antimicrobial Efficacy of Copper Alloy Furnishing
294 in the Clinical Environment: A Crossover Study. *Infection Control and Hospital Epidemiology*
295 **33:3-9.**
- 296 16. **Kohn, W. G., A. S. Collins, J. L. Cleveland, J. A. Harte, K. J. Eklund, and D. M. Malvitz.**
297 2003. Guidelines for infection control in dental health-care settings--2003. *MMWR Recomm*
298 *Rep* **52:1-61.**
- 299 17. **Kramer, A., I. Schwebke, and G. Kampf.** 2006. How long do nosocomial pathogens persist on
300 inanimate surfaces? A systematic review. *BMC Infect Dis* **6:130.**
- 301 18. **Lewis, T., C. Griffith, M. Gallo, and M. Weinbren.** 2008. A modified ATP benchmark for
302 evaluating the cleaning of some hospital environmental surfaces. *J Hosp Infect* **69:156-63.**
- 303 19. **Malik, R. E., R. A. Cooper, and C. J. Griffith.** 2003. Use of audit tools to evaluate the efficacy
304 of cleaning systems in hospitals. *Am J Infect Control* **31:181-7.**
- 305 20. **Marais, F., S. Mehtar, and L. Chalkley.** 2010. Antimicrobial efficacy of copper touch surfaces
306 in reducing environmental bioburden in a South African community healthcare facility. *J Hosp*
307 *Infect* **74:80-2.**
- 308 21. **Martinez, J. A., R. Ruthazer, K. Hansjosten, L. Barefoot, and D. R. Snyderman.** 2003. Role of
309 environmental contamination as a risk factor for acquisition of vancomycin-resistant enterococci
310 in patients treated in a medical intensive care unit. *Arch Intern Med* **163:1905-12.**
- 311 22. **Michels, H. T., S. A. Wilks, J. O. Noyce, and C. W. Keevil.** 2005. Presented at the Materials
312 Science and Technology Conference, Copper for the 21st Century Symposium, Pittsburgh, PA,
313 September 25-28, 2005.
- 314 23. **Mulvey, D., P. Redding, C. Robertson, C. Woodall, P. Kingsmore, D. Bedwell, and S. J.**
315 **Dancer.** 2011. Finding a benchmark for monitoring hospital cleanliness. *J Hosp Infect* **77:25-30.**
- 316 24. **Noyce, J. O., H. Michels, and C. W. Keevil.** 2006. Potential use of copper surfaces to reduce
317 survival of epidemic methicillin-resistant *Staphylococcus aureus* in the healthcare environment. *J*
318 *Hosp Infect* **63:289-97.**
- 319 25. **Noyce, J. O., H. Michels, and C. W. Keevil.** 2006. Use of copper cast alloys to control
320 *Escherichia coli* O157 cross-contamination during food processing. *Appl Environ Microbiol*
321 **72:4239-44.**
- 322 26. **O'Doherty, A. J., P. G. Murphy, and R. A. Curran.** 1989. Risk of *Staphylococcus aureus*
323 transmission during ultrasound investigation. *J Ultrasound Med* **8:619-20.**
- 324 27. **Oie, S., I. Hosokawa, and A. Kamiya.** 2002. Contamination of room door handles by
325 methicillin-sensitive/methicillin-resistant *Staphylococcus aureus*. *J. Hosp. Infect.* **51:140-143.**
- 326 28. **Rutala, W. A., D. J. Weber, and H. I. C. P. A. C. (HICPAC).** 2008. Guideline for Disinfection
327 and Sterilization in Healthcare Facilities, 2008, Healthcare Infection Control Practices Advisory
328 Committee (HICPAC), vol. _____
329 http://www.cdc.gov/hicpac/Disinfection_Sterilization/acknowledg.html. United States of
330 America.
- 331 29. **Schabrun, S., and L. Chipchase.** 2006. Healthcare equipment as a source of nosocomial
332 infection: a systematic review. *J Hosp Infect* **63:239-45.**

- 333 30. **Scott, R. D.** 2009. The Direct Medical Costs of Healthcare-Associated Infections in U.S.
334 Hospitals and the Benefits of Prevention. PDF.
- 335 31. **Sehulster, L., and R. Y. Chinn.** 2003. Guidelines for environmental infection control in health-
336 care facilities. Recommendations of CDC and the Healthcare Infection Control Practices
337 Advisory Committee (HICPAC). *MMWR Recomm Rep* **52**:1-42.
- 338 32. **Shaughnessy, M. K., R. L. Micielli, D. D. DePestel, J. Arndt, C. L. Strachan, K. B. Welch,**
339 **and C. E. Chenoweth.** 2011. Evaluation of hospital room assignment and acquisition of
340 *Clostridium difficile* infection. *Infect Control Hosp Epidemiol* **32**:201-6.
- 341 33. **Siegel, J. D., E. Rhinehart, M. Jackson, and L. Chiarello.** 2007. 2007 Guideline for Isolation
342 Precautions: Preventing Transmission of Infectious Agents in Health Care Settings. *Am J Infect*
343 *Control* **35**:S65-164.
- 344 34. **Spaulding, E. H.** 1968. Chemical disinfection of medical and surgical materials., p. 517-531. *In*
345 C. Lawrence and S. S. Block (ed.), *Disinfection, sterilization, and preservation*. Lea & Febiger,
346 Philadelphia.
- 347 35. **Stiefel, U., J. L. Cadnum, B. C. Eckstein, D. M. Guerrero, M. A. Tima, and C. J. Donskey.**
348 2011. Contamination of hands with methicillin-resistant *Staphylococcus aureus* after contact
349 with environmental surfaces and after contact with the skin of colonized patients. *Infect Control*
350 *Hosp Epidemiol* **32**:185-7.
- 351 36. **United States Environmental Protection Agency.** 2008. EPA registers copper-containing alloy
352 products. <http://www.epa.gov/opp00001/factsheets/copper-alloy-products.htm>.
- 353 37. **Wagenvoort, J. H., and R. J. Penders.** 1997. Long-term in-vitro survival of an epidemic
354 MRSA phage-group III-29 strain. *J Hosp Infect* **35**:322-5.
- 355 38. **Wagenvoort, J. H., W. Sluijsmans, and R. J. Penders.** 2000. Better environmental survival of
356 outbreak vs. sporadic MRSA isolates. *J Hosp Infect* **45**:231-4.
- 357 39. **Weaver, L., H. T. Michels, and C. W. Keevil.** 2008. Survival of *Clostridium difficile* on copper
358 and steel: futuristic options for hospital hygiene. *J Hosp Infect* **68**:145-51.
- 359 40. **White, L. F., S. J. Dancer, C. Robertson, and J. McDonald.** 2008. Are hygiene standards
360 useful in assessing infection risk? *Am J Infect Control* **36**:381-4.
- 361
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Table 1
Antimicrobial Copper Alloys Used to Surface or Fabricate High Touched Items

	Copper Alloy	Component Fabricated	Description	% Copper Content
IV stands	C710 and C706	Pole(s)	Copper Nickel	80% / 90%
	C693	IV hanger loops	Brass	75%
	C87610	Base	Silicon bronze	90%
	C706	Handle	Copper nickel	90%
	C706	Brackets	Copper nickel	90%
Patient bed—Side rails	C110	Top of rails	Copper	99.99%
Over-bed table	C706	Table top	Copper nickel	90%
	C110	Table bottom	Copper	99.9%
	C464	Release lever	Naval brass	60%
Visitors' chair (arms)	C706	Arm rests	Copper nickel	90%
Nurse call button	C638	Buttons	Aluminum bronze	80%
	C260	Clamshell (Hospitals 1&3)	Cartridge brass	80%
Computer Mouse	C260	Computer mouse (Hospital 2 Only)	Cartridge brass	70%
Data Input devices	C524	Base of Monitor bezel (Hospitals 1 & 2)	Phosphor- Bronze	90%
	C710	Laptop palm rest (Hospital 3 Only)	Copper nickel	80%

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368 **Figure Legends**369 **Figure 1. Representative high touch objects and their respective placement in the ICU.**

370 **FIGURE 2. Assessment of the Inherent Microbial Burden Associated with High Touch**
371 **Objects.** The average concentration of bacteria, described by type was
372 determined from samples collected from six inanimate objects for a period of 23
373 months (N=668 rooms). Bed rails, Dark blue bars; Call buttons (Hospitals 1 & 3
374 and Mice (Hospital 2), Red Bars; Arms of the Chair, Yellow Bar; Tray Table, Light
375 Blue Bar; Data Input Device (Base of Monitor Bezel Hospitals 1 & 2 and Palm rest
376 of laptop computer, Hospital 3), Purple; and IV Pole Grey Bars, (the call button
377 represents values obtained from call buttons at 2 sites and a computer mouse from
378 the third due to the absence of a call button

379 **Figure 3. COPPER LOWERED THE MB FOUND ON COMMON HIGH TOUCH OBJECTS. Panel A.**
380 Comparison of the average MB between rooms with (Green Bars, N=501 rooms)
381 and without (Red Bars, N=511 rooms) copper surfaced items. Samples were
382 collected period of 21 months, processed and statistically analyzed as described in
383 the methods (*denotes a p value <0.05). **Panels B (Non-Copper Objects) and C,**
384 **(Copper Objects)** described the average concentration of bacteria, by type,
385 recovered from Bed rails, (Dark blue bars); Call buttons (Hospitals 1&3 and Mice
386 (Hospital 2), (Red Bars); Arms of the Chair, (Yellow Bars); Tray Table, (Light Blue
387 Bar); Data Input Device (Base of Monitor Bezel Hospitals 1 & 2 and Palm rest of
388 laptop computer, Hospital 3), Purple Bar); and IV Pole (Grey Bars), (the call button
389 represents values obtained from call buttons at 2 sites and a computer mouse from
390 the third due to the absence of a call button at that location).
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392 **Figure 4. Frequency distribution of the MB resident on all objects, by type, during the**
393 **intervention.** The MB observed for each sample from rooms with copper surfaced
394 objects are described on the left, those without copper surfaced objects are
395 described on right. The concentration of bacteria observed for each sample was
396 placed into one of three categories, 0 cfu/100cm² (green bar), 1-250 cfu/100cm²
397 (yellow bar), >250 cfu/100cm² (red bar) the final percentage, rounded to the
398 nearest whole number, of each category was determined.











